

REVISED FINAL

PHASE II RCRA FACILITY INVESTIGATION
REPORT FOR 16 SOLID WASTE MANAGEMENT UNITS

AT

FORT STEWART, GEORGIA
VOLUME I OF III

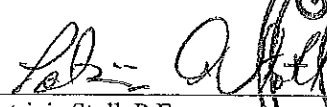
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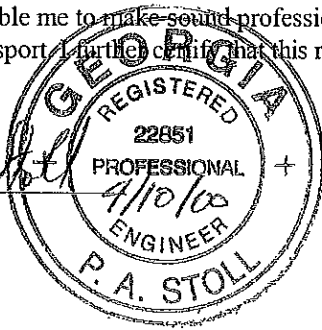
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The undersigned certifies that I am a qualified groundwater scientist who has received a baccalaureate or postgraduate degree in the natural sciences or engineering and have sufficient training and experience in groundwater hydrology and related fields, as demonstrated by state registration and completion of accredited university courses, to enable me to make sound professional judgments regarding groundwater monitoring and contaminant fate and transport. I further certify that this report was prepared by myself or a subordinate working under my direction.


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CONTENTS

VOLUME I

| | |
|---|-------|
| TABLES..... | xxii |
| FIGURES | xlili |
| ACRONYMS AND ABBREVIATIONS..... | li |
| EXECUTIVE SUMMARY | ES-1 |
| 1.0 INTRODUCTION..... | 1-1 |
| 1.1 OBJECTIVES AND SCOPE OF THE INVESTIGATION..... | 1-1 |
| 1.2 REPORT ORGANIZATION | 1-2 |
| 2.0 SITE HISTORY AND CONTAMINANTS | 2-1 |
| 2.1 INSTALLATION DESCRIPTION | 2-1 |
| 2.2 SITE LOCATION AND HISTORY | 2-1 |
| 2.3 PREVIOUS INVESTIGATIONS..... | 2-9 |
| 2.4 PRELIMINARY CONCEPTUAL SITE MODEL | 2-9 |
| 2.4.1 Camp Oliver Landfill (SWMU 2) and TAC-X Landfill (SWMU 3)..... | 2-9 |
| 2.4.2 Inactive EOD Areas (SWMUs 9, 10, and 11) | 2-9 |
| 2.4.3 Active EOD Area (SWMU 12A)..... | 2-9 |
| 2.4.4 Old Fire Training Area (SWMU 14)..... | 2-10 |
| 2.4.5 DRMO Hazardous Waste Storage Area (SWMU 17)..... | 2-10 |
| 2.4.6 Industrial Wastewater Treatment Plant (SWMU 18) | 2-10 |
| 2.4.7 Old Sludge Drying Beds (SWMU 19) | 2-11 |
| 2.4.8 Old Radiator Shop/Paint Booth (SWMU 24B)..... | 2-11 |
| 2.4.9 Thirty-two OWSs at 29 Sites Associated with Motorpools (SWMUs 27A through 27V)..... | 2-11 |
| 2.4.10 Evans Heliport POL Storage Facility (SWMU 29), DEH Asphalt Tanks (SWMU 31), and Supply Diesel Tank (SWMU 32)..... | 2-12 |
| 2.4.11 DEH Equipment Wash Rack (SWMU 34)..... | 2-12 |
| 2.4.12 NGTC Equalization Basin (SWMU 37) | 2-12 |
| 3.0 PHYSICAL CHARACTERISTICS OF THE SITE | 3-1 |
| 3.1 DEMOGRAPHICS | 3-1 |
| 3.2 TOPOGRAPHY | 3-1 |
| 3.3 SURFACE DRAINAGE..... | 3-1 |
| 3.4 REGIONAL GEOLOGY | 3-2 |
| 3.5 SOILS..... | 3-2 |
| 3.6 HYDROGEOLOGY | 3-4 |
| 3.7 ECOLOGY..... | 3-4 |
| 3.8 METEOROLOGY | 3-5 |
| 4.0 SUMMARY OF INVESTIGATION ACTIVITIES | 4-1 |
| 4.1 SAMPLING METHODOLOGIES..... | 4-1 |
| 4.1.1 Soil Sampling..... | 4-1 |
| 4.1.2 Groundwater Sampling | 4-8 |
| 4.1.3 Surface Water and Sediment Sampling..... | 4-11 |

CONTENTS (continued)

| | | |
|-------|--|------|
| 4.1.4 | Wastewater and Sediment (Sludge) Sampling | 4-11 |
| 4.1.5 | Investigation-derived Waste Management | 4-11 |
| 4.2 | DATA QUALITY ASSESSMENT..... | 4-11 |
| 5.0 | CONTAMINANT NATURE AND EXTENT | 5-1 |
| 5.1 | BACKGROUND DATA ANALYSIS | 5-1 |
| 5.2 | SURFACE SOIL | 5-1 |
| 5.3 | SUBSURFACE SOIL | 5-3 |
| 5.4 | GROUNDWATER..... | 5-3 |
| 5.5 | SURFACE WATER..... | 5-4 |
| 5.6 | SEDIMENT..... | 5-4 |
| 5.7 | SITE-RELATED CONTAMINANTS | 5-4 |
| 6.0 | CONTAMINANT FATE AND TRANSPORT..... | 6-1 |
| 6.1 | INTRODUCTION..... | 6-1 |
| 6.2 | PHYSICAL AND CHEMICAL PROPERTIES..... | 6-1 |
| 6.2.1 | Metals..... | 6-1 |
| 6.2.2 | Organic Compounds | 6-2 |
| 6.3 | CONCEPTUAL SITE MODEL..... | 6-6 |
| 6.3.1 | Water Balance Components | 6-7 |
| 6.3.2 | Contaminant Release Mechanisms and Migration Pathways | 6-7 |
| 6.4 | FATE AND TRANSPORT ANALYSIS | 6-8 |
| 6.4.1 | Soil Leachability Analysis..... | 6-8 |
| 6.4.2 | Vadose Zone SESOIL Modeling of the Contaminant Migration Constituents of Potential Concern..... | 6-10 |
| 6.4.3 | Saturated Zone Groundwater Modeling | 6-11 |
| 6.4.4 | Natural Attenuation of the Contaminant Migration Constituents of Potential Concern..... | 6-12 |
| 6.4.5 | Identification of Soil Remedial Levels..... | 6-13 |
| 7.0 | HUMAN HEALTH PRELIMINARY RISK EVALUATION METHODOLOGY | 7-1 |
| 7.1 | DATA EVALUATION..... | 7-1 |
| 7.2 | EXPOSURE EVALUATION | 7-3 |
| 7.2.1 | Receptor Assessment | 7-3 |
| 7.2.2 | Migration Pathway Analysis | 7-4 |
| 7.2.3 | Identification of Exposure Pathways..... | 7-5 |
| 7.3 | SELECTION OF SCREENING VALUES | 7-6 |
| 7.3.1 | Screening Values for Soils and Sediments..... | 7-7 |
| 7.3.2 | Screening Values for Groundwater | 7-9 |
| 7.3.3 | Screening Values for Surface Water | 7-9 |
| 7.4 | RISK EVALUATION | 7-10 |
| 7.5 | UNCERTAINTY | 7-10 |
| 7.6 | REMEDIAL LEVELS FOR PROTECTION OF HUMAN HEALTH | 7-11 |
| 8.0 | ECOLOGICAL PRELIMINARY RISK EVALUATION..... | 8-1 |
| 8.1 | ECOLOGICAL SCREENING VALUE COMPARISON (Step i)..... | 8-4 |
| 8.2 | PRELIMINARY PROBLEM FORMULATION (Step ii) | 8-4 |
| 8.2.1 | Environmental Setting..... | 8-7 |

CONTENTS (continued)

| | | |
|----------|---|--------|
| 8.2.2 | Surface Soil..... | 8-12 |
| 8.2.3 | Surface Water and Groundwater..... | 8-13 |
| 8.2.4 | Sediment..... | 8-14 |
| 8.3 | PRELIMINARY ECOLOGICAL EFFECTS EVALUATION (Step iii)..... | 8-15 |
| 8.4 | PRELIMINARY EXPOSURE ESTIMATE (Step iv)..... | 8-24 |
| 8.5 | PRELIMINARY RISK CALCULATION (Step v)..... | 8-34 |
| 8.6 | UNCERTAINTIES | 8-34 |
| 9.0 | RESULTS OF THE RCRA FACILITY INVESTIGATION AT THE 16 SWMUS RESULTING IN RECOMMENDATION OF NO FURTHER ACTION..... | 9-1 |
| 9.1 | SWMU 14: OLD FIRE TRAINING AREA..... | 9.1-1 |
| 9.1.1 | History and Description of SWMU 14, Old Fire Training Area | 9.1-1 |
| 9.1.2 | Summary of Phase II RCRA Facility Investigation Activities..... | 9.1-2 |
| 9.1.3 | Physical Characteristics of the Site | 9.1-3 |
| 9.1.4 | Nature and Extent of Contamination..... | 9.1-3 |
| 9.1.5 | Fate and Transport Considerations..... | 9.1-5 |
| 9.1.6 | Human Health Preliminary Risk Evaluation of SWMU 14..... | 9.1-6 |
| 9.1.7 | Ecological Preliminary Risk Evaluation of SWMU 14 | 9.1-7 |
| 9.1.8 | Conclusions and Risk Management and Site Recommendations for SWMU 14..... | 9.1-9 |
| 9.2 | SWMU 17: DRMO HAZARDOUS WASTE STORAGE AREA..... | 9.2-1 |
| 9.2.1 | History and Description of SWMU 17, DRMO Hazardous Waste Storage Area..... | 9.2-1 |
| 9.2.2 | Summary of Phase II RCRA Facility Investigation Activities..... | 9.2-1 |
| 9.2.3 | Physical Characteristics of the Site | 9.2-2 |
| 9.2.4 | Nature and Extent of Contamination..... | 9.2-3 |
| 9.2.5 | Fate and Transport Considerations..... | 9.2-5 |
| 9.2.6 | Human Health Preliminary Risk Evaluation of SWMU 17..... | 9.2-6 |
| 9.2.7 | Ecological Preliminary Risk Evaluation of SWMU 17 | 9.2-7 |
| 9.2.8 | Conclusions and Risk Management and Site Recommendations for SWMU 17..... | 9.2-9 |
| | ATTACHMENT 9.2A SWMU 17: DRMO HAZARDOUS WASTE STORAGE AREA, HUMAN HEALTH BASELINE RISK ASSESSMENT | 9.2A-1 |
| 9.2A.1 | IDENTIFICATION OF COPCS | 9.2A-3 |
| 9.2A.2 | EXPOSURE ASSESSMENT | 9.2A-3 |
| 9.2A.2.1 | Exposure Setting | 9.2A-3 |
| 9.2A.2.2 | Identification of Potential Receptor Populations and Exposure Pathways | 9.2A-3 |
| 9.2A.2.3 | Estimation of Exposure Concentrations | 9.2A-4 |
| 9.2A.2.4 | Quantification of Exposure | 9.2A-4 |
| 9.2A.3 | TOXICITY ASSESSMENT | 9.2A-5 |
| 9.2A.4 | RISK CHARACTERIZATION RESULTS..... | 9.2A-5 |
| 9.2A.4.1 | Current Land-use Scenarios | 9.2A-5 |
| 9.2A.4.2 | Future Land-use Scenarios | 9.2A-5 |
| 9.2A.5 | UNCERTAINTY ASSESSMENT..... | 9.2A-6 |
| 9.2A.6 | RISK SUMMARY..... | 9.2A-6 |

CONTENTS (continued)

| | | |
|----------|---|--------|
| 9.3 | SWMU 19: OLD SLUDGE DRYING BEDS | 9.3-1 |
| 9.3.1 | History and Description of SWMU 19, Old Sludge Drying Beds..... | 9.3-1 |
| 9.3.2 | Summary of Investigative Activities | 9.3-1 |
| 9.3.3 | Physical Characteristics of the Site | 9.3-1 |
| 9.3.4 | Nature and Extent of Contamination..... | 9.3-2 |
| 9.3.5 | Fate and Transport Considerations..... | 9.3-5 |
| 9.3.6 | Human Health Preliminary Risk Evaluation of SWMU 19..... | 9.3-5 |
| 9.3.7 | Ecological Preliminary Risk Evaluation of SWMU 19 | 9.3-6 |
| 9.3.8 | Conclusions and Risk Management and Site Recommendations for SWMU 19..... | 9.3-8 |
| | ATTACHMENT 9.3A SWMU 19: OLD SLUDGE DRYING BEDS, HUMAN HEALTH BASELINE RISK ASSESSMENT | 9.3A-1 |
| 9.3A.1 | IDENTIFICATION OF COPCS | 9.3A-3 |
| 9.3A.2 | EXPOSURE ASSESSMENT | 9.3A-3 |
| 9.3A.2.1 | Exposure Setting | 9.3A-3 |
| 9.3A.2.2 | Identification of Potential Receptor Populations and Exposure Pathways | 9.3A-3 |
| 9.3A.2.3 | Estimation of Exposure Concentrations | 9.3A-4 |
| 9.3A.2.4 | Quantification of Exposure | 9.3A-5 |
| 9.3A.3 | TOXICITY ASSESSMENT | 9.3A-5 |
| 9.3A.4 | RISK CHARACTERIZATION RESULTS | 9.3A-5 |
| 9.3A.4.1 | Current Land-use Scenarios | 9.3A-5 |
| 9.3A.4.2 | Future Land-use Scenarios | 9.3A-5 |
| 9.3A.5 | UNCERTAINTY ASSESSMENT..... | 9.3A-6 |
| 9.3A.6 | RISK SUMMARY | 9.3A-6 |
| 9.3A.7 | REMEDIAL LEVELS | 9.3A-7 |
| 9.3A.7.1 | Derivation of Remedial Levels..... | 9.3A-7 |
| 9.3A.7.2 | Remedial Level Recommendations | 9.3A-8 |
| 9.4 | SWMU 27A: 3D SQUADRON 7TH CAVALRY MOTORPOOL, BUILDING 1339A..... | 9.4-1 |
| 9.4.1 | History and Description of SWMU 27A, 3d Squadron 7th Cavalry Motorpool, Building 1339A..... | 9.4-1 |
| 9.4.2 | Summary of Investigative Activities | 9.4-1 |
| 9.4.3 | Physical Characteristics of the Site | 9.4-1 |
| 9.4.4 | Nature and Extent of Contamination..... | 9.4-2 |
| 9.4.5 | Fate and Transport Considerations..... | 9.4-3 |
| 9.4.6 | Human Health Preliminary Risk Evaluation of SWMU 27A, Building 1339A | 9.4-3 |
| 9.4.7 | Ecological Preliminary Risk Evaluation of SWMU 27A, Building 1339A | 9.4-4 |
| 9.4.8 | Conclusions and Risk Management and Site Recommendations for SWMU 27A, Building 1339A | 9.4-5 |
| 9.5 | SWMU 27A: 3D SQUADRON 7TH CAVALRY MOTORPOOL, BUILDING 1339B | 9.5-1 |
| 9.5.1 | History and Description of SWMU 27A, 3d Squadron 7th Cavalry Motorpool, Building 1339B..... | 9.5-1 |
| 9.5.2 | Summary of Investigative Activities | 9.5-1 |

CONTENTS (continued)

| | | |
|-----------|--|----------|
| 10.0 | RESULTS OF THE RCRA FACILITY INVESTIGATION AT THE 16 SWMUS RESULTING IN RECOMMENDATION OF FURTHER INVESTIGATION OR A CAP | 10-1 |
| 10.1 | SWMU 2: CAMP OLIVER LANDFILL | 10.1-1 |
| 10.1.1 | History and Description of SWMU 2, Camp Oliver Landfill..... | 10.1-1 |
| 10.1.2 | Summary of Phase II RCRA Facility Investigation Activities..... | 10.1-2 |
| 10.1.3 | Physical Characteristics of the Site | 10.1-3 |
| 10.1.4 | Nature and Extent of Contamination..... | 10.1-4 |
| 10.1.5 | Fate and Transport Considerations..... | 10.1-7 |
| 10.1.6 | Human Health Preliminary Risk Evaluation of SWMU 2..... | 10.1-9 |
| 10.1.7 | Ecological Preliminary Risk Evaluation of SWMU 2 | 10.1-11 |
| 10.1.8 | Conclusions and Risk Management and Site Recommendations for SWMU 2 | 10.1-14 |
| | ATTACHMENT 10.1A SWMU 2: CAMP OLIVER LANDFILL, HUMAN HEALTH BASELINE RISK ASSESSMENT..... | 10.1A-1 |
| 10.1A.1 | IDENTIFICATION OF COPCS | 10.1A-3 |
| 10.1A.2 | EXPOSURE ASSESSMENT | 10.1A-3 |
| 10.1A.2.1 | Exposure Setting | 10.1A-3 |
| 10.1A.2.2 | Identification of Potential Receptor Populations and Exposure Pathways | 10.1A-3 |
| 10.1A.2.3 | Estimation of Exposure Concentrations | 10.1A-5 |
| 10.1A.2.4 | Quantification of Exposure | 10.1A-6 |
| 10.1A.3 | TOXICITY ASSESSMENT | 10.1A-7 |
| 10.1A.4 | RISK CHARACTERIZATION RESULTS | 10.1A-7 |
| 10.1A.4.1 | Current Land-use Scenarios | 10.1A-7 |
| 10.1A.4.2 | Future Land-use Scenarios | 10.1A-8 |
| 10.1A.5 | UNCERTAINTY ASSESSMENT..... | 10.1A-9 |
| 10.1A.6 | RISK SUMMARY | 10.1A-10 |
| 10.1A.7 | REMEDIAL LEVELS | 10.1A-11 |
| 10.1A.7.1 | Derivation of Remedial Levels..... | 10.1A-11 |
| 10.1A.7.2 | Remedial Level Recommendations | 10.1A-12 |
| 10.2 | SWMU 3: TAC-X LANDFILL..... | 10.2-1 |
| 10.2.1 | History and Description of SWMU 3, TAC-X Landfill..... | 10.2-1 |
| 10.2.2 | Summary of Phase II RCRA Facility Investigation Activities..... | 10.2-2 |
| 10.2.3 | Physical Characteristics of the Site | 10.2-3 |
| 10.2.4 | Nature and Extent of Contamination..... | 10.2-4 |
| 10.2.5 | Fate and Transport Considerations..... | 10.2-7 |
| 10.2.6 | Human Health Preliminary Risk Evaluation of SWMU 3..... | 10.2-9 |
| 10.2.7 | Ecological Preliminary Risk Evaluation of SWMU 3 | 10.2-11 |
| 10.2.8 | Conclusions and Risk Management and Site Recommendations for SWMU 3 | 10.2-15 |
| | ATTACHMENT 10.2A SWMU 3: TAC-X LANDFILL, HUMAN HEALTH BASELINE RISK ASSESSMENT | 10.2A-1 |
| 10.2A.1 | IDENTIFICATION OF COPCS | 10.2A-3 |
| 10.2A.2 | EXPOSURE ASSESSMENT | 10.2A-3 |
| 10.2A.2.1 | Exposure Setting | 10.2A-3 |

CONTENTS (continued)

| | | |
|-----------|---|----------|
| 10.2A.2.2 | Identification of Potential Receptor Populations and Exposure Pathways | 10.2A-3 |
| 10.2A.2.3 | Estimation of Exposure Concentrations | 10.2A-5 |
| 10.2A.2.4 | Quantification of Exposure | 10.2A-6 |
| 10.2A.3 | TOXICITY ASSESSMENT | 10.2A-7 |
| 10.2A.4 | RISK CHARACTERIZATION RESULTS | 10.2A-8 |
| 10.2A.4.1 | Current Land-use Scenarios | 10.2A-8 |
| 10.2A.4.2 | Future Land-use Scenarios | 10.2A-9 |
| 10.2A.5 | UNCERTAINTY ASSESSMENT | 10.2A-11 |
| 10.2A.6 | RISK SUMMARY | 10.2A-11 |
| 10.2A.7 | REMEDIAL LEVELS | 10.2A-12 |
| 10.3 | SWMU 9: INACTIVE EOD AREA IN RED CLOUD RANGE, HOTEL AREA | 10.3-1 |
| 10.3.1 | History and Description of SWMU 9, Inactive EOD Area in Red Cloud Range, Hotel Area | 10.3-1 |
| 10.3.2 | Conclusions and Risk Management and Site Recommendations for SWMU 9 | 10.3-2 |
| 10.4 | SWMU 10: INACTIVE EOD AREA NORTH OF THE GARRISON AREA | 10.4-1 |
| 10.4.1 | History and Description of SWMU 10, Inactive EOD Area North of the Garrison Area | 10.4-1 |
| 10.4.2 | Summary of Phase II RCRA Facility Investigation Activities | 10.4-2 |
| 10.4.3 | Physical Characteristics of the Site | 10.4-2 |
| 10.4.4 | Nature and Extent of Contamination | 10.4-3 |
| 10.4.5 | Conclusions and Risk Management and Site Recommendations for SWMU 10 | 10.4-4 |
| 10.5 | SWMU 11: INACTIVE EOD AREA LOCATED THREE MILES NORTH OF THE GARRISON AREA | 10.5-1 |
| 10.5.1 | History and Description of SWMU 11, Inactive EOD Area Located Three Miles North of the Garrison Area | 10.5-1 |
| 10.5.2 | Summary of Phase II RCRA Facility Investigation Activities | 10.5-2 |
| 10.5.3 | Physical Characteristics of the Site | 10.5-2 |
| 10.5.4 | Nature and Extent of Contamination | 10.5-3 |
| 10.5.5 | Conclusions and Risk Management and Site Recommendations for SWMU 11 | 10.5-4 |
| 10.6 | SWMU 12A: ACTIVE EOD AREA CONTAINING OPEN DETONATION UNIT AND OPEN BURN UNIT | 10.6-1 |
| 10.6.1 | History and Description of SWMU 12A, Active EOD Area Containing Open Detonation Unit and Open Burn Unit | 10.6-1 |
| 10.6.2 | Summary of Phase II RCRA Facility Investigation Activities | 10.6-3 |
| 10.6.3 | Physical Characteristics of the Site | 10.6-4 |
| 10.6.4 | Nature and Extent of Contamination | 10.6-4 |
| 10.6.5 | Fate and Transport Considerations | 10.6-7 |
| 10.6.6 | Human Health Preliminary Risk Evaluation of SWMU 12A | 10.6-8 |
| 10.6.7 | Ecological Preliminary Risk Evaluation of SWMU 12A | 10.6-10 |

CONTENTS (continued)

| | | |
|------|---------------------------------------|------|
| 11.0 | CONCLUSIONS AND RECOMMENDATIONS | 11-1 |
| 11.1 | CONCLUSIONS | 11-1 |
| 11.2 | RECOMMENDATIONS | 11-2 |
| 12.0 | REFERENCES | 12-1 |

VOLUME II

APPENDICES A through G

| | | |
|---|--|-----|
| A | DIRECT-PUSH TECHNOLOGY AND BORING LOGS | A-1 |
| B | MONITORING WELL CONSTRUCTION DIAGRAMS | B-1 |
| C | QUALITY CONTROL SUMMARY REPORT | C-1 |
| D | COMPARISON OF METAL DATA FROM THE PHASE I AND PHASE II RCRA FACILITY INVESTIGATIONS | D-1 |
| E | GEOTECHNICAL LABORATORY TEST RESULTS | E-1 |
| F | BACKGROUND DATA SUMMARY | F-1 |
| G | CHAINS OF CUSTODY | G-1 |

VOLUME III

APPENDICES H through L

| | | |
|---|---|-----|
| H | ANALYTICAL LABORATORY DATA | H-1 |
| I | HUMAN HEALTH RISK ASSESSMENT METHODOLOGY | I-1 |
| J | TOXICITY PROFILES FOR CONTAMINANTS OF POTENTIAL CONCERN | J-1 |
| K | SEASONAL SOIL COMPARTMENT MODEL | K-1 |
| L | GEORGIA ENVIRONMENTAL PROTECTION DIVISION LETTER OF CONCURRENCE, MEETING MINUTES, AND AMENDED COMMENT RESPONSE TABLE (APRIL 2000) | L-1 |

TABLES

| | | |
|--|--|--------|
| ES-1 | Summary of Site-related Contaminants | ES-6 |
| ES-2 | Summary of CMCOPCs, HHCOPCs, and ECOPCs | ES-9 |
| ES-3 | SWMU-specific Recommendations | ES-12 |
| 2-1 | List of Solid Waste Management Units Included in This Report | 2-7 |
| 4-1 | Summary of Media and Investigation Technologies for the SWMUs | 4-2 |
| 4-2 | Summary of Media Sampled and Laboratory Analyses Performed for the SWMUs | 4-4 |
| 5-1 | Background Media Summary | 5-2 |
| 6-1 | List of Distribution Coefficients for the Inorganic SRCs at the 16 SWMUs | 6-2 |
| 6-2 | Physical and Chemical Properties of Organic Site-related Contaminants | 6-3 |
| 7-1 | Generalized Site Descriptions and Associated Receptor Populations | 7-4 |
| 8-1 | Ecological Screening Values for Surface Water and Sediment | 8-5 |
| 8-2 | Habitat Types at 16 SWMUs | 8-8 |
| 8-3 | Derivation of NOAELs for Mammal Test Species | 8-16 |
| 8-4 | Derivation of NOAELs for Bird Test Species | 8-20 |
| 8-5 | Derivation of NOAELs and Screening Toxicity Reference Values for Mammal Receptors | 8-25 |
| 8-6 | Derivation of NOAELs and Screening Toxicity Reference Values for Bird Receptors | 8-29 |
| 8-7 | Exposure Parameters for Surrogate Species | 8-33 |
| 8-8 | Exposure Parameters for Surrogate Species for Supplementary Risk Calculations | 8-37 |
| 8-9 | Derivation of LOAEL Toxicity Reference Values for Mammal Test Species | 8-38 |
| 8-10 | Derivation of LOAEL Toxicity Reference Values for Bird Test Species | 8-39 |
| 8-11 | Derivation of LOAEL Toxicity Reference Values for Mammal Receptors | 8-40 |
| 8-12 | Derivation of LOAEL Toxicity Reference Values for Bird Receptors | 8-41 |
| SWMU 14: Old Fire Training Area | | |
| 9.1-1 | Summary of RFA Results, SWMU 14 | 9.1-11 |
| 9.1-2 | Summary of Phase I RFI Results, SWMU 14 | 9.1-12 |
| 9.1-3 | Monitoring Well Construction Summary, SWMU 14 | 9.1-13 |
| 9.1-4 | Summary of Geotechnical Analyses, SWMU 14 | 9.1-13 |
| 9.1-5 | Well Development Summary, SWMU 14 | 9.1-14 |
| 9.1-6 | Field Parameter Measurements during Groundwater Sampling, SWMU 14 | 9.1-14 |
| 9.1-7 | Summary of Analytes Detected in Surface Soil, SWMU 14 | 9.1-15 |
| 9.1-8 | Summary of Analytes Detected in Subsurface Soil, SWMU 14 | 9.1-16 |
| 9.1-9 | Summary of Analytes Detected in Groundwater, SWMU 14 | 9.1-17 |
| 9.1-10 | Summary of Site-related Contaminants, SWMU 14 | 9.1-18 |
| 9.1-11 | GSSL Screening of Site-related Contaminants in Soil, SWMU 14 | 9.1-18 |
| 9.1-12 | Human Health Risk Screening for Surface Soil, Subsurface Soil, and Groundwater, SWMU 14 | 9.1-19 |
| 9.1-13 | Ecological Screening Value Comparison for Analytes Detected in Groundwater, SWMU 14 | 9.1-20 |
| 9.1-14 | Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 14 | 9.1-21 |
| 9.1-15 | Preliminary Risk Calculations for ECOPCs in Groundwater, SWMU 14 | 9.1-22 |

TABLES (continued)

SWMU 37: NGTC Equalization Basin

| | | |
|---------|--|---------|
| 9.30-1 | Summary of Geotechnical Analyses at NGTC Equalization Basin, SWMU 37 | 9.30-9 |
| 9.30-2 | Summary of Analytes Detected in Surface Soil, SWMU 37 | 9.30-9 |
| 9.30-3 | Summary of Analytes Detected in Subsurface Soil, SWMU 37 | 9.30-10 |
| 9.30-4 | Summary of Analytes Detected in Groundwater, SWMU 37 | 9.30-10 |
| 9.30-5 | Summary of Analytes Detected in Equalization Basin Wastewater, SWMU 37 | 9.30-11 |
| 9.30-6 | Summary of Analytes Detected in Equalization Basin Sediment, SWMU 37 | 9.30-11 |
| 9.30-7 | Summary of Site-related Contaminants, SWMU 37 | 9.30-12 |
| 9.30-8 | GSSL Screening of Site-related Contaminants in Soil and Sediment, SWMU 37 | 9.30-13 |
| 9.30-9 | Human Health Risk Screening for Surface Soil, Subsurface Soil, and Groundwater, SWMU 37 | 9.30-14 |
| 9.30-10 | Ecological Screening Value Comparison for Analytes Detected in Groundwater, SWMU 37 | 9.30-14 |
| 9.30-11 | Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 37 | 9.30-15 |
| 9.30-12 | Preliminary Risk Calculations for ECOPCs in Groundwater, SWMU 37 | 9.30-15 |

SWMU 2: Camp Oliver Landfill

| | | |
|---------|---|----------|
| 10.1-1 | Summary of Phase I RFI Results, SWMU 2 | 10.1-18 |
| 10.1-2 | Monitoring Well Construction Summary, SWMU 2 | 10.1-18 |
| 10.1-3 | Summary of Geotechnical Analyses, SWMU 2 | 10.1-19 |
| 10.1-4 | Well Development Summary, SWMU 2 | 10.1-19 |
| 10.1-5 | Field Parameter Measurements during Groundwater and Surface Water Sampling, SWMU 2 | 10.1-20 |
| 10.1-6 | Summary of Analytes Detected in Surface Soil, SWMU 2 | 10.1-21 |
| 10.1-7 | Summary of Analytes Detected in Subsurface Soil, SWMU 2 | 10.1-23 |
| 10.1-8 | Summary of Analytes Detected in Groundwater, SWMU 2 | 10.1-24 |
| 10.1-9 | Summary of Analytes Detected in Surface Water and Sediment, SWMU 2 | 10.1-27 |
| 10.1-10 | Summary of Site-related Contaminants, SWMU 2 | 10.1-28 |
| 10.1-11 | GSSL Screening of Site-related Contaminants in Soil and Sediment, SWMU 2 | 10.1-29 |
| 10.1-12 | Human Health Risk Screening for Surface Soil, Subsurface Soil, Groundwater, and Sediment, SWMU 2 | 10.1-30 |
| 10.1-13 | Ecological Screening Value Comparison for Analytes Detected in Sediment, SWMU 2 | 10.1-32 |
| 10.1-14 | Ecological Screening Value Comparison for Analytes Detected in Groundwater, SWMU 2 | 10.1-32 |
| 10.1-15 | Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 2 | 10.1-33 |
| 10.1-16 | Preliminary Risk Calculations for ECOPCs in Groundwater, SWMU 2 | 10.1-35 |
| 10.1-17 | Supplemental Risk Calculations for ECOPCs in Surface Soil for American Robin, SWMU 2 | 10.1-36 |
| 10.1-18 | Supplemental Risk Calculations for ECOPCs in Groundwater for Mink, SWMU 2 | 10.1-37 |
| 10.1-19 | Supplemental Risk Calculations for ECOPCs in Groundwater for Green Heron, SWMU 2 | 10.1-38 |
| 10.1A-1 | Selected Exposure and Modeled Concentrations, SWMU 2 | 10.1A-15 |
| 10.1A-2 | Exposure Parameters for Potential Receptor Populations, SWMU 2 | 10.1A-16 |
| 10.1A-3 | Estimated Intakes for Current Installation Worker, SWMU 2 | 10.1A-18 |
| 10.1A-4 | Estimated Intakes for Current Juvenile Trespasser, SWMU 2 | 10.1A-18 |
| 10.1A-5 | Estimated Intakes for Future On-site Installation Worker, SWMU 2 | 10.1A-19 |

TABLES (continued)

| | | |
|----------|---|----------|
| 10.1A-6 | Estimated Intakes for Future On-site Juvenile Trespasser, SWMU 2..... | 10.1A-19 |
| 10.1A-7 | Estimated Intakes for Future On-site Resident Child, SWMU 2..... | 10.1A-20 |
| 10.1A-8 | Estimated Intakes for Future On-site Resident Adult, SWMU 2..... | 10.1A-20 |
| 10.1A-9 | Estimated Intakes for Future Off-site Installation Worker, SWMU 2..... | 10.1A-21 |
| 10.1A-10 | Estimated Intakes for Future Off-site Resident Child, SWMU 2..... | 10.1A-21 |
| 10.1A-11 | Estimated Intakes for Future Off-site Resident Adult, SWMU 2..... | 10.1A-22 |
| 10.1A-12 | Estimated Intakes for Future Off-site Juvenile Wader, SWMU 2..... | 10.1A-22 |
| 10.1A-13 | Estimated Intakes for Future Off-site Sportsman, SWMU 2..... | 10.1A-22 |
| 10.1A-14 | Toxicity Values for Constituents of Potential Concern, SWMU 2..... | 10.1A-23 |
| 10.1A-15 | Hazard Indices and Carcinogenic Risks for Current On-site Installation Worker, SWMU 2..... | 10.1A-24 |
| 10.1A-16 | Hazard Indices and Carcinogenic Risks for Current Juvenile Trespasser, SWMU 2..... | 10.1A-24 |
| 10.1A-17 | Hazard Indices and Carcinogenic Risks for Future On-site Installation Worker, SWMU 2..... | 10.1A-25 |
| 10.1A-18 | Hazard Indices and Carcinogenic Risks for Future On-site Juvenile Trespasser, SWMU 2..... | 10.1A-25 |
| 10.1A-19 | Hazard Indices and Carcinogenic Risks for Future On-site Resident Child, SWMU 2..... | 10.1A-26 |
| 10.1A-20 | Carcinogenic Risks for Future On-site Resident Adult, SWMU 2..... | 10.1A-26 |
| 10.1A-21 | Hazard Indices and Carcinogenic Risks for Future Off-site Installation Worker, SWMU 2..... | 10.1A-27 |
| 10.1A-22 | Hazard Indices and Carcinogenic Risks for Future Off-site Resident Child, SWMU 2..... | 10.1A-27 |
| 10.1A-23 | Carcinogenic Risks for Future Off-site Resident Adult, SWMU 2..... | 10.1A-28 |
| 10.1A-24 | Hazard Indices for Future Off-site Juvenile Wader, SWMU 2..... | 10.1A-28 |
| 10.1A-25 | Hazard Indices for Future Off-site Sportsman, SWMU 2..... | 10.1A-28 |
| 10.1A-26 | Remedial Levels for Soil Based on Direct Exposure, SWMU 2..... | 10.1A-29 |
| 10.1A-27 | Target Groundwater Concentrations for Contaminant Migration Constituents of Concern, SWMU 2..... | 10.1A-29 |
| 10.1A-28 | Remedial Levels for Soil Based on Protection of Groundwater and Surface Water, SWMU 2..... | 10.1A-30 |

SWMU 3: TAC-X Landfill

| | | |
|---------|--|---------|
| 10.2-1 | Summary of Phase I RFI Results, SWMU 3..... | 10.2-20 |
| 10.2-2 | Monitoring Well Construction Summary, SWMU 3..... | 10.2-20 |
| 10.2-3 | Summary of Geotechnical Analyses, SWMU 3..... | 10.2-21 |
| 10.2-4 | Well Development Summary, SWMU 3..... | 10.2-21 |
| 10.2-5 | Field Parameter Measurements during Groundwater and Surface Water Sampling, SWMU 3..... | 10.2-22 |
| 10.2-6 | Summary of Analytes Detected in Surface Soil, SWMU 3..... | 10.2-23 |
| 10.2-7 | Summary of Analytes Detected in Subsurface Soil, SWMU 3..... | 10.2-24 |
| 10.2-8 | Summary of Analytes Detected in Groundwater, SWMU 3..... | 10.2-25 |
| 10.2-9 | Summary of Analytes Detected in Surface Water and Sediment, SWMU 3..... | 10.2-27 |
| 10.2-10 | Summary of Site-related Contaminants, SWMU 3..... | 10.2-28 |
| 10.2-11 | GSSL Screening of Site-related Contaminants in Soil, SWMU 3..... | 10.2-29 |
| 10.2-12 | GSSL Screening of Site-related Contaminants in Sediment, SWMU 3..... | 10.2-30 |

TABLES (continued)

| | | |
|----------|--|----------|
| 10.2-13 | Human Health Risk Screening for Surface Soil, Subsurface Soil, Groundwater, Surface Water, and Sediment, SWMU 3 | 10.2-31 |
| 10.2-14 | Ecological Screening Value Comparison for Analytes Detected in Surface Water, SWMU 3..... | 10.2-34 |
| 10.2-15 | Ecological Screening Value Comparison for Analytes Detected in Sediment, SWMU 3 | 10.2-35 |
| 10.2-16 | Ecological Screening Value Comparison for Analytes Detected in Groundwater, SWMU 3 | 10.2-36 |
| 10.2-17 | Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 3 | 10.2-37 |
| 10.2-18 | Preliminary Risk Calculations for ECOPCs in Surface Water, SWMU 3 | 10.2-38 |
| 10.2-19 | Preliminary Risk Calculations for ECOPCs in Sediment, SWMU 3 | 10.2-39 |
| 10.2-20 | Preliminary Risk Calculations for ECOPCs in Groundwater, SWMU 3 | 10.2-40 |
| 10.2-21 | Supplemental Risk Calculations for ECOPCs in Surface Soil for American Robin, SWMU 3 | 10.2-41 |
| 10.2-22 | Supplemental Risk Calculations for ECOPCs in Surface Water for Mink, SWMU 3..... | 10.2-42 |
| 10.2-23 | Supplemental Risk Calculations for ECOPCs in Surface Water for Green Heron, SWMU 3 | 10.2-43 |
| 10.2-24 | Supplemental Risk Calculations for ECOPCs in Sediment for Raccoon, SWMU 3 | 10.2-44 |
| 10.2-25 | Supplemental Risk Calculations for ECOPCs in Groundwater for Mink, SWMU 3..... | 10.2-45 |
| 10.2-26 | Supplemental Risk Calculations for ECOPCs in Groundwater for Green Heron, SWMU 3 | 10.2-46 |
| 10.2A-1 | Selected Exposure and Modeled Concentrations, SWMU 3 | 10.2A-14 |
| 10.2A-2 | Exposure Parameters for Potential Receptor Populations, SWMU 3 | 10.2A-15 |
| 10.2A-3 | Estimated Intakes for Current On-site Installation Worker, SWMU 3 | 10.2A-18 |
| 10.2A-4 | Estimated Intakes for Current On-site Juvenile Trespasser, SWMU 3 | 10.2A-18 |
| 10.2A-5 | Estimated Intakes for Current Off-site Juvenile Wader, SWMU 3..... | 10.2A-19 |
| 10.2A-6 | Estimated Intakes for Future On-site Installation Worker, SWMU 3 | 10.2A-19 |
| 10.2A-7 | Estimated Intakes for Future On-site Juvenile Trespasser, SWMU 3..... | 10.2A-20 |
| 10.2A-8 | Estimated Intakes for Future On-site Resident Child, SWMU 3 | 10.2A-20 |
| 10.2A-9 | Estimated Intakes for Future On-site Resident Adult, SWMU 3..... | 10.2A-21 |
| 10.2A-10 | Estimated Intakes for Future Off-site Installation Worker, SWMU 3 | 10.2A-21 |
| 10.2A-11 | Estimated Intakes for Future Off-site Juvenile Wader, SWMU 3 | 10.2A-22 |
| 10.2A-12 | Estimated Intakes for Future Off-site Resident Child, SWMU 3 | 10.2A-22 |
| 10.2A-13 | Estimated Intakes for Future Off-site Resident Adult, SWMU 3 | 10.2A-23 |
| 10.2A-14 | Toxicity Values for Constituents of Potential Concern, SWMU 3 | 10.2A-23 |
| 10.2A-15 | Hazard Indices and Carcinogenic Risks for Current On-site Installation Worker, SWMU 3 | 10.2A-24 |
| 10.2A-16 | Hazard Indices and Carcinogenic Risks for Current On-site Juvenile Trespasser, SWMU 3 | 10.2A-24 |
| 10.2A-17 | Hazard Indices and Carcinogenic Risks for Current Off-site Juvenile Wader, SWMU 3 | 10.2A-25 |
| 10.2A-18 | Hazard Indices and Carcinogenic Risks for Future On-site Installation Worker, SWMU 3 | 10.2A-25 |
| 10.2A-19 | Hazard Indices and Carcinogenic Risks for Future On-site Juvenile Trespasser, SWMU 3 | 10.2A-26 |
| 10.2A-20 | Hazard Indices and Carcinogenic Risks for Future On-site Resident Child, SWMU 3 | 10.2A-27 |
| 10.2A-21 | Carcinogenic Risks for Future On-site Resident Adult, SWMU 3 | 10.2A-27 |

TABLES (continued)

| | | |
|----------|---|----------|
| 10.2A-22 | Hazard Indices and Carcinogenic Risks for Future Off-site Installation Worker, SWMU 3 | 10.2A-28 |
| 10.2A-23 | Hazard Indices and Carcinogenic Risks for Future Off-site Juvenile Wader, SWMU 3 | 10.2A-29 |
| 10.2A-24 | Hazard Indices and Carcinogenic Risks for Future Off-site Resident Child, SWMU 3 | 10.2A-30 |
| 10.2A-25 | Carcinogenic Risks for Future Off-site Resident Adult, SWMU 3 | 10.2A-30 |
| 10.2A-26 | Remedial Levels for Surface Soil, SWMU 3 | 10.2A-31 |
| 10.2A-27 | Remedial Levels for Surface Water, SWMU 3 | 10.2A-31 |

SWMU 9: Inactive EOD Area in Red Cloud Range, Hotel Area

| | | |
|--------|--|--------|
| 10.3-1 | Summary of Phase I RFI Results for Inactive EOD Area in Red Cloud Range, Hotel Area (SWMU 9) | 10.3-2 |
|--------|--|--------|

SWMU 10: Inactive EOD Area North of the Garrison Area

| | | |
|--------|---|--------|
| 10.4-1 | Summary of Phase I RFI Results, SWMU 10 | 10.4-6 |
| 10.4-2 | Field Parameter Measurements during Surface Water Sampling, SWMU 10 | 10.4-6 |
| 10.4-3 | Summary of Analytes Detected in Surface Soil, SWMU 10 | 10.4-6 |
| 10.4-4 | Summary of Analytes Detected in Surface Water and Sediment, SWMU 10 | 10.4-7 |
| 10.4-5 | Summary of Site-related Contaminants, SWMU 10 | 10.4-7 |

SWMU 11: Inactive EOD Area Located Three Miles North of the Garrison Area

| | | |
|--------|--|--------|
| 10.5-1 | Summary of Phase I RFI Results, SWMU 11 | 10.5-5 |
| 10.5-2 | Summary of Analytes Detected in Phase II RFI Surface Soil, SWMU 11 | 10.5-5 |
| 10.5-3 | Summary of Site-related Contaminants, SWMU 11 | 10.5-5 |

SWMU 12A: Active EOD Area Containing Open Detonation Unit and Open Burn Unit

| | | |
|---------|---|---------|
| 10.6-1 | Summary of Phase I RFI Results, SWMU 12A | 10.6-19 |
| 10.6-2 | Summary of Subpart X Permit Application Sampling Event Results, SWMU 12A | 10.6-20 |
| 10.6-3 | Monitoring Well Construction Summary, SWMU 12A | 10.6-25 |
| 10.6-4 | Summary of Geotechnical Analyses, SWMU 12A | 10.6-25 |
| 10.6-5 | Well Development Summary, SWMU 12A | 10.6-26 |
| 10.6-6 | Field Parameter Measurements during Groundwater and Surface Water Sampling, SWMU 12A | 10.6-26 |
| 10.6-7 | Summary of Analytes Detected in Surface Soil, SWMU 12A | 10.6-27 |
| 10.6-8 | Summary of Analytes Detected in Subsurface Soil, SWMU 12A | 10.6-29 |
| 10.6-9 | Summary of Analytes Detected in Groundwater, SWMU 12A | 10.6-30 |
| 10.6-10 | Summary of Analytes Detected in Surface Water, SWMU 12A | 10.6-32 |
| 10.6-11 | Summary of Analytes Detected in Sediment, SWMU 12A | 10.6-33 |
| 10.6-12 | Summary of Site-related Contaminants, SWMU 12A | 10.6-34 |
| 10.6-13 | GSSL Screening of Site-related Contaminants in Soil, SWMU 12A | 10.6-35 |
| 10.6-14 | GSSL Screening of Site-related Contaminants in Sediment, SWMU 12A | 10.6-35 |
| 10.6-15 | Human Health Risk Screening for Surface Soil, Subsurface Soil, Groundwater, Surface Water, and Sediment, SWMU 12A | 10.6-36 |
| 10.6-16 | Ecological Screening Value Comparison for Analytes Detected in Surface Water, SWMU 12A | 10.6-38 |

FIGURES

| | | |
|--|--|---------|
| 2-1 | Regional Location Map for Fort Stewart Military Reservation, Georgia..... | 2-2 |
| 2-2 | Location Map for Fort Stewart Military Reservation, Georgia..... | 2-3 |
| 2-3 | Location Map for SWMUs Outside the Garrison Area, Fort Stewart, Georgia..... | 2-5 |
| 2-4 | Location Map of SWMUs Within the Garrison Area, Fort Stewart, Georgia..... | 2-6 |
| 3-1 | Approximate Geological Column for the FSMR Showing the Principal Hydrostratigraphy..... | 3-3 |
| 7-1 | General Process for Assessing Risk and Selecting Remedial Levels for Human Receptors..... | 7-2 |
| 8-1 | General Process for Assessing Risk and Selecting Remedial Levels for Ecological Receptors..... | 8-3 |
| SWMU 14: Old Fire Training Area | | |
| 9.1-1 | Sampling Locations and Phase I RFI Analytical Results in Subsurface Soil, SWMU 14..... | 9.1-23 |
| 9.1-2 | Cross Section A-A', SWMU 14..... | 9.1-25 |
| 9.1-3 | Cross Section B-B', SWMU 14..... | 9.1-26 |
| 9.1-4 | Groundwater Potentiometric Surface Map, SWMU 14..... | 9.1-27 |
| 9.1-5 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 14..... | 9.1-29 |
| 9.1-6 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 14..... | 9.1-30 |
| 9.1-7 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 14..... | 9.1-31 |
| 9.1-8 | Potential Migration and Exposure Pathways, SWMU 14..... | 9.1-32 |
| SWMU 17: DRMO Hazardous Waste Storage Area | | |
| 9.2-1 | Sampling Locations, SWMU 17..... | 9.2-22 |
| 9.2-2 | Cross Section A-A', SWMU 17..... | 9.2-23 |
| 9.2-3 | Cross Section B-B', SWMU 17..... | 9.2-24 |
| 9.2-4 | Groundwater Potentiometric Surface Map, SWMU 17..... | 9.2-25 |
| 9.2-5 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 17..... | 9.2-27 |
| 9.2-6 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 17..... | 9.2-28 |
| 9.2-7 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 17..... | 9.2-29 |
| 9.2-8 | Potential Migration and Exposure Pathways, SWMU 17..... | 9.2-30 |
| 9.2A-1 | Specific Conceptual Site Model for Acetone in Groundwater, SWMU 17..... | 9.2A-10 |
| SWMU 19: Old Sludge Drying Beds | | |
| 9.3-1 | Sampling Locations, SWMU 19..... | 9.3-25 |
| 9.3-2 | Summary of Analytical Results in Surface Soil, SWMU 19..... | 9.3-27 |
| 9.3-3 | Summary of Analytical Results in Subsurface Soil, SWMU 19..... | 9.3-28 |
| 9.3-4 | Summary of Analytical Results in Groundwater, SWMU 19..... | 9.3-29 |
| 9.3-5 | Potential Migration and Exposure Pathways, SWMU 19..... | 9.3-30 |
| SWMU 27A: 3d Squadron 7th Cavalry Motorpool, Building 1339A | | |
| 9.4-1 | Sampling Locations, SWMU 27A (Building 1339A)..... | 9.4-12 |
| 9.4-2 | Summary of Analytical Results in Surface Soil, Subsurface Soil, and Groundwater, SWMU 27A (Building 1339A)..... | 9.4-13 |
| 9.4-3 | Potential Migration and Exposure Pathways, SWMU 27A (Building 1339A)..... | 9.4-14 |

FIGURES (continued)

SWMU 32: Supply Diesel Tank

| | | |
|--------|---|---------|
| 9.28-1 | Sampling Locations and Phase I RFI Analytical Results for Surface Soil, SWMU 32 | 9.28-18 |
| 9.28-2 | Cross Section A-A', SWMU 32 | 9.28-19 |
| 9.28-3 | Cross Section B-B', SWMU 32 | 9.28-20 |
| 9.28-4 | Groundwater Potentiometric Surface Map, SWMU 32 | 9.28-21 |
| 9.28-5 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 32 | 9.28-23 |
| 9.28-6 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 32 | 9.28-24 |
| 9.28-7 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 32 | 9.28-25 |
| 9.28-8 | Potential Migration and Exposure Pathways, SWMU 32 | 9.28-26 |

SWMU 34: DEH Equipment Wash Rack

| | | |
|--------|--|---------|
| 9.29-1 | Sampling Locations, SWMU 34 | 9.29-19 |
| 9.29-2 | Cross Section A-A', SWMU 34 | 9.29-21 |
| 9.29-3 | Cross Section B-B', SWMU 34 | 9.29-22 |
| 9.29-4 | Groundwater Potentiometric Surface Map, SWMU 34 | 9.29-23 |
| 9.29-5 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 34 | 9.29-25 |
| 9.29-6 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 34 | 9.29-26 |
| 9.29-7 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 34 | 9.29-27 |
| 9.29-8 | Potential Migration and Exposure Pathways, SWMU 34 | 9.29-28 |

SWMU 37: NGTC Equalization Basin

| | | |
|--------|---|---------|
| 9.30-1 | Sampling Locations, SWMU 37 | 9.30-16 |
| 9.30-2 | Summary of Analytical Results in Surface Soil, SWMU 37 | 9.30-17 |
| 9.30-3 | Summary of Analytical Results in Subsurface Soil, SWMU 37 | 9.30-18 |
| 9.30-4 | Summary of Analytical Results in Groundwater, SWMU 37 | 9.30-19 |
| 9.30-5 | Potential Migration and Exposure Pathways, SWMU 37 | 9.30-20 |

SWMU 2: Camp Oliver Landfill

| | | |
|---------|---|----------|
| 10.1-1 | Sampling Locations and Phase I RFI Analytical Results in Surface Soil, SWMU 2 | 10.1-39 |
| 10.1-2 | Cross Section A-A', SWMU 2 | 10.1-41 |
| 10.1-3 | Cross Section B-B', SWMU 2 | 10.1-42 |
| 10.1-4 | Groundwater Potentiometric Surface Map, SWMU 2 | 10.1-43 |
| 10.1-5 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 2 | 10.1-45 |
| 10.1-6 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 2 | 10.1-46 |
| 10.1-7 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 2 | 10.1-47 |
| 10.1-8 | Summary of Phase II RFI Analytical Results in Sediment, SWMU 2 | 10.1-48 |
| 10.1-9 | Potential Migration and Exposure Pathways, SWMU 2 | 10.1-49 |
| 10.1-10 | Location of Chromium and Mercury in Soil above Remedial Levels, SWMU 2 | 10.1-50 |
| 10.1A-1 | Migration and Exposure Pathways for Receptors Addressed in the Baseline Risk Assessment, SWMU 2 | 10.1A-31 |

SWMU 3: TAC-X Landfill

| | | |
|--------|---|---------|
| 10.2-1 | Sampling Locations and Phase I RFI Analytical Results in Surface Soil, SWMU 3 | 10.2-47 |
| 10.2-2 | Cross Section A-A', SWMU 3 | 10.2-49 |

FIGURES (continued)

| | | |
|---|--|----------|
| 10.2-3 | Cross Section B-B', SWMU 3 | 10.2-50 |
| 10.2-4 | Groundwater Potentiometric Surface Map, SWMU 3 | 10.2-51 |
| 10.2-5 | Deep Groundwater Potentiometric Surface Map (March 1998), SWMU 3 | 10.2-52 |
| 10.2-6 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 3 | 10.2-53 |
| 10.2-7 | Summary of Phase II RFI Analytical Results in Subsurface Soil, SWMU 3 | 10.2-54 |
| 10.2-8 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 3 | 10.2-55 |
| 10.2-9 | Summary of Phase II RFI Analytical Results in Surface Water and Sediment, SWMU 3 | 10.2-56 |
| 10.2-10 | Potential Migration and Exposure Pathways, SWMU 3 | 10.2-57 |
| 10.2A-1 | Migration and Exposure Pathways for Receptors Addressed in the Baseline Risk Assessment, SWMU 3 | 10.2A-32 |
| SWMU 9: Inactive EOD Area in Red Cloud Range, Hotel Area | | |
| 10.3-1 | Sampling Locations, SWMU 9 | 10.3-3 |
| SWMU 10: Inactive EOD Area North of the Garrison Area | | |
| 10.4-1 | Sampling Locations and Phase I RFI Analytical Results for Surface Soil, SWMU 10 | 10.4-8 |
| 10.4-2 | Summary of Phase II RFI Analytical Results in Surface Soil, SWMU 10 | 10.4-9 |
| 10.4-3 | Summary of Phase II RFI Analytical Results in Surface Water and Sediment, SWMU 10 | 10.4-10 |
| SWMU 11: Inactive EOD Area Located Three Miles North of the Garrison Area | | |
| 10.5-1 | Sampling Locations and Phase I RFI Analytical Results for Surface Soil, SWMU 11 | 10.5-6 |
| 10.5-2 | Summary of Phase II RFI Analytical Results in Surface Soil and Groundwater, SWMU 11 | 10.5-7 |
| SWMU 12A: Active EOD Area Containing Open Detonation Unit and Open Burn Unit | | |
| 10.6-1 | Sampling Locations and Phase I RFI and Subpart X Analytical Results in Surface Soil and Subsurface Soil, SWMU 12A | 10.6-47 |
| 10.6-2 | Cross Section A-A', SWMU 12A | 10.6-48 |
| 10.6-3 | Cross Section B-B', SWMU 12A | 10.6-49 |
| 10.6-4 | Groundwater Potentiometric Surface Map, SWMU 12A | 10.6-50 |
| 10.6-5 | Summary of Phase II RFI Analytical Results in Surface Soil and Subsurface Soil, SWMU 12A | 10.6-51 |
| 10.6-6 | Summary of Phase II RFI Analytical Results in Groundwater, SWMU 12A | 10.6-52 |
| 10.6-7 | Summary of Phase II RFI Analytical Results in Surface Water and Sediment, SWMU 12A | 10.6-53 |
| 10.6-8 | Potential Migration and Exposure Pathways, SWMU 12A | 10.6-54 |
| 10.6A-1 | Migration and Exposure Pathways for Receptors Addressed in the Baseline Risk Assessment, SWMU 12A | 10.6A-43 |
| SWMU 18: Industrial Wastewater Treatment Plant | | |
| 10.7-1 | Phase I RFI Sampling Locations, SWMU 18 | 10.7-52 |
| 10.7-2 | Phase II RFI Sampling Locations, SWMU 18 | 10.7-53 |
| 10.7-3 | Cross Section A-A', SWMU 18 | 10.7-55 |

ACRONYMS

| | |
|---------|---|
| ADA | Air Defense Artillery |
| ADD | average daily dose |
| amsl | above mean sea level |
| ARAR | applicable or relevant and appropriate requirement |
| Army | U.S. Army |
| AST | aboveground storage tank |
| ASTM | American Society for Testing and Materials |
| AT123D | Analytical Transient 1-, 2-, 3-Dimensional |
| AUF | area use factor |
| AWQC | Ambient Water Quality Criteria |
| BAF | bioaccumulation factor |
| BCF | bioconcentration factor |
| bgs | below ground surface |
| BHC | benzene hexachloride |
| BN | battalion |
| BTEX | benzene, toluene, ethylbenzene, and xylenes |
| CAP | Corrective Action Plan |
| CB | chemical/biological |
| CERCLA | Comprehensive Environmental Response, Compensation, and Liability Act |
| CMCOC | contaminant migration chemical of concern |
| CMCOPC | contaminant migration constituent of potential concern |
| COC | contaminant of concern |
| COPC | contaminant of potential concern |
| CSM | Conceptual Site Model |
| DAF | dilution attenuation factor |
| DEH | Directorate of Engineering and Housing |
| DF | dilution factor |
| DO | dissolved oxygen |
| DOL | Directorate of Logistics |
| DPT | direct-push technology |
| DPW | Directorate of Public Works |
| DQO | data quality objective |
| DRMO | Defense Reutilization and Marketing Organization |
| ECOPC | ecological contaminant of potential concern |
| EOD | Explosive Ordnance Disposal |
| EPA | U.S. Environmental Protection Agency |
| EPRE | ecological preliminary risk evaluation |
| EP Tox | Extraction Procedure Toxicity |
| ERA | Ecological Risk Assessment |
| ESV | ecological screening value |
| FR | <u>Federal Register</u> |
| FSMR | Fort Stewart Military Reservation |
| GAF | gastrointestinal absorption factor |
| GEPD | Georgia Environmental Protection Division |
| GSSL | generic soil screening level |
| HAZWRAP | Hazardous Waste Remedial Actions Program |
| HELP | Hydrologic Evaluation of Landfill Performance |

ACRONYMS (continued)

| | |
|--------|--|
| HHCOC | human health chemical of concern |
| HHCOPC | human health contaminant of potential concern |
| HHPRE | human health preliminary risk evaluation |
| HI | hazard index |
| HMX | octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine |
| HQ | hazard quotient |
| IDW | investigation-derived waste |
| IEUBK | Integrated Exposure Uptake Biokinetic |
| ILCR | incremental lifetime cancer risk |
| IRA | Interim Removal Action |
| IWTP | Industrial Wastewater Treatment Plant |
| LAS | Land Application System |
| LOAEL | lowest observed adverse effect level |
| MCL | maximum contaminant level |
| MOGAS | motor gasoline |
| NFA | no further action |
| NGTC | National Guard Training Center |
| NGVD | National Geodetic Vertical Datum |
| NOAEL | no observed adverse effect level |
| NPDES | National Pollutant Discharge Elimination System |
| NTU | nephelometric turbidity unit |
| OB | open burn |
| OD | open detonation |
| ODAST | One-dimensional Analytical Solute Transport |
| OWS | oil/water separator |
| PAH | polycyclic aromatic hydrocarbon |
| PCB | polychlorinated biphenyl |
| PETN | pentaerythrite tetranitrate |
| PID | photoionization detector |
| POL | petroleum, oil, and lubricants |
| POTW | publicly owned treatment works |
| PVC | polyvinyl chloride |
| QA | quality assurance |
| QAPP | Quality Assurance Project Plan |
| QC | quality control |
| QCSR | Quality Control Summary Report |
| RBC | risk-based concentration |
| RBCA | Risk-based Corrective Action |
| RCRA | Resource Conservation and Recovery Act |
| RDX | hexahydro-1,3,5-trinitro-1,3,5-triazine |
| Redox | oxidation-reduction |
| RFA | RCRA Facility Assessment |
| RFI | RCRA Facility Investigation |
| SAIC | Science Applications International Corporation |
| SAP | Sampling and Analysis Plan |
| SDWA | Safe Drinking Water Act |
| SESOIL | Seasonal Soil Compartment Model |
| SMCL | secondary maximum contaminant level |

ACRONYMS (continued)

| | |
|--------|--|
| SPT BN | Support Battalion |
| SRC | site-related contaminant |
| SSL | soil screening level |
| SQB | sediment quality benchmark |
| SVOC | semivolatile organic compound |
| SWMU | Solid Waste Management Unit |
| TAC | Tactical Air Command |
| TC | toxicity characteristic |
| TCLP | Toxicity Characteristic Leaching Procedure |
| TDS | total dissolved solids |
| TEF | toxicity equivalence factor |
| TOC | total organic carbon |
| TPH | total petroleum hydrocarbons |
| TRPH | total recoverable petroleum hydrocarbons |
| TRV | toxicity reference value |
| USACE | U.S. Army Corps of Engineers |
| USGS | U.S. Geological Survey |
| UST | underground storage tank |
| UXO | unexploded ordnance |
| VOC | volatile organic compound |
| WQS | water quality standard |

EXECUTIVE SUMMARY

This report summarizes the results of the Phase II Resource Conservation and Recovery Act (RCRA) Facility Investigation (RFI) for the 16 Solid Waste Management Units (SWMUs) at Fort Stewart, Georgia. The 16 SWMUs include: Camp Oliver Landfill, SWMU 2; TAC-X Landfill, SWMU 3; Inactive EOD Area in Red Cloud Range, Hotel Area, SWMU 9; Inactive EOD Area North of Garrison Area, SWMU 10; Inactive EOD Area Located Approximately Three Miles Northeast of Garrison Area, SWMU 11; Active EOD Containing Open Detonation Unit and Open Burn Unit, SWMU 12A; Old Fire Training Area, SWMU 14; DRMO Hazardous Waste Storage Area, SWMU 17; Industrial Wastewater Treatment Plant, SWMU 18; Old Sludge Drying Beds, SWMU 19; Old Radiator Shop/Paint Booth, SWMU 24B; Motorpools, SWMUs 27A through 27V; Evans Army Heliport POL Storage Facility, SWMU 29; DEH Asphalt Tanks, SWMU 31; Supply Diesel Tank, SWMU 32; DEH Equipment Wash Rack, SWMU 34; and NGTC Equalization Basin, SWMU 37. Four of the 16 sites—Old Sludge Drying Beds, SWMU 19; Old Radiator Shop/Paint Booth, SWMU 24B; Motorpools, SWMUs 27A through 27V; and NGTC Equalization Basin, SWMU 37—had not been investigated previously and were investigated as Phase I RFIs. This report has been prepared by Science Applications International Corporation (SAIC) for the U.S. Army Corps of Engineers (USACE), Savannah District, under Contract DACA21-95-D-0022, Delivery Order No. 0009. The RFI was conducted in accordance with USACE Guidance EM 200-1-3 and the Georgia Environmental Protection Division (GEPD)—approved Sampling and Analysis Plan (SAP) (SAIC 1997).

The 16 SWMUs investigation consisted of 38 SWMU sites (including 22 motorpool sites) as designated under Hazardous Waste Permit HW-045. The sites were divided into 45 distinct geographic areas for investigation. Seven (SWMUs 2, 3, 9, 10, 11, 12A, and 29) of the 38 SWMUs are located outside the garrison area. The remaining 31 (SWMUs 14, 17, 18, 19, 24B, 27A through 27V, 31, 32, 34, and 37) are located within the garrison area.

OBJECTIVES AND SCOPE OF THE INVESTIGATION

The specific objectives of the Phase I and Phase II RFIs for the 16 SWMUs at Fort Stewart, Georgia, as defined in the Phase II RFI SAP (SAIC 1997) (approved by the GEPD in October 1997) are listed below.

Phase I RFI

- Determine if contamination of the environment has occurred.
- Determine whether contaminants, if present, constitute a threat to human health or the environment.
- Determine the need for future action and/or no further action (NFA).

Phase II RFI

- Determine the horizontal and vertical extent of contamination.
- Determine whether contaminants present a threat to human health or the environment.

- Determine the need for future action and/or NFA.
- Gather data necessary to support a Corrective Action Plan (CAP), if warranted.

The information provided in this report is based upon data collected previously during the Phase I RFI (if available) and data collected as part of the Phase II field sampling and analysis. At some of the sites, the Phase II sampling program incorporated an observational approach to sampling, as defined in the Phase II RFI SAP (SAIC 1997). This observational approach used field screening techniques to determine the horizontal and vertical extent of contamination at the SWMU and to identify suitable locations for installation of permanent monitoring wells. The scope of the fieldwork for the Phase I and Phase II sites included the activities listed below.

Phase I Sites

- Collection of direct-push soil samples using a push probe.
- Collection of direct-push groundwater samples using a push probe.
- Installation of permanent groundwater monitoring points or monitoring wells to confirm the nature of potential contamination at a specific push-probe location.
- Collection of surface water and sediment samples at SWMUs at which surface water and sediment were available.
- Surveying of the positions of all sample locations.

Phase II Sites

- Collection of direct-push soil samples using a push probe.
- Collection of direct-push groundwater samples using a push probe, including vertical-profile probes.
- Installation of permanent groundwater monitoring wells both upgradient and downgradient of the site.
- Groundwater sampling at existing monitoring wells (if available) and sampling of newly installed wells around the SWMUs.
- Collection of surface water and sediment samples at SWMUs at which surface water and sediment were available.
- Surveying of the positions of all sample locations.

Nature and Extent of Contamination

Site-related contaminants (SRCs) were identified for each site by comparing the analytical results obtained from soil, groundwater, surface water, and sediment against the reference background criteria. Contaminants with concentrations above the reference background criteria were identified as SRCs. The results of the chemical analyses on surface soil, subsurface soil, and groundwater were screened against the reference

background criteria for the Fort Stewart Military Reservation. Surface water and sediment were screened against site-specific background criteria.

In general, reference background samples were collected from each medium at locations upgradient or upstream of each site so as to be representative of naturally occurring conditions at sites under investigation. Upgradient or upstream samples were not collected at sites under a Phase I RFI (i.e., SWMUs 19, 24B, 27A through 27V and 37). The reference background concentrations for surface soil, subsurface soil, and groundwater were calculated as two times the average concentration of all of the locations selected to be in the background data set. If a chemical was not detected at a site, then one-half the detection limit was used as the concentration when calculating the reference mean background concentration. Surface water and sediment background samples were collected during the Phase II RFI and applied to the SWMUs on a site-specific basis.

Inorganics were considered to be SRCs if their concentrations were above the reference background concentrations, while organics were considered SRCs if they were simply detected because organic constituents are considered to potentially be man-made. SRCs from the nature and extent of contamination evaluation were further evaluated as potential concerns based upon fate and transport characteristics and upon their potential risk to human health and ecological receptors. A summary of SRCs by medium for each SWMU is presented in Table ES-1.

Fate and Transport Analysis

Fate and transport analysis was performed on each SWMU. This analysis included developing a site-specific Conceptual Site Model (CSM) identifying potential contaminant release and migration pathways and determining the potential for SRCs in surface soil, subsurface soil, and/or sediment to migrate to groundwater.

The maximum concentrations of the SRCs determined from nature and extent analysis were compared to U.S. Environmental Protection Agency (EPA) Generic Soil Screening Levels (GSSLs). Generally, if contaminant concentrations in soil fall below the GSSLs and there are no significant ecological receptors of concern, then no further study or action is warranted. SRCs were identified as contaminant migration constituents of potential concern (CMCOPCs) if they were detected at concentrations that exceeded their respective GSSLs. To evaluate leaching of CMCOPCs from soil to groundwater at the 16 SWMUs, groundwater concentrations of CMCOPCs were compared to maximum contaminant levels (MCLs). If an MCL for a chemical was not available, the groundwater concentration was compared to the risk-based concentration, as established by EPA Region III (EPA 1999b). A summary of the results of the fate and transport analysis (CMCOPCs) is presented in Table ES-2.

A weight-of-evidence approach was used to evaluate each CMCOPC identified based on leaching to groundwater. In some instances, the potential impact of CMCOPCs to groundwater, and possibly to surface water, was evaluated (modeled concentrations were compared to risk-based criteria) in a human health baseline risk assessment. CMCOPCs that indicated a potential risk to human health (i.e., that exceeded risk-based screening criteria) from modeling were identified as contaminant migration chemicals of concern, and remedial levels were developed based on protection of groundwater. SWMUs for which a human health baseline risk assessment was performed are identified in Table ES-2.

Human Health Preliminary Risk Evaluation

A human health preliminary risk evaluation (HHPRE) using a Step i risk evaluation approach based on guidance from GEPD was performed for each SWMU to determine the potential human health risks associated with the maximum concentrations of identified SRCs. The Step i risk evaluation involves the components listed below.

- For inorganics, compare detected concentrations to naturally occurring background levels to determine if detected inorganics are naturally occurring or are associated with past activities at the site.
- Identify potential migration and exposure pathways associated with the site and identify potential exposure scenarios to determine appropriate action levels.
- Identify available risk-based action levels for each contaminant detected above background levels or develop levels if they do not exist.
- Compare sample concentrations to action levels to determine if site conditions warrant further evaluation.

Chemicals that exceeded action levels were identified as human health contaminants of potential concern (HHCOPCs). A summary of the HHPRE results (HHCOPCs) is presented in Table ES-2.

A weight-of-evidence approach was used to evaluate each HHCOPC identified in the preliminary risk assessment. In some instances, HHCOPCs were evaluated further in a human health baseline risk assessment. HHCOPCs and/or CMCOPCs (see previous section) that either had hazard indices of 0.1 or incremental lifetime cancer risks of 1×10^{-6} were identified as human health contaminants of concern. Remedial levels were developed that were protective of the most sensitive receptor population, based on a minimum risk level of 3.0 for the total hazard index and 1×10^{-4} for the total incremental lifetime cancer risk. SWMUs for which a human health baseline risk assessment was performed are identified in Table ES-2.

Ecological Preliminary Risk Evaluation

An ecological preliminary risk evaluation (EPRE) based on guidance from GEPD was performed to determine the potential risk to ecological receptors associated with the maximum concentrations of the identified SRCs. The EPRE compared measured concentrations of detected substances to conservative ecological screening values to identify substances detected at the facility that pose a potential hazard to ecological receptors and that are identified as ecological contaminants of potential concern (ECOPCs). A summary of the results of the EPRE (ECOPCs) is presented in Table ES-2.

A weight-of-evidence approach was used to evaluate each ECOPC identified in the preliminary risk evaluation. In some instances, ECOPCs were evaluated further in a supplemental preliminary risk evaluation (SPRE). The SPRE presented a comparison of more realistic exposure estimates to toxicity reference values based on the lowest observed adverse effects levels. The exposure estimates were calculated using measured concentrations and more realistic exposure assumptions such as diets, absorption efficiencies, and area use factors. SWMUs for which an SPRE was performed are identified in Table ES-2.

Conclusions and Recommendations

A weight-of-evidence approach was used with the results from the fate and transport evaluation, HHPRE, human health baseline risk assessment (if performed), EPRE, and SPRE (if performed) to determine the recommendation for each SWMU. The recommendations fell into the following three categories:

- **No Further Action:** NFA was recommended for a SWMU if: (1) the contaminant levels in soil, groundwater, surface water, and sediment were below the reference background criteria, fate and transport values (GSSLs), and/or human health or ecological screening criteria or (2) significant uncertainty was evident, indicating minimal potential risk of migration to groundwater and/or a surface water body and/or to human health and ecological receptors.

- **Additional Investigation (Phase II RFI or additional monitoring):** A Phase II RFI or additional monitoring was recommended if the nature and extent of potential contaminants had not been determined, and further investigation or additional monitoring was required to evaluate extent or potential migration in the future.
- **Corrective Action Plan:** A CAP was recommended if the nature and extent of contamination at a SWMU was determined by the Phase II RFI, there was a potential risk of migration of contaminants to groundwater and/or surface water bodies or a potential risk to human health and ecological receptors, or institutional controls need to be applied to protect the health and safety of humans coming in contact with the site (i.e., inactive EOD areas). Such a site requires a CAP to evaluate appropriate remedial actions to eliminate or minimize these potential risks.

The recommendations for each SWMU are presented in Table ES-3.

Table ES-1. Summary of Site-related Contaminants

| SWMU | Type of Investigation | Site-related Contaminants | | | | |
|----------------------|-----------------------|--|--------------------------------------|---|----------------------------|--|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment |
| 2 | Phase II | 2 VOCs, 14 pest., 1 SVOC, and 6 metals | 1 VOC, 3 pest., 1 SVOC, and 3 metals | 3 VOCs and 3 metals | None | alpha-Chlordane |
| 3 | Phase II | 4 pest., BEHP, As, Cr, and Pb | 2 VOCs, BEHP, 3 pest., Cr, and Cd | 3 VOCs, 3 pest., Ba, Cd, Cr, Pb, and Hg | 1 SVOC, As, Ba, Cr, and Pb | 6 VOCs, As, Ba, Cr, Pb, Hg, and Se |
| 9 | Phase I ^d | As, Cr, and Ag | NC | NC | NP | NP |
| 10 | Phase II | As, Ba, Cr, and Pb | NC ^b | None | Cd, Cr, and Hg | As, Ba, and Pb |
| 11 | Phase II | As, Ba, Cr, Pb, and Ag | NC ^b | None | NP | NP |
| 12A | Phase II | 3 SVOCs, 4 exp., and 16 metals | Al, As, Ba, Cr, Fe, Pb, and V | BEHP, 1 exp., and 8 metals | RDX, Pb, Mn, and Hg | 1 SVOC, 1 exp., and 9 metals |
| 14 | Phase I | 2 VOCs, BEHP, and Hg | 5 VOCs, Cr, and Hg | 1 VOC, Pb, and Hg | NP | NP |
| 17 | Phase II | 1 VOC | 3 VOCs | 3 VOCs and Pb | None | None |
| 18 | Phase II | 1 VOC, Pb, and Hg | 5 VOCs, 2 SVOCs, Ba, Cr, Pb, and Hg | 9 VOCs, Ba, Cd, and Pb | 1 SVOC and Ba | (6 VOCs, 4 SVOCs, As, Ba, Cd, Cr, Pb, Hg, Se, and Ag) ^c |
| 19 | Phase I | 4 VOCs, 7 pest., and 5 metals | 6 VOCs, 9 pest., and 5 metals | BEHP, 7 pest., and 3 metals | NP | NP |
| 24B | Phase I | 1 VOC, 10 SVOCs, and 6 metals | 2 VOCs | 1 VOC, 11 SVOCs, and Hg | NP | NP |
| 27A (Bldg. 1339A) | Phase I | None | 2 VOCs and 3 SVOCs | 2 VOCs and BEHP | NP | NP |
| 27A (Bldg. 1339B) | Phase I | BEHP and Pb | 2 VOCs | 1 VOC | NP | NP |
| 27A (Bldg. 1322) | Phase I | 3 VOCs and Pb | 3 VOCs | Acetone | NP | NP |
| 27B | Phase I | None | 1 VOC | ND | NP | NP |
| 27C | Phase I | 1 VOC | 2 VOCs and 1 SVOC | 4 VOCs | NP | NP |
| 27D | Phase I | 3 VOCs | 1 VOC | None | NP | NP |
| 27E (Bldg. 1628) | Phase I | None | 1 VOC | None | NP | NP |

Note: Footnotes appear on page ES-8.

Table ES-1. Summary of Site-related Contaminants (continued)

| SWMU | Type of Investigation | Site-related Contaminants | | | | |
|------------------------|-----------------------|---------------------------|------------------------------|---------------------|-----------------|-----------------|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment |
| 27E (Bldg. 1720) | Phase I | NC | 2 VOCs and BEHP | 1 SVOC | NP | NP |
| 27F (NW Bldg. 1340) | Phase I | NC | 3 VOCs and Pb | 10 VOCs and 4 SVOCs | NP | NP |
| 27F (NE Bldg. 1340) | Phase I | 3 VOCs | 8 VOCs and 4 SVOCs | None | NP | NP |
| 27G | Phase I | NC | 3 VOCs | 1 SVOC | NP | NP |
| 27H (Bldg. 1071) | Phase I | NC | 2 VOCs, 11 SVOCs, Pb, and Hg | 1 VOC and 9 SVOCs | NP | NP |
| 27H (Bldg. 1056) | Phase I | NC | 1 VOC, 1 SVOC, Cd, and Pb | 2 VOCs and 4 SVOCs | NP ^d | NP ^d |
| 27I (Block 9900) | Phase I | NC | 1 VOC and Pb | None | NC | 1 VOC and Pb |
| 27I (Block 10300) | Phase I | NC | None | None | Pb | None |
| 27J (Bldg. 10535) | Phase I | None | None | 1 VOC and 1 SVOC | NP | NP |
| 27J (Bldg. 10531) | Phase I | 1 VOC and 1 SVOC | NC | 2 SVOCs | NP | NP |
| 27K | Phase I | NC | 4 VOCs | 1 VOC | NP | NP |
| 27L (Block 10200) | Phase I | None | 1 VOC and 1 SVOC | 8 VOCs and 2 SVOCs | Acetone | None |
| 27M (Block 10100) | Phase I | 1 VOC and Pb | 2 SVOCs and Pb | 1 VOC | NC | Pb |
| 27N (Block 9800) | Phase I | NC | 2 SVOCs and Pb | None | NC | 5 SVOCs |
| 27O (Block 9700) | Phase I | Pb | None | 1 SVOC | 1 VOC | Pb |
| 27P (Block 9500) | Phase I | 1 VOC and 1 SVOC | 1 VOC, 6 SVOCs, and Pb | None | NC | 1 VOC and Pb |

Note: Footnotes appear on page ES-8.

Table ES-1. Summary of Site-related Contaminants (continued)

| SWMU | Type of Investigation | Site-related Contaminants | | | |
|---------------------|-----------------------|-------------------------------------|---------------------------|---------------------------------|---|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water |
| 27Q (Block 9400) | Phase I | Pb | None | None | NC |
| 27R | Phase I | None | 1 VOC and 2 SVOCs | None | NP |
| 27S | Phase I | NC | 6 VOCs | None | NP |
| 27T | Phase I | 4 SVOCs | None | 1 VOC and 1 SVOC | NC |
| 27U | Phase I | 1 VOC and Pb | 2 VOCs and Pb | 4 VOCs | NP |
| 27V | Phase I | 1 VOC and Pb | 1 VOC and Pb | None | NP |
| 29 | Phase II | 8 VOCs and Ag | 16 VOCs and 14 SVOCs | 3 VOCs, 3 SVOCs, As, Ba, and Cr | NP |
| 31 | Phase II and IRA | None | 6 VOCs and 17 SVOCs | 4 VOCs | NP |
| 32 | Phase II | 2 VOCs, Ba, Cd, Cr, Pb, and Hg | 2 VOCs, Pb, and Hg | 4 VOCs and 2 SVOCs | NP |
| 34 | Phase II | 4 VOCs, 2 SVOCs, Ba, Cd, Pb, and Hg | 1 VOC, Ba, Cd, Cr, and Pb | 3 VOCs | NP |
| 37 | Phase I | 1 VOC and Hg | 2 VOCs and Hg | 4 VOCs | NP |
| | | | | | (4 VOCs, Ba, Cd, Cr, Pb, Hg, and Se) ^c |

^aPhase II RFI was not required at this time. The Phase II RFI will be conducted upon closure of the Red Cloud Range, Hotel Area.

^bPer the GEPD-approved SAP, subsurface soil was not collected because subsurface soil sampling in an EOD area requires approval by the Secretary of the Army.²

^cResults from sediment within the NGTC Equalization Basin.

^dSediment was collected; however, the oil/water separator does not discharge to the drainage ditch.

BEHP = Bis(2-ethylhexyl)phthalate.

NA = Not applicable.

NC = Not collected based on field screening results or because no medium (i.e., surface water) was available during the RFI.

ND = Not detected.

NP = No pathway exists.

SVOC = Semivolatile organic compound.

VOC = Volatile organic compound.

Table ES-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs

| SWMU | CMCOPCs | | | HHCOPCs | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|-------------------|--|--|---------------------|-----------------|------------------------|------------------------|-----------------|--------------------------|-----------------------------|--------------------|------------------------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 2 | 2 pest., Ar, Ca, and Hg | None | As and Cr | None | None | None | None | 4,4'-DDE, Cd, Cr, and Pb | 1 VOC, Pb, and Hg | None | None | HHBRA and SPRE |
| 3 | None | As | As | None | 1 pest. and Hg | 1 SVOC, As, Cr, and Pb | As | Pb and Cr | 2 pest., Ba, Cd, Pb, and Hg | 1 SVOC, Ba, and Pb | 2 VOCs, As, Ba, and Se | HHBRA and SPRE |
| 9 ^a | NA ^a | NA ^a | NA ^a | NC ^a | NC ^a | NP | NP | NA ^a | NC ^a | NP | NP | |
| 10 | NA ^b | NA ^b | NA ^b | NC | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | |
| 11 | NA ^b | NA ^b | NA ^b | NC | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NP | NP | |
| 12A | Ar, Cd, Cr, Pb, Ag, 1 SVOC, and 2 exp. | None | As and Pb | As | BEHP | Hg | None | 1 SVOC, Cd, Cr, and Pb | BEHP | Pb and Hg | Ba | HHBRA and SPRE |
| 14 | 1 VOC | NA | None | None | None | NP | NP | None | Pb, Hg, and 1 VOC | NP | NP | |
| 17 | None | None | None | None | 1 VOC | None | None | None | 1 VOC and Pb | None | None | HHBRA |
| 18 | Cr and Hg | (1 VOC, 1 SVOC, Ar, Ba, Cd, Cr, Hg, and Se) ^f | None | None | 3 VOCs and Pb | 1 SVOC | As | Pb | 4 VOCs, Ba, and Pb | Ba and BEHP | None | HHBRA and SPRE |
| 19 | 2 pest. | NA | None | None | BEHP, 2 pest., and As | NP | NP | Cd, Pb, and 1 pest. | BEHP, 5 pest., Ba, and Hg | NP | NP | HHBRA and SPRE |
| 24B | 1 VOC, 3 SVOCs, and Pb | NA | 4 SVOCs, As, and Pb | None | 1 VOC, 9 SVOCs, and Hg | NP | NP | NP | Hg and 9 SVOCs | NP | NP | |
| 27A (Bldg. 1339A) | None | NA | None | None | BEHP | NP | NP | None | 1 VOC and BEHP | NP | NP | |
| 27A (Bldg. 1339B) | None | NA | None | None | Benzene | NP | NP | Pb | Xylenes | NP | NP | |
| 27A (Bldg. 1322) | None | NA | None | None | Acetone | NA | NA | Pb | None | NP | NP | HHBRA |

Note: Footnotes appear on page ES-11.

Table ES-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs (continued)

| SWMU | CMCOPCs | | | HHCOPCs | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|---------------------|---------|----------|--------------|-----------------|--------------------|---------------|----------|--------------|--------------------|---------------|----------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 27B | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27C | None | NP | None | None | None | NP | NP | None | 2 VOCs | NP | NP | |
| 27D | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27E (Bldg. 1628) | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27E (Bldg. 1720) | None | NP | NA | None | None | NP | NP | NA | 1 SVOC | NP | NP | |
| 27F (NW Bldg. 1340) | None | NP | NA | None | 4 VOCs and 4 SVOCs | NP | NP | NA | 2 VOCs and 4 SVOCs | NP | NP | |
| 27F (NE Bldg. 1340) | 1 VOC | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27G | None | NP | NA | None | 1 SVOC | NP | NP | NA | None | NP | NP | HHBRA |
| 27H (Bldg. 1071) | 2 SVOCs | NP | NC | 1 SVOC | 1 VOC and 7 SVOCs | NP | NP | NC | 8 SVOCs | NP | NP | |
| 27H (Bldg. 1056) | None | NP | NC | None | 3 SVOCs | NP | NP | NC | 2 SVOCs | NP | NP | |
| 27I (Block 9900) | None | None | NC | None | None | NC | None | NC | None | NC | Pb | |
| 27I (Block 10300) | None | NA | NC | None | None | Pb | None | NC | None | Pb | None | |
| 27J (Bldg. 10535) | None | NP | None | None | None | NP | NP | None | 1 VOC and 1 SVOC | NP | NP | |
| 27J (Bldg. 10531) | None | NP | None | NC | 1 SVOC | NP | NP | None | 2 SVOCs | NP | NP | |
| 27K | None | NP | NA | None | None | NP | NP | NA | None | NP | NP | |
| 27L (Block 10200) | None | None | None | None | 4 VOCs and 2 SVOCs | Acetone | None | None | 2 VOCs and 1 SVOC | None | None | |
| 27M (Block 10100) | 1 VOC | None | None | None | 1 VOC | NC | None | Pb | None | NC | Pb | |
| 27N (Block 9800) | None | None | NA | None | None | NC | 1 SVOC | NA | None | NC | None | HHBRA |

Note: Footnotes appear on page ES-11.

Table ES-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs (continued)

| SWMU | CMCOPCs | | HHCOPCs | | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|------------------|------------------|---------------------------|--------------|-----------------|------------------------|---------------|----------|----------------|------------------------|---------------|----------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 27O (Block 9700) | None | None | None | NA | None | None | None | Pb | 1 SVOC | None | Pb | |
| 27P (Block 9500) | None | None | None | None | None | NC | None | None | None | NC | Pb | |
| 27Q (Block 9400) | None | None | None | NA | None | NC | None | Pb | None | NC | Pb | |
| 27R | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27S | None | NP | NA | None | None | NP | NP | NA | None | NP | NP | |
| 27T | None | Cd | 1 SVOC | None | None | NA | 4 SVOCs | None | 1 SVOC | NA | Cd | HHBRA |
| 27U | None | NP | None | None | Benzene | NP | NP | Pb | None | NP | NP | |
| 27V | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 29 | 7 VOCs | NP | None | None | 1 VOC, 2 SVOCs, and As | NP | NP | None | 1 VOC, 2 SVOCs, and Ba | NP | NP | HHBRA |
| 31 | 1 VOC and 1 SVOC | NP | None | None | Acetone | NP | NP | None | Xylenes | NP | NP | HHBRA |
| 32 | 1 VOC | NP | None | None | Acetone | NP | NP | Cd, Pb, and Cr | 1 VOC and 1 SVOC | NP | NP | HHBRA |
| 34 | 2 VOCs | NP | None | None | Acetone | NP | NP | Cd and Pb | 1 VOC | NP | NP | HHBRA |
| 37 | 1 VOC | 1 VOC ^b and Cd | None | None | Benzene | NP | NP | None | Xylenes | NP | NA | |

^aPhase II RFI was not required at this time. The Phase II RFI will be conducted upon closure of the Red Cloud Range, Hotel Area.

^bWith the concurrence of GEPA, fate and transport analysis and human health and ecological preliminary risk assessments were deemed unnecessary. SRCs were determined solely on comparison to background criteria (see Table ES-1).

^cResults from sediment within the NGTC Equalization Basin.

BEHP = Bis(2-ethylhexyl)phthalate.

HHBRA = Human health baseline risk assessment.

NA = Not applicable.

NC = Sample not collected based on field screening results or because no medium (i.e., surface water) was available during the RFI.

NP = No pathway exists.

SVOC = Semivolatile organic compound.

VOC = Volatile organic compound.

Table ES-3. SWMU-specific Recommendations

| SWMU | Recommendation |
|---------------------------|---|
| 2 | CAP |
| 3 | CAP |
| 9 | CAP |
| 10 | CAP |
| 11 | CAP |
| 12A | Long-term compliance monitoring and CAP |
| 14 | NFA |
| 17 | NFA |
| 18 | Long-term monitoring and CAP |
| 19 | NFA |
| 24B | Phase II RFI |
| 27A (Building 1339A) | NFA |
| 27A (Building 1339B) | NFA |
| 27A (Building 1322) | NFA |
| 27B | NFA |
| 27C | NFA |
| 27D | NFA |
| 27E (Building 1628) | NFA |
| 27E (Building 1720) | NFA |
| 27F (NW Building 1340) | Phase II RFI |
| 27F (NE Building 1340) | NFA |
| 27G | NFA |
| 27H (Building 1071) | Phase II RFI |

| SWMU | Recommendation |
|-------------------------|----------------|
| 27H (Building 1056) | Phase II RFI |
| 27I (Block 9900) | NFA |
| 27I (Block 10300) | NFA |
| 27J (Building 10535) | NFA |
| 27J (Building 10531) | Phase II RFI |
| 27K | NFA |
| 27L (Block 10200) | Phase II RFI |
| 27M (Block 10100) | NFA |
| 27N (Block 9800) | NFA |
| 27O (Block 9700) | NFA |
| 27P (Block 9500) | NFA |
| 27Q (Block 9400) | NFA |
| 27R | NFA |
| 27S | NFA |
| 27T | Phase II RFI |
| 27U | NFA |
| 27V | NFA |
| 29 | CAP |
| 31 | NFA |
| 34 | NFA |
| 32 | NFA |
| 37 | NFA |

1.0 INTRODUCTION

This report summarizes the results of the Resource Conservation and Recovery Act (RCRA) Facility Investigation (RFI) for 16 Solid Waste Management Units (SWMUs), at Fort Stewart, Georgia. The 16 SWMUs include: Camp Oliver Landfill, SWMU 2; Tactical Air Command (TAC)-X Landfill, SWMU 3; Inactive Explosive Ordnance Disposal (EOD) Area in Red Cloud Range, Hotel Area, SWMU 9; Inactive EOD Area North of Garrison Area, SWMU 10; Inactive EOD Area Located Approximately Three Miles Northeast of Garrison Area, SWMU 11; Active EOD Containing Open Detonation (OD) Unit and Open Burn (OB) Unit, SWMU 12A; Old Fire Training Area, SWMU 14; Defense Reutilization and Marketing Organization (DRMO) Hazardous Waste Storage Area, SWMU 17; Industrial Wastewater Treatment Plant (IWTP), SWMU 18; Old Sludge Drying Beds, SWMU 19; Old Radiator Shop/Paint Booth, SWMU 24B; Motorpools, SWMUs 27A through 27V; Evans Army Heliport Petroleum, Oil, and Lubricants (POL) Storage Facility, SWMU 29; Directorate of Engineering and Housing (DEH) Asphalt Tanks, SWMU 31; Supply Diesel Tank, SWMU 32; DEH Equipment Wash Rack, SWMU 34; and National Guard Training Center (NGTC) Equalization Basin, SWMU 37. Four of the 16 sites—Old Sludge Drying Beds, SWMU 19; Old Radiator Shop/Paint Booth, SWMU 24B; Motorpools, SWMUs 27A through 27V; and NGTC Equalization Basin, SWMU 37—had not been previously investigated and were investigated as Phase I RFIs. This report has been prepared by Science Applications International Corporation (SAIC) for the U.S. Army (Army) Corps of Engineers (USACE)—Savannah District under Contract DACA21-95-D-0022, Delivery Order No. 0009. The RFI was conducted in accordance with USACE Guidance EM 200-1-3.

1.1 OBJECTIVES AND SCOPE OF THE INVESTIGATION

The specific objectives of the Phase I and Phase II RFIs for the 16 SWMUs at Fort Stewart, Georgia, as defined in the Phase II RFI Sampling and Analysis Plan (SAP) (SAIC 1997) [approved by the Georgia Environmental Protection Division (GEPD) in October 1997] are listed below.

Phase I RFI

- Determine if contamination of the environment has occurred.
- Determine whether contaminants, if present, constitute a threat to human health or the environment.
- Determine the need for future action and/or no further action (NFA).

Phase II RFI

- Determine the horizontal and vertical extent of contamination.
- Determine whether contaminants present a threat to human health or the environment.
- Determine the need for future action and/or NFA.
- Gather data necessary to support a Corrective Action Plan (CAP), if warranted.

The information provided in this report is based upon data collected previously during the Phase I RFI (if available) and data collected as part of the Phase II field sampling and analysis. At some of the sites, the

Phase II sampling program incorporated an observational approach to sampling, as defined in the Phase II RFI SAP (SAIC 1997). This observational approach used field screening techniques to determine the horizontal and vertical extent of contamination at the SWMU and to identify suitable locations for installation of permanent monitoring wells. The scope of the fieldwork for the Phase I and II sites included the activities listed below.

Phase I Sites

- Collection of direct-push soil samples using a push probe.
- Collection of direct-push groundwater samples using a push probe.
- Collection of surface water and sediment samples at SWMUs where surface water and/or sediment was available.
- Surveying of the positions of all sample locations.

Phase II Sites

- Collection of direct-push soil samples using push probes.
- Collection of direct-push groundwater samples using push probes, including vertical-profile probes.
- Installation of permanent groundwater monitoring wells both upgradient and downgradient of the site.
- Groundwater sampling at existing monitoring wells (if available) and sampling of newly installed wells around the SWMUs.
- Collection of surface water and sediment samples at SWMUs where surface water and/or sediment was available.
- Surveying of the positions of all sample locations.

1.2 REPORT ORGANIZATION

This Phase II RFI Report consists of three volumes: 12 chapters of text in Volume I, 7 appendices in Volume II, and a final appendix in Volume III. Chapter 1.0 describes the purpose of this investigation, summarizes the scope of work performed, and presents the organization of the report. General information is presented in Chapters 2.0 through 8.0. Chapter 2.0 describes the Fort Stewart Military Reservation (FSMR) Installation and discusses the history of the FSMR and FSMR regulator history. Chapter 3.0 presents the regional setting of the FSMR, including the demographics, topography, regional geology and hydrogeology, surface drainage, soils, and ecology. Chapter 4.0 summarizes the investigation activities and methodologies used in completing the Phase II RFI fieldwork. Chapter 5.0 describes the results of the background interpretation for surface soil, subsurface soil, groundwater, surface water, and sediment and their relationship to each site. Chapter 6.0 identifies general considerations affecting contaminant fate and transport. Chapter 7.0 presents the general methodology for the human health preliminary risk evaluation (HHPRE), and Chapter 8.0 presents the general methodology for the ecological preliminary risk evaluation (EPRE).

SWMU-specific information corresponding to Chapters 2.0 through 8.0 is presented in Chapters 9.0 and 10.0, including site-specific conclusions on nature and extent of contaminants, fate and transport, HHPRE, and EPRE. Chapter 9.0, identified by a gray tab, designates in sequential order the SWMUs that are recommended for NFA because contaminant levels in soil, groundwater, sediment, and surface water are below reference background criteria or the sites do not pose a risk to human health and the environment based on human health and ecological risk assessments. Chapter 10.0, identified by a blue tab, designates in sequential order the SWMUs that are recommended for additional investigation or a CAP. Secondary tabs are used to separate the individual SWMUs behind the gray or blue tab. Chapter 11.0 presents conclusions and recommendations identifying the SWMUs being recommended for NFA or SWMUs that indicate risk to human health or environment and are recommended for additional investigation or a CAP. References are presented in Chapter 12.0.

Volume II of this report contains nine appendices. Appendix A contains the direct-push technology and boring logs. Appendix B contains monitoring well construction diagrams. Appendix C is the Quality Control Summary Report. Appendix D provides a comparison of metal data from the Phase I and Phase II RFIs. Appendix E contains the geotechnical laboratory test results. Appendix F is the background data summary. Appendix G contains the chain-of-custody forms.

Volume III of this report contains five appendices. Appendix H provides the analytical data results. In addition, the analytical data are provided in electronic format (i.e., on CDs). Appendix I presents the methodology for the human health baseline risk assessment. Appendix J contains the toxicity profiles for contaminants of potential concern (COPCs). Appendix K presents Fate and Transport Input Data and Model Description. Appendix L presents the revised response to GEPD comments received on the final Phase II RFI Report for 16 SWMUs submitted in February 1999 and the meeting minutes for the comment response meeting with GEPD held on September 14, 1999.

2.0 SITE HISTORY AND CONTAMINANTS

2.1 INSTALLATION DESCRIPTION

Fort Stewart (then known as Camp Stewart) was established in June 1940 as an antiaircraft artillery training center. Between January and September 1945, the Installation operated as a prisoner-of-war camp. The Installation was deactivated in September 1945. In August 1950 Fort Stewart was reactivated to train antiaircraft artillery units for the Korean Conflict. The training mission was expanded to include armor training in 1953. Fort Stewart was designated a permanent Army installation in 1956 and became a flight training center in 1966. Aviation training at the Fort Stewart facilities was phased out in 1973. In January 1974 the 1st Battalion, 75th Infantry was activated at Fort Stewart. Fort Stewart then became a training and maneuver area, providing tank, field artillery, helicopter gunnery, and small arms training for regular Army and National Guard units. The 24th Infantry Division, which was reflagged as the 3d Infantry Division in May 1966, was permanently stationed at Fort Stewart in 1975. These activities comprise the Installation's primary mission today.

The FSMR is located in portions of Liberty, Bryan, Long, Tattnall, and Evans counties, Georgia, approximately 40 miles west-southwest of Savannah, Georgia (Figures 2-1 and 2-2). The cantonment, or garrison area, of the FSMR is located within the Liberty County portion of the FSMR, on the southern boundary of the reservation. Hinesville, Georgia, is the nearest city to the garrison area and is located immediately south of the reservation's boundary.

2.2 SITE LOCATION AND HISTORY

The Phase I RFIs were conducted in response to a RCRA Facility Assessment (RFA) submitted to the GEPD in June 1990. The RFA listed 24 SWMUs requiring some type of RFI action (Geraghty and Miller 1992). The objective of the Phase I investigations at Fort Stewart was to determine if a release to the environment had occurred from any of the 24 identified SWMUs and to decide if the site had the potential for a release to the environment. After the Phase I RFI Report, 11 oil/water separators (OWSs) were added under SWMU 27 for a total of 32 OWSs distributed over 29 sites. Each site represents a distinct geographic area requiring an investigation. Sixteen of the original 24 SWMUs presented in the Phase I RFI Report were recommended for a Phase II RFI, or initial investigation (Phase I), and are presented in this Phase II RFI Report. In addition, a recently identified site, the NGTC Equalization Basin (SWMU 37), is also presented in this report. Thus, 17 SWMUs were investigated: 13 Phase II RFIs (SWMUs 2, 3, 9, 10, 11, 12, 14, 17, 18, 29, 31, 32, and 34) and four Phase I RFIs (SWMUs 19, 24B, 27A through 27V, and 37). Site descriptions for the SWMUs are presented under the respective SWMU sections in Chapters 9.0 and 10.0.

Seven of the 16 SWMUs are located outside the garrison area (i.e., SWMUs 2, 3, 9, 10, 11, 12A, and 29) and are presented in Figure 2-3. The remaining SWMUs (i.e., SWMUs 14, 17, 18, 19, 24B, 27A through 27V, 31, 32, 34, and 37) are located within the garrison area and are presented in Figure 2-4. Table 2-1 is a summary table identifying the SWMU designation from the Phase I Report, the SWMU designation from the Hazardous Waste Permit (HW-045), the level of investigation (Phase I or Phase II), and the site investigation requirements as determined by geographic location.

Table 2-1. List of Solid Waste Management Units Included in This Report

| SWMU Number | SWMU Name on Hazardous Waste Permit HW-045 | SWMU Name Phase I RFI Designation | Number of Site Investigations under Each SWMU | Investigation Type |
|-------------|--|--|---|-----------------------|
| 2 | Camp Oliver Landfill | Camp Oliver Landfill, SWMU 2 (FST-002) | 1 | Phase II |
| 3 | TAC-X Landfill | TAC-X Landfill, SWMU 3 (FST-003) | 1 | Phase II |
| 9 | Inactive EOD Area in Red Cloud Range, Hotel Area | Inactive EOD Area, SWMU 9 (FST-009) | " | " |
| 10 | Inactive EOD Area North of Garrison Area | Inactive EOD Area, SWMU 10 (FST-010) | 1 | Phase II |
| 11 | Inactive EOD Area Located Approximately Three Miles Northeast of Garrison Area | Inactive EOD Area, SWMU 11 (FST-011) | 1 | Phase II |
| 12A | Active EOD Containing Open Detonation Unit and Open Burn Unit | Active EOD Area, SWMU 12 (FST-012) | 1 | Phase II |
| 12B | Open Detonation (OD) Unit | Not identified in Phase I RFI | 1 ^b | Phase II ^b |
| 12C | Open Burn (OB) Unit | Not identified in Phase I RFI | 1 ^b | Phase II ^b |
| 14 | Old Fire Training Area | Old Fire Training Area, SWMU 14 (FST-014) | 1 | Phase II |
| 17 | DRMO Hazardous Waste Storage Area | Hazardous Waste Storage Area, SWMU 17 (FST-017) | 1 | Phase II |
| 18 | Industrial Wastewater Treatment Plant | Industrial Wastewater Treatment Plant, SWMU 18 (FST-018) | 1 | Phase II |
| 19 | Old Sludge Drying Beds | Old Sludge Drying Beds, SWMU 19 (FST-019) | 1 | Phase I |
| 24B | Old Radiator Shop/Paint Booth | Paint Booth, SWMU 24B (FST-24B) | 1 | Phase I |
| 27A | 3d Squadron, 7th Cavalry Motorpool and Four Associated Oil/Water Separators | 2d Squadron, 4th Cavalry | 3 | Phase I |
| 27B | 1st BN, 3d ADA Motorpool and Associated Oil/Water Separator | 1st BN, 5th ADA | 1 | Phase I |
| 27C | 92d ECB (H) Motorpool and associated Oil/Water Separator | 92d ECB (H) | 1 | Phase I |
| 27D | 26th SPT BN Motorpool and Associated Two Oil/Water Separators | 224th SPT BN | 1 | Phase I |
| 27E | 703d SPT BN (Main) Motorpool and Associated Two Oil/Water Separators | 724th SPT BN (Main) | 2 | Phase I |
| 27F | 3d Engineer Brigade Motorpool and Associated Two Oil/Water Separators | 24th Inf. Div Engineer Brigade | 2 | Phase I |
| 27G | DISCOM Motorpool and Associated Oil/Water Separator | DISCOM | 1 | Phase I |

Note: Footnotes appear on page 2-8.

Table 2-1. List of Solid Waste Management Units Included in This Report (continued)

| SWMU Number | SWMU Name on Hazardous Waste Permit HW-045 | SWMU Name Phase I RFI Designation | Number of Site Investigations under Each SWMU | Investigation Type |
|--|--|---|---|--------------------|
| 27H | DOL Maintenance Motorpool and Associated Two Oil/Water Separators | DOL Maintenance | 2 | Phase I |
| 27I | NGTC Block 9900, 10300 Motorpool and Associated Two Oil/Water Separators | NGTC Block 9900, 10300 | 2 | Phase I |
| 27J | GANG MATES Motorpool and Associated Two Oil/Water Separators | GANG MATES | 2 | Phase I |
| 27K | 3d BN, 69th Armor Motorpool Wash Rack and Oil/Water Separator | 3d BN, 69th Armor | 1 | Phase I |
| 27L | NGTC Block 10200 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27M | NGTC Block 10100 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27N | NGTC Block 9800 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27O | NGTC Block 9700 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27P | NGTC Block 9500 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27Q | NGTC Block 9400 Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27R | 396 Transportation Company Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27S | Two 103d MI BN Wash Racks and Associated Two Oil/Water Separators | Not identified in Phase I RFI | 1 | Phase I |
| 27T | 293 MP Company Wash Rack and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27U | Two Wright Army Airfield Wash Racks and Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 27V | Auto Craft Center Oil/Water Separator | Not identified in Phase I RFI | 1 | Phase I |
| 29 | Evans Army Heliport POL Storage Facility | Evans Army Heliport POL Storage Facility, SWMU 29 (FST-029) | 1 | Phase II |
| 31 | DEH Asphalt Tanks | DEH Asphalt Tanks, SWMU 31 (FST-031) | 1 | Phase II |
| 32 | Supply Diesel Tank | Supply Diesel Tank, SWMU 32 (FST-032) | 1 | Phase II |
| 34 | DEH Equipment Wash Rack | DEH Equipment Wash Rack, SWMU 34 (FST-034) | 1 | Phase II |
| 37 | NGTC Equalization Basin | Not identified in Phase I RFI | 1 | Phase I |
| Total Number of Site Investigations ^c | | | 44 | |

^aBecause SWMU 9 is contained within an active range, the site investigation at SWMU 9 was deferred until the range is closed.

^bSWMUs 12B and 12C are contained within SWMU 12A.

^cTotal number includes 32 OWSs at 29 sites.

2.3 PREVIOUS INVESTIGATIONS

Results of previous investigations are presented under the respective SWMU sections in Chapters 9.0 and 10.0.

2.4 PRELIMINARY CONCEPTUAL SITE MODEL

Based upon a review of the history of operations at the SWMUs and the results of the Phase I RFI, representative preliminary Conceptual Site Models (CSMs) were developed identifying the potential migration pathways of contaminants. Discussions for the preliminary CSMs for each SWMU or groups of SWMUs are presented in the following sections. The final CSMs are presented under the respective SWMUs in Chapters 9.0 and 10.0.

2.4.1 Camp Oliver Landfill (SWMU 2) and TAC-X Landfill (SWMU 3)

Materials such as municipal/sanitary solid waste and construction and demolition debris were disposed of at the Camp Oliver Landfill and TAC-X Landfill using trench-and-fill landfill construction techniques.

The buried material and decomposition products associated with the landfills and the leachate generated from the infiltration/percolation of precipitation and the contact with fluctuating groundwater levels represent a potential source of contaminants at the former landfills. Leachate migration represents the most likely pathway for contaminant migration at the landfills. Potential contaminant pathways include surface runoff, migration of leachate from seeps to surface water, and migration of leachate to groundwater.

The characteristics of leachate are determined by the character of the buried material. Typically, leachate from municipal/sanitary landfills has a low pH (less than 6), significant negative oxidation-reduction (Redox) potential, and no dissolved oxygen (DO), which is indicative of reducing conditions. The leachate often has high concentrations of organic compounds, with volatile organic compounds (VOCs) being more prevalent in younger landfills (during active decomposition). The reducing conditions facilitate the dissolution of metals, which may be indicated by high total dissolved solids (TDS) concentrations and high specific conductance. The main COPCs at the former landfills are VOCs, semivolatile organic compounds (SVOCs), and metals.

2.4.2 Inactive EOD Areas (SWMUs 9, 10, and 11)

Explosive residuals, unexploded ordnance (UXO), and RCRA metals are the main concerns at the inactive EOD areas. The sites are characterized by small craters where ordnance was disposed of on the surface. The Phase I RFI indicated only one detection of explosive residuals in the soil across the three sites. Contamination at the surface from the disposal of ordnance represents the main potential migration pathway for contamination at the inactive EOD sites. The main COPCs are explosive constituents and RCRA metals. Surface water drainages are not present at SWMU 9 or SWMU 11; therefore, constituents are not likely to migrate to surface water. Surface drainage from SWMU 10 may potentially impact Taylors Creek, which is east of SWMU 10.

2.4.3 Active EOD Area (SWMU 12A)

The Active EOD Area (SWMU 12A) contains OD and OB treatment areas. Contamination at the surface from the disposal of ordnance represents the main potential migration pathway for contamination at the active EOD site. Explosive residuals, UXO, organic contaminants (SVOCs only), and RCRA metals are the COPCs at the active EOD areas. Surface drainage at SWMU 12A flows to Canoochee Creek and the swampy area located northeast and southeast, respectively, of the site.

3.0 PHYSICAL CHARACTERISTICS OF THE SITE

3.1 DEMOGRAPHICS

The FSMR is located in portions of Liberty, Bryan, Long, Tattnall, and Evans counties, Georgia, approximately 40 miles west-southwest of Savannah (Figure 2-1). The cantonment, or garrison area, of the FSMR is located within Liberty County, Georgia (Figure 2-2). Liberty County occupies 328,768 acres and had a total population of 52,745 in 1990. Forty-one percent of the county population lives in Hinesville, the largest city in Liberty County. The total population of Fort Stewart in 1990 was 13,774, 55 percent of which were employed by the Armed Forces. Forty-one percent of the Fort Stewart population lived in group quarters, while the remaining population lived in households (U.S. Department of Commerce 1990).

3.2 TOPOGRAPHY

The FSMR occupies a low-lying, flat region on the coastal plain of Georgia. Surface elevations range from approximately 20 feet to 100 feet above mean sea level (amsl) within the FSMR and generally decrease from northwest to southeast across the reservation. The topography is dominated by terraces dissected by surface water drainages. The terraces are remnants of sea level fluctuations. The four terraces present within the FSMR are the Wicomico, Penholoway, Talbot, and Pamlico (Metcalf and Eddy 1996a).

3.3 SURFACE DRAINAGE

The principal surface water body accepting drainage from the FSMR is the Canoochee River, which joins the Ogeechee River (part of the northwestern boundary of the reservation). Canoochee Creek is a tributary of the Canoochee River that drains much of the western portion of the FSMR.

The site-specific surface drainage characteristics of each SWMU are described in their respective sections in Chapters 9.0 and 10.0. Generally, the surface water drainage (if present) at the SWMUs consists of shallow swales or drainage ditches not directly linked to specific surface water bodies (i.e., creeks or rivers). The drainage swales/ditches (if present) typically drain to a low or depression area where the surface water percolates into the soil.

As discussed in Chapter 2.0, SWMUs outside the garrison area (SWMUs 2, 3, 9, 10, 11, 12A, and 29) are separate, individual remote locations and have site-specific surface water features and drainage characteristics. Only four of these sites, SWMUs 2, 3, 10, and 12A have surface water bodies in their immediate proximity.

At the majority of the SWMUs within the garrison area (SWMUs 14, 17, 18, 24B, 27A-V, 31, 32, and 34), the surface water drainage (if present) consists of shallow swales or drainage ditches to collect and direct surface water runoff from the area in which the SWMU happens to be located (e.g., motorpool) and are not specific to the SWMU in question. Therefore, the surface water/sediment is potentially influenced by many other sources.

The potential surface water drainage from SWMUs contained within the garrison primarily drains to two areas of the garrison area. SWMUs 14, 17, 18, 24B, 27A-H, 27K, 27K, 31, and 34 drain to a swales or ditches that ultimately may drain to Mill and/or Taylors Creek. However, the primary surface water drainage at these SWMUs is percolation into surface/subsurface soil.

SWMUs 27I and 27L-27T (all OWSs) are located along Troupe Avenue, and potential surface runoff from these SWMUs may migrate to the drainage ditch/swale along Troupe Avenue that ultimately drains to a low area southeast of Troupe Avenue. The effluent from eight of the SWMUs along Troupe Avenue (27I, Block 9900; 27I, Block 10300; 27L, Block 10200; 27M, Block 10100; 27N, Block 9800; 27O; 27P; and 27Q) discharges directly into the drainage ditch/swale along Troupe Avenue and adjacent to the sites. SWMUs 27I, 27J, and 37 are also located in the southeastern portion of the garrison area; however, the surface drainage does not influence the Troupe Avenue drainage swale(s). Potential runoff from SWMUs 27I and 27J is allowed to percolate into adjacent surface/subsurface soil. Potential runoff from SWMU 37 may migrate to a drainage ditch downgradient of the SWMU; however, surface runoff primarily is allowed to percolate into the surface/subsurface soil.

3.4 REGIONAL GEOLOGY

The FSMR is located within the coastal plain physiographic province. This province is typified by southeastward-dipping strata that increase in thickness from 0 foot at the fall line (located approximately 350 miles inland from the Atlantic coast) to approximately 4,200 feet at the coast. State geologic records describe a probable petroleum exploration well (the No. 1 Jelks-Rogers) located in the region as having encountered crystalline basement rocks at a depth of 4,254 feet below ground surface (bgs). This well provided the most complete record for Cretaceous, Tertiary, and Quaternary sedimentary strata. Figure 3-1 presents a geologic column for the Tertiary and Quaternary sections in the Fort Stewart area.

The Cretaceous section is approximately 1,970 feet in thickness and is dominated by clastics. The Tertiary section is approximately 2,170 feet in thickness and is dominated by limestone, with a 175-foot-thick cap of dark green phosphatic clay. This clay is regionally extensive and is known as the Hawthorn Group. The interval from approximately 110 feet to the surface is Quaternary in age and composed primarily of sand with interbeds of clay or silt. This section is undifferentiated (Metcalf and Eddy 1996a).

State geologic records contain information regarding a well drilled in October 1942, 1.8 miles north of Flemington at Liberty Field of Camp Stewart (now known as Fort Stewart). This well is believed to have been an artesian well located approximately 0.25 mile north of the runway at Wright Army Airfield within the FSMR. The log for this well describes a 410-foot section, the lowermost 110 feet of which consisted predominantly of limestone above which 245 feet of dark green phosphatic clay typical of the Hawthorn Group were encountered. The uppermost 55-foot interval was Quaternary-age interbedded sands and clays. The top 15 feet of these sediments were described as sandy clay (Metcalf and Eddy 1996a).

3.5 SOILS

The major soil types in the area of FSMR range from well-drained, nearly pure sand to poorly drained mixtures of loam, sand, and clay. The soils lack natural strength and are vulnerable to erosion if stripped of vegetation. Boring logs showing the types of soils encountered under the Phase II RFI at the SWMUs, including soil from screening probes, groundwater screening probes, and monitoring well boreholes, are given in Appendix A.

Geotechnical analyses were conducted on one bulk sample and one Shelby tube sample taken from the monitoring well boreholes. The bulk samples were analyzed for grain-size distribution [in accordance with American Society for Testing and Materials (ASTM) D422], moisture content (ASTM D2216), and Atterberg limits (ASTM D4318). In addition, the Shelby tube samples were analyzed for specific gravity (ASTM D854), porosity (EM 1110-2-1906), and permeability (ASTM D5084). Results of the geotechnical analyses are

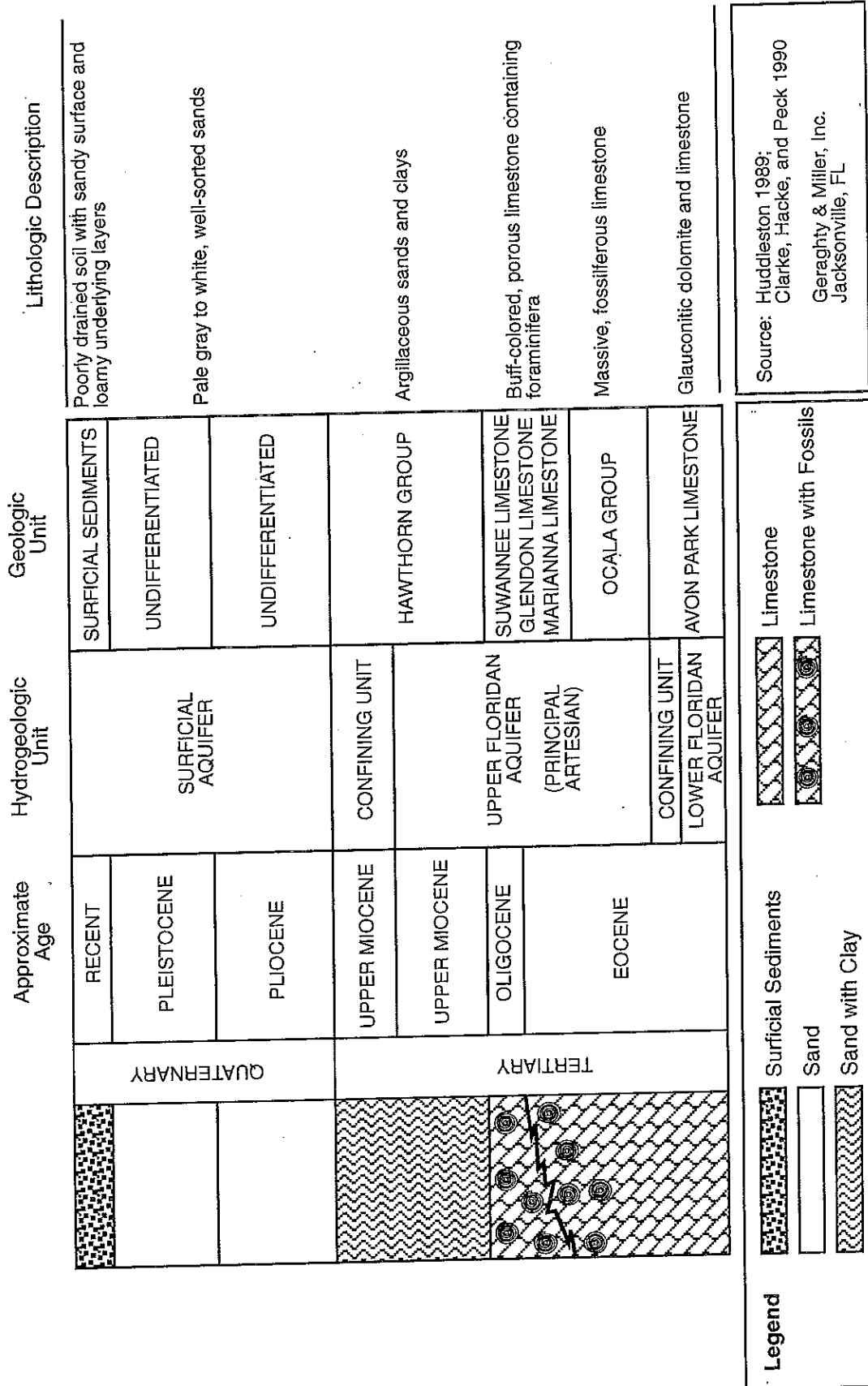


Figure 3-1. Approximate Geological Column for the FSMR Showing the Principal Hydrostratigraphy

summarized in each SWMU section as presented in Chapters 9.0 and 10.0. The geotechnical laboratory data sheets and chains of custody are included in Appendix E.

3.6 HYDROGEOLOGY

The hydrogeology in the vicinity of the FSMR is dominated by two aquifers, referred to as the Principal Artesian and the surficial aquifer, that are separated by a confining unit (Figure 3-1).

The Principal Artesian aquifer is the lowermost hydrologic unit; is regionally extensive from South Carolina through Georgia, Alabama, and most of Florida; and is regionally known as the Floridan Aquifer. This aquifer is subdivided into upper and lower hydrogeologic units. The upper hydrogeologic unit is composed primarily of Miocene-age argillaceous sands and clays and Oligocene- to Eocene-age limestones (including the Ocala Group and the Suwannee Limestone, where present) at the top. The upper hydrogeologic unit ranges in thickness from 200 feet to 260 feet and is most productive where it is thickest and where secondary permeability is most developed. The lower hydrologic unit is comprised of the Eocene-age Avon Park Limestone at the base. The transmissivity of the aquifer in the Savannah area ranges from about 28,000 square feet/day to 33,000 square feet/day (Krause and Randolph 1989). Groundwater from this aquifer is primarily used for drinking water (Arora 1984). Thirteen groundwater production wells are used for potable water supply on the FSMR, and one additional production well is available for use in fire protection.

The confining layer for the Principal Artesian aquifer is the phosphatic clays of the upper Hawthorn Group. These sediments are regionally extensive and range from 60 feet to 80 feet in thickness at the FSMR. There are minor occurrences of aquifer material within the Hawthorn Group; however, they have limited utilization (Miller 1990).

The uppermost hydrologic unit is the surficial aquifer, which consists of widely varying amounts of sand, silt, and clay ranging from 35 feet to 150 feet in thickness. This aquifer is primarily used for domestic lawn and agricultural irrigation, with wells typically yielding 2 gallons to 180 gallons per minute.

Water levels were measured from temporary piezometers at the SWMUs. The resulting data were used to determine the placement of permanent monitoring wells around respective SWMUs.

Water levels were also measured in newly installed and existing monitoring wells around the SWMUs. The potentiometric surface based on the water levels in the monitoring wells is presented for the respective SWMUs in Chapters 9.0 and 10.0.

3.7 ECOLOGY

Approximately 7.8 square miles of the 436.8 square miles at the FSMR comprise the garrison area. The remainder is used for ranges and training areas (approximately 11 percent) or held as non-use areas.

Eighty-four percent of the land is forested (approximately 367.2 square miles). Sixty-six percent of the forest area is pine, with the major species including the slash, loblolly, and longleaf pines. Thirty-four percent of the forest is composed of river bottomlands and swamps whose major species include tupelo, other gum trees, water oak, and bald cypress trees. The open range and training areas comprise 11 percent of the Installation and consist of grasses, shrubs, and scrub tree (oak) growth.

Aquatic habitats on the FSMR include a number of natural or man-made ponds and lakes, the Canoochee River, Canoochee Creek and its tributaries, and a number of bottomland swamps and pools. The Ogeechee River borders the installation along its northeast boundary. Organic detritus content is high, and dark coloring of the water is not unusual. Dense growths of aquatic vegetation are also typical, especially during the summer months.

Both terrestrial and aquatic fauna are abundant in the unimproved areas of the FSMR. Major game species found on the installation include white-tailed deer, feral hog, wild turkey, rabbit, squirrel, and bobwhite in addition to numerous other mammal, bird, reptile, and amphibian species (Environmental Science and Engineering 1982). Dominant fish include bluegill, largemouth bass, crappie, sunfish, channel catfish, minnows, and shiners. Three federally listed threatened or endangered species reside at the FSMR: the American bald eagle, Eastern indigo snake, and red-cockaded woodpecker.

3.8 METEOROLOGY

Fort Stewart has a humid, subtropical climate with long, hot summers. Average temperatures range from 50°F in the winter to 80°F in the summer. Average annual precipitation is 48 inches, with slightly over half falling from June through September. Prolonged drought is rare in the area, but severe local storms (tornadoes and hurricanes) do occur. Under normal conditions, wind speeds rarely exceed 5 knots, but gusty winds of more than 25 knots may occur during summer thunderstorms (Geraghty and Miller 1992).

4.0 SUMMARY OF INVESTIGATION ACTIVITIES

4.1 SAMPLING METHODOLOGIES

This section describes the general RFI procedures and methodology followed at the 16 SWMUs from January 13 through June 12, 1998. In addition, sampling to address specific GEPC comments on the final Phase II RFI for 16 SWMUs was performed during field mobilizations conducted July 10, 1999, through November 4, 1999. The sampling methodologies and types of testing for physical and chemical characterization of the site are also described. Specific numbers of samples and types of physical and chemical characterization as well as locations are presented under the respective SWMU discussions in Chapters 9.0 and 10.0. The sampling strategy included installation and collection of direct-push soil and groundwater samples, installation and sampling of monitoring wells, groundwater sampling of existing monitoring wells, and surface water and sediment sampling. Table 4-1 presents a summary table of the media sampled, with the number of samples collected per medium for each SWMU investigated. Table 4-2 presents a summary of the types of analyses performed on each medium at each site.

4.1.1 Soil Sampling

Soil sampling was conducted using two methods: (1) direct-push methods and (2) hollow-stem augers during installation of soil borings and monitoring wells.

4.1.1.1 Direct-push soil sampling

Direct-push soil probes were completed within or around the perimeter of a SWMU to evaluate the potential extent of contamination. The direct-push soil probes were selected for the following reasons:

- to determine the extent of contamination in surface and subsurface soil at the SWMU and
- to minimize generation of investigation-derived waste (IDW).

The direct-push soil samples were taken using a 4-foot macro sampler by pushing the sampler from the ground surface down to the water table in continuous 4-foot intervals. Each 4-foot sample was split into two 2-foot samples. The headspace of the soil samples was field-tested for VOCs using a photoionization detector (PID). The sample from each boring having the highest detected organic vapor concentration in the headspace gas was then sent off-site for quantitative laboratory analysis for VOCs with rapid (24-hour) turnaround. If no VOCs were detected in the headspace gas, then the sample from the 2-foot interval directly above the water table was sent for analysis because COPCs at the SWMUs would tend to be distributed at the soil/water interface. These samples served to confirm the presence or absence of contamination using quantitative data. Results of the laboratory VOC analyses are presented in Chapters 9.0 and 10.0. Logs for the direct-push soil probes showing headspace readings and depths sampled are included in Appendix A.

Samples designated for possible VOC laboratory analysis were collected first from each interval using a stainless steel spoon and placed into laboratory sample containers. A portion of the remaining sample was then placed into containers designated for headspace analyses. The remaining portion of the sample was used for field lithologic description.

Table 4-1. Summary of Media and Investigation Technologies for the SWMUs

| Area | Media or Investigation Technology | | | | | | | | | | | | |
|--|-----------------------------------|------------------|--------------------|---|---|----------------------|---------------------------|---------------------------------|---|----------------------------|-------------------------|------------------------|----------------------|
| | Surface Water Samples | Sediment Samples | Wastewater Samples | Direct-push Groundwater Screening Samples | Direct-push Vertical-profile Samples ^a | Surface Soil Samples | Soil Borings ^b | Direct-push Groundwater Samples | 3/4-Inch-diameter Permanent Monitoring Point ^c | Monitoring Wells Installed | Subsurface Soil Samples | Existing Wells Sampled | Geotechnical Samples |
| SWMU 2 Camp Oliver Landfill | 2 | 2 | | | 1 | 11 | 6 | 3 | | 3 | 8 | 4 | 2 |
| SWMU 3 TAC-X Landfill | 2 | 3 | | 3 | 1 | 10 | 7 | 3 | | 4 | 7 | 4 | 4 |
| SWMU 9 Inactive EOD Area ^d | | | | | | | | | | | | | |
| SWMU 10 Inactive EOD Area | 2 | 2 | | 8 | | 3 | | | | | | | |
| SWMU 11 Inactive EOD Area | | | | 8 | | 3 | | | | | | | |
| SWMU 12A Active EOD Area | 5 | 5 | | | | 11 | | | | 3 | 3 | 7 | 3 |
| SWMU 14 Old Fire Training Area | | | | | 1 | 9 | 10 | | | 4 | 11 | 4 | 4 |
| SWMU 17 DRMO Hazardous Waste Storage | 2 | 2 | | 6 | 1 | 6 | | | | 3 | 3 | | 3 |
| SWMU 18 IWTP | 8 ^e | 8 ^e | 2 | 25 | 14 | 6 | 10 | | | 9 | 12 | | 10 |
| SWMU 19 Old Sludge Drying Beds | | | | | | 8 | | 8 | 1 | | 8 | | |

Note: Footnotes appear on page 4-3.

Table 4-1. Summary of Media and Investigation Technologies for the SWMUs (continued)

| Area | Media or Investigation Technology | | | | | | | | | | | | |
|--|-----------------------------------|------------------|--------------------|---|---|----------------------|---------------------------|---------------------------------|---|----------------------------|-------------------------|------------------------|----------------------|
| | Surface Water Samples | Sediment Samples | Wastewater Samples | Direct-push Groundwater Screening Samples | Direct-push Vertical-profile Samples ^a | Surface Soil Samples | Soil Borings ^b | Direct-push Groundwater Samples | 3/4-Inch-diameter Permanent Monitoring Point ^c | Monitoring Wells Installed | Subsurface Soil Samples | Existing Wells Sampled | Geotechnical Samples |
| SWMU 24B Old Radiators Shop/Paint Booth | | | | | | 5 | | 6 | | | 4 | | |
| SWMUs 27A-27V Motorpools | 3 | 12 | | 119 | | | 5 | 119 | 13 | 5 ^d | 119 ^e | | |
| SWMU 29 Evans Heliport POL | | | | 18 | 2 | 15 | 9 | | | 9 | 12 | | 6 |
| SWMU 31 DEH Asphalt Tanks | | | | 9 | 1 | 6 | 3 | | | 3 | 7 | | 3 |
| SWMU 32 Supply Diesel Tank | | | | 4 | 1 | 6 | 3 | | | 3 | 6 | | 3 |
| SWMU 34 DEH Wash Rack | | | | 4 | 1 | 6 | 3 | | | 3 | 6 | | 3 |
| SWMU 37 NGTC Equalization Basin | | 4 ^e | 4 ^e | 6 | 1 | 4 | | | | | 5 | | 1 |

^aThe temporary piezometer location at the site indicating the highest concentration from screening was the initial vertical profile. If required, subsequent vertical profiles were installed based on field screening results and groundwater flow direction. Therefore, three groundwater samples were taken for each vertical profile (i.e., 10 feet to 30 feet bgs, 30 feet to 40 feet bgs, and 40 feet to 50 feet bgs) unless site-specific subsurface conditions (i.e., refusal) prevented sampling.

^bIncludes borings for installation of monitoring wells.

^cWith the concurrence of GEPD, 3/4-inch-diameter permanent monitoring points were installed, and the groundwater was resampled to confirm direct-push groundwater results.

^dBecause SWMU 9 is located within an active range, the Phase II RFI was deferred until closure of the range.

^eSediment and surface water samples included samples collected from their respective equalization basins.

^fWith the concurrence of GEPD, a 2-inch PVC monitoring well was installed, and the groundwater was resampled to confirm direct-push groundwater results.

^gContinuous soil sampling was performed. The soil sample indicating the highest organic concentration from screening was sent off-site for analysis.

Table 4-2. Summary of Media Sampled and Laboratory Analyses Performed for the SWMUs

| SWMU Number | SWMU Name | VOCs | SVOCs | TPH-Recoverable ^a | Pesticides/PCBs | RCRA Metals | Lead | Explosive Constituents | Geotechnical ^{b,c} |
|------------------------|--|------|-------|------------------------------|-----------------|----------------|------|------------------------|-----------------------------|
| <i>Surface Soil</i> | | | | | | | | | |
| SWMU 2 | Camp Oliver Landfill | X | X | | X | X | | | |
| SWMU 3 | TAC-X Landfill | X | X | | X | X | | | |
| SWMU 9 ^d | Inactive EOD Area ^d | | | | | | | | |
| SWMU 10 | Inactive EOD Area | | | | | X | | X | |
| SWMU 11 | Inactive EOD Area | | | | | X | | X | |
| SWMU 12A | Active EOD Area | | X | | | X | | X | |
| SWMU 14 | Old Fire Training Area | X | X | | | X | | | |
| SWMU 17 | DRMO Hazardous Waste Storage Area | X | X | | | X | | | |
| SWMU 18 | Industrial Wastewater Treatment Plant | X | X | | | X | | | |
| SWMU 19 | Old Sludge Drying Bed | X | X | | X | X | | | |
| SWMU 24B | Old Radiator Shop/Paint Booth | X | X | | | X | | | |
| SWMUs 27A-27V | Motorpools | X | X | | | X ^e | X | | |
| SWMU 29 | Evans Army Heliport POL Storage Facility | X | X | | | X | | | |
| SWMU 31 | DEH Asphalt Tanks | X | X | | | X | | | |
| SWMU 32 | Supply Diesel Tank | X | X | | | X | | | |
| SWMU 34 | DEH Equipment Wash Rack | X | X | | | X | | | |
| SWMU 37 | NGTC Equalization Basin | X | X | | | X | | | |
| <i>Subsurface Soil</i> | | | | | | | | | |
| SWMU 2 | Camp Oliver Landfill | X | X | | X | X | | | X |
| SWMU 3 | TAC-X Landfill | X | X | | X | X | | | X |
| SWMU 9 ^d | Inactive EOD Area ^d | | | | | | | | |
| SWMU 10 | Inactive EOD Area | | | | | | | | |
| SWMU 11 | Inactive EOD Area | | | | | | | | |
| SWMU 12A | Active EOD Area | | X | | | X | | X | X |
| SWMU 14 | Old Fire Training Area | X | X | | | X | | | X |
| SWMU 17 | DRMO Hazardous Waste Storage Area | X | X | | | X | | | X |
| SWMU 18 | Industrial Wastewater Treatment Plant | X | X | | | X | | | X |
| SWMU 19 | Old Sludge Drying Bed | X | X | | X | X | | | |

Note: Footnotes appear on page 4-7.

Table 4-2. Summary of Media Sampled and Laboratory Analyses Performed for the SWMUs (continued)

| SWMU Number | SWMU Name | VOCs | SVOCs | TPH-Recoverable ^a | Pesticides/PCBs | RCRA Metals | Lead | Explosive Constituents | Geotechnical ^{b,c} |
|--------------------------------|--|------|-------|------------------------------|-----------------|----------------|------|------------------------|-----------------------------|
| SWMU 24B | Old Radiator Shop/Paint Booth | X | X | | | X | | | |
| SWMUs 27A-27V | Motorpools | X | X | | | X ^e | X | | |
| SWMU 29 | Evans Army Heliport POL Storage Facility | X | X | | | X | | | X |
| SWMU 31 | DEH Asphalt Tanks | X | X | | | X | | | X |
| SWMU 32 | Supply Diesel Tank | X | X | | | X | | | X |
| SWMU 34 | DEH Equipment Wash Rack | X | X | | | X | | | X |
| SWMU 37 | NGTC Equalization Basin | X | X | | | X | | | X |
| <i>Sediment</i> | | | | | | | | | |
| SWMU 2 | Camp Oliver Landfill | X | X | | X | X | | | |
| SWMU 3 | TAC-X Landfill | X | X | | X | X | | | |
| SWMU 9 ^d | Inactive EOD Area ^d | | | | | | | | |
| SWMU 10 | Inactive EOD Area | | | | | X | | X | |
| SWMU 11 | Inactive EOD Area | | | | | | | | |
| SWMU 12A | Active EOD Area | | X | | | X | | X | |
| SWMU 14 | Old Fire Training Area | | | | | | | | |
| SWMU 17 | DRMO Hazardous Waste Storage Area | X | X | | | X | | | |
| SWMU 18 | Industrial Wastewater Treatment Plant | X | X | | | X | | | |
| SWMU 19 | Old Sludge Drying Bed | | | | | | | | |
| SWMU 24B | Old Radiator Shop/Paint Booth | | | | | | | | |
| SWMUs 27A-27V | Motorpools | X | X | | | X ^e | X | | |
| SWMU 29 | Evans Army Heliport POL Storage Facility | | | | | | | | |
| SWMU 31 | DEH Asphalt Tanks | | | | | | | | |
| SWMU 32 | Supply Diesel Tank | | | | | | | | |
| SWMU 34 | DEH Equipment Wash Rack | | | | | | | | |
| SWMU 37 | NGTC Equalization Basin | X | X | | | X | | | |
| <i>Groundwater^f</i> | | | | | | | | | |
| SWMU 2 | Camp Oliver Landfill | X | X | | X | X | | | |
| SWMU 3 | TAC-X Landfill | X | X | | X | X | | | |

Note: Footnotes appear on page 4-7.

Table 4-2. Summary of Media Sampled and Laboratory Analyses Performed for the SWMUs (continued)

| SWMU Number | SWMU Name | VOCs | SVOCs | TPH-Recoverable ^a | Pesticides/PCBs | RCRA Metals | Lead | Explosive Constituents | Geotechnical ^{b,c} |
|----------------------|--|------|-------|------------------------------|-----------------|----------------|------|------------------------|-----------------------------|
| SWMU 9 ^d | Inactive EOD Area ^d | | | | | | | | |
| SWMU 10 | Inactive EOD Area | | | | | | | X | |
| SWMU 11 | Inactive EOD Area | | | | | | | X | |
| SWMU 12A | Active EOD Area | | X | | | X ^g | | X | |
| SWMU 14 | Old Fire Training Area | X | X | | | X | | | |
| SWMU 17 | DRMO Hazardous Waste Storage Area | X | X | X | | X | | | |
| SWMU 18 | Industrial Wastewater Treatment Plant | X | X | X | | X | | | |
| SWMU 19 | Old Sludge Drying Bed | X | X | | X | X | | | |
| SWMU 24B | Old Radiator Shop/Paint Booth | X | X | | | X | | | |
| SWMUs 27A-27V | Motorpools | X | X | | | X ^e | X | | |
| SWMU 29 | Evans Army Heliport POL Storage Facility | X | X | X | | X | | | |
| SWMU 31 | DEH Asphalt Tanks | X | X | X | | X | | | |
| SWMU 32 | Supply Diesel Tank | X | X | X | | X | | | |
| SWMU 34 | DEH Equipment Wash Rack | X | X | X | | X | | | |
| SWMU 37 | NGTC Equalization Basin | X | | X | | | | | |
| <i>Surface Water</i> | | | | | | | | | |
| SWMU 2 | Camp Oliver Landfill | X | X | | X | X | | | |
| SWMU 3 | TAC-X Landfill | X | X | | X | X | | | |
| SWMU 9 ^d | Inactive EOD Area ^d | | | | | | | | |
| SWMU 10 | Inactive EOD Area | | | | | X | | X | |
| SWMU 11 | Inactive EOD Area | | | | | | | | |
| SWMU 12A | Active EOD Area | | X | | | X | | X | |
| SWMU 14 | Old Fire Training Area | | | | | | | | |
| SWMU 17 | DRMO Hazardous Waste Storage Area | X | X | | | X | | | |
| SWMU 18 | Industrial Wastewater Treatment Plant | X | X | | | X | | | |
| SWMU 19 | Old Sludge Drying Bed | | | | | | | | |
| SWMU 24B | Old Radiator Shop/Paint Booth | | | | | | | | |

Note: Footnotes appear on page 4-7.

Table 4-2. Summary of Media Sampled and Laboratory Analyses Performed for the SWMUs (continued)

| SWMU Number | SWMU Name | VOCs | SVOCs | TPH-Recoverable ^a | Pesticides/PCBs | RCRA Metals | Lead | Explosive Constituents | Geotechnical ^{b,c} |
|----------------------------|--|------|-------|------------------------------|-----------------|----------------|------|------------------------|-----------------------------|
| SWMUs 27A-27V ^h | Motorpools | X | X | | | X ^e | X | | |
| SWMU 29 | Evans Army Heliport POL Storage Facility | | | | | | | | |
| SWMU 31 | DEH Asphalt Tanks | | | | | | | | |
| SWMU 32 | Supply Diesel Tank | | | | | | | | |
| SWMU 34 | DEH Equipment Wash Rack | | | | | | | | |
| SWMU 37 | NGTC Equalization Basin | X | X | | | X | | | |

^aPerformed on groundwater screening samples only.

^bOne soil sample from each monitoring well was analyzed for grain-size distribution, Atterberg limits, and moisture content.

^cOne soil sample from each SWMU was analyzed for permeability, porosity, specific gravity, and total organic carbon.

^dBecause SWMU 9 is located within an active range, the Phase II RFI was deferred until closure of the active range.

^eRCRA metals analysis was performed on media at two of the motorpools.

^fOne groundwater sample from each site was analyzed for ferric iron and sulfate to be used in the future to perform a Risk-based Corrective Action.

^gAdditional metals analyses were performed to meet Subpart X monitoring requirements.

^hSurface water was available at some sites only after rainfall events.

X = Analyzed for on all samples.

4.1.1.2 Permanent monitoring points

Permanent monitoring points were installed at some SWMUs under Phase I investigation to confirm groundwater sample results obtained from direct-push groundwater sampling. The confirmation resampling and the installation of the permanent monitoring points at each location were approved by GEPD prior to the installation. The permanent monitoring points were installed using techniques similar to those described for direct-push groundwater sampling (see Section 4.1.2.1). A 3/4-inch-inside-diameter polyvinyl chloride (PVC) monitoring point with a 10-foot screened interval with a filter pack was installed to intersect the water table. The permanent monitoring point was installed approximately 1 foot away from the location to be resampled. Groundwater samples were collected using the same procedures as those described for direct-push groundwater sampling (see Section 4.1.2.1).

4.1.1.3 Soil sampling at soil boring and monitoring well locations

Soil samples were also taken during the drilling of boreholes for the installation of soil borings and monitoring wells using the hollow-stem auger drilling method. Auger-drilled soil boreholes were advanced using 4.25-inch-inside-diameter hollow-stem augers using either a CME-55 or Ingersoll-Rand A-300 drill rig. The borehole samples were collected to obtain the following:

- relatively undisturbed samples for geotechnical testing,

- lithographic descriptions of the soil profile at each monitoring well,
- background surface and subsurface soil samples for characterization, and
- surface and subsurface soil samples for characterizing the nature and extent of contaminants.

During the drilling of each soil borehole, soil samples were collected with a split-barrel sampler continuously over 5-foot intervals from the ground surface to the water table. The 5-foot core was split into two 2.5-foot sections. A portion of each 2.5-foot section was field-tested for VOC headspace gas using a PID. As with the direct-push soil samples, the borehole sample having the highest detected organic vapor concentration in the headspace gas was then sent off-site for quantitative laboratory analysis. If no VOCs were detected in the headspace gas, then the sample from the 2.5-foot interval directly above the soil/water interface was sent for analysis because contaminants typically tend to be distributed at the water table interface. In addition, the surface sample taken from a depth of 0 to 1 foot bgs was sent off-site for analysis for use in the HHPRE and EPRE; therefore, two soil samples were collected from each borehole for chemical analyses. Results of the chemical analyses are presented under the respective SWMUs in Chapters 9.0 and 10.0. Boring logs for the drilling of soil borings and monitoring wells are included in Appendix A.

Decontamination of drilling and downhole sampling equipment was accomplished in accordance with the procedures specified in the Phase II RFI SAP (SAIC 1997). These procedures included washing with water and phosphate-free detergent, rinsing alternately with water and isopropyl alcohol, air drying, and placing the equipment on clean plastic or wrapping it in plastic or aluminum foil to prevent cross-contamination.

At monitoring well boreholes, one soil sample from the screened interval in each borehole was analyzed for geotechnical parameters to support contaminant transport evaluation. Bulk soil samples were taken from all monitoring well boreholes. The soil was collected directly from the 5-foot split-barrel core and placed into containers. The samples were tested for moisture content, Atterberg limits, and grain-size distribution. At one well at each site, a relatively undisturbed sample was collected for geotechnical analysis using a thin-walled (Shelby) tube sampler. The Shelby tube sampler was inserted into the hollow-stem auger string and hydraulically pushed approximately 2 feet. The ends of the Shelby tube sampler were sealed with wax to preserve the moisture content in accordance with ASTM D1587-94, and the tubes were shipped to an off-site laboratory for analysis. The Shelby tube sample was tested for soil porosity, specific gravity, and permeability. Soil from these locations was also sampled for total organic carbon (TOC).

4.1.2 Groundwater Sampling

4.1.2.1 Direct-push groundwater sampling

The direct-push groundwater samples were taken for the following reasons:

- to delineate the extent of contamination in groundwater,
- to determine the most appropriate locations for monitoring wells based on contamination, and
- to estimate the approximate direction of groundwater flow to determine the most appropriate locations for downgradient monitoring wells.

A single grab sample of groundwater was obtained at the water table from the direct-push locations. Multiple grab samples of groundwater at varying depth intervals were collected at vertical-profile stations to measure

the vertical distribution of contamination. The locations of the direct-push groundwater probes are presented under each SWMU in Chapters 9.0 and 10.0.

The direct-push groundwater samples at the water table probe locations were taken using direct-push sampling techniques (Dietrich Power Punch devices mounted on a Mobil B-47 drilling rig or Geoprobe devices mounted on trucks). The sampling device used for shallow groundwater sampling had a 7/8-inch-inside-diameter screen/casing. The device was pushed down to between 3 feet and 5 feet below the level of the water table that was encountered, and a grab groundwater sample was retrieved at the water table using a peristaltic pump or stainless steel bailer. At the vertical-profile locations, multiple depths were sampled from the same hole using a dual wall system that prevented cross-contamination of the sample intervals to a maximum of 50 feet or when the Hawthorn layer was reached, whichever was encountered first. The Hawthorn layer is a confining unit that prevents downward migration of contaminants. The samples were then sent off-site for laboratory analyses for site-specific screening parameters with rapid (24-hour) turnaround. The site-specific screening parameter was typically VOCs; however, explosive constituents were analyzed at SWMUs where EOD disposal had historically occurred. Results of the site-specific screening analyses are presented under the respective SWMUs in Chapters 9.0 and 10.0.

To assist in estimating the direction of groundwater flow, water levels were measured in temporary piezometers that were set in the direct-push holes. Following installation of all temporary piezometers, each piezometer was surveyed for horizontal and vertical elevation. Unfortunately, field parameter measurements were not taken during groundwater screening sampling as proposed in the SAP because an insufficient amount of sample was available. The results of these measurements are presented under the respective SWMUs in Chapters 9.0 and 10.0.

4.1.2.2 Monitoring well installation and development

Monitoring wells were installed at SWMUs under Phase II investigation and with the concurrence of GEPD at some Phase I RFI sites (SWMUs 27F, Northeast of Building 1340; 27J, Building 10535; 27N, Block 9800; 27S; and 27U) to confirm or deny potential contamination at selected Geoprobe locations. The monitoring well boreholes were drilled using the hollow-stem-auger drilling method. Auger-drilled monitoring well boreholes were advanced using 4.25-inch inside diameter hollow-stem augers using a CME-55 or Ingersoll Rand A-300 drill rig. The resulting borehole was approximately 9.5 inches in diameter. The total depth of each borehole was dictated by the depth at which the water table was encountered. Boreholes were drilled to allow 7 feet of screen (total screen length 10 feet) to be placed beneath the water table.

The wells were constructed of 2-inch-diameter Schedule 40 PVC with flush-threaded couplings. Well screens were constructed of factory-slotted pipe in 10-foot-long sections. Slot size, determined from the sieve analysis results from the direct-push soil probes and field sieve analyses, was 0.008 inch (No. 8 slot). No. 8 slot size was used for all monitoring well screens. Filter pack materials consisted of DSI Extra-fine Sand. A 2-foot-thick bentonite seal consisting of commercially available pellets was placed above the filter pack. After placement of the pellets, a small volume of approved water was used to hydrate the pellets for a minimum of one hour. Grout composed of Type I Portland cement, 3 pounds of dry powdered bentonite per 94-pound sack of dry cement, and a maximum of 7 gallons of approved water per sack of cement was placed in the remaining annulus starting at just above the bentonite seal using a rigid-side discharge tremie pipe. The annulus was grouted to the ground surface for aboveground completions and to approximately 2 feet bgs for flush-mounted completions. The site requirements determined whether a monitoring well was completed aboveground or flush. For aboveground completions a protective iron/steel casing was installed as the grout was being placed within the well annulus. The protective casing was set approximately 5 feet below and extended approximately 3 feet above the ground. A protective concrete pad with measurements of 3 feet by 3 feet square and with a thickness of 4 inches was poured around the exterior of the protective casing, and four steel posts were

installed to protect the aboveground completions. Well construction diagrams are presented in Appendix B. Well construction details are summarized under each respective SWMU in Chapters 9.0 and 10.0.

The wells were installed such that the screened interval bisected the water table so that any nonaqueous-phase liquid floating on the water table surface could be detected in the well.

The wells were developed throughout the field investigation a minimum of 48 hours after installation. Well development was accomplished using a downhole positive-displacement pump. A surge block was used to agitate and mobilize particulates around the well screen by rapidly surging the surge block up and down. Well development continued until field parameters stabilized, turbidity was less than 10 nephelometric turbidity units (NTUs), sediment within the well was less than 0.1 foot, a minimum of five times the standing water volume in the well had been removed or a maximum of 12 hours had passed, and five times the volume of any water added during completion had been removed. Water quality parameters (pH, conductivity, temperature, DO, Eh, and turbidity) were measured during well development to verify that all field parameters had reached equilibrium. At some wells turbidity remained higher than 10 NTUs after 12 hours of development, but all other development criteria had been met. Well development is summarized under each respective SWMU in Chapters 9.0 and 10.0.

4.1.2.3 Monitoring well sampling

Groundwater sampling was conducted at least 14 days after well development. Monitoring wells were sampled using low-flow micropurging techniques to minimize the volume of purge water, the disturbance of the aquifer, and, thereby, the turbidity of the sample. The wells were checked for free product, and water levels were measured prior to purging and sampling. Field parameters (pH, conductivity, temperature, DO, Eh, and turbidity) were monitored during micropurging. The purge rate was adjusted, as necessary, to avoid purging any well to dryness and to equal the recharge of the aquifer. Purging was considered complete when the field parameters stabilized within plus or minus 10 percent after a minimum of three readings at 5-minute intervals and a turbidity of less than 10 NTUs was achieved. Purging times varied, requiring from 8 hours to 12 hours of purging to attain a turbidity of less than 10 NTUs. At some wells turbidity did not reduce to below 10 NTUs after 6 consecutive hours of purging. In such cases a field decision was made to increase the target turbidity to 25 NTUs, and up to 4 additional hours of purging were performed to achieve 25 NTUs. After a total of 10 hours of purging, if the turbidity still had not dropped below 25 NTUs, filtered and unfiltered metal samples were taken. A list of wells that did not achieve 25 NTUs even after 10 hours of purging and the associated results of field parameter measurements recorded at the end of purging in each well are presented under the respective SWMUs in Chapters 9.0 and 10.0.

Sampling of each monitoring well began immediately after completion of purging, using the same micropurging pump. Groundwater samples were transferred directly into laboratory sample containers, with the portion designated for volatile organic analysis taken first. Filtered groundwater samples were collected by attaching a 0.45-micron filter to the end of the low-flow pump sampling line. A field test kit was used to measure ferric iron at selected monitoring wells. Ferric iron analyses were randomly distributed across existing and new wells. Groundwater samples were then sent off-site for laboratory chemical analysis. Total iron, ferric iron, and sulfate analyses were performed at selected locations to support contaminant fate and transport modeling and potential remedial alternative development.

4.1.3 Surface Water and Sediment Sampling

Surface water and sediment samples were collected at upstream (background) and downstream sampling locations where surface water was a potential migration pathway. Surface water samples were collected first, and then field measurements were taken for pH, specific conductance, temperature, DO, and turbidity. Sediment samples were collected using stainless steel scoops or an Eckman dredge depending on the depth of the water. Samples were then sent off-site for laboratory chemical analysis.

4.1.4 Wastewater and Sediment (Sludge) Sampling

Surface water and sediment (sludge) were collected in the equalization basin of the IWTP (SWMU 18) and the NGTC Equalization Basin (SWMU 37). In addition, influent and effluent wastewater was collected at SWMU 18.

4.1.5 Investigation-derived Waste Management

IDW was managed in accordance with the procedures specified in the Phase II RFI SAP (SAIC 1997). All IDWs were determined to be nonhazardous materials. Solid wastes were disposed of by transporting the material to the Fort Stewart Sanitary Landfill for use as daily cover. Liquid wastes were disposed of at the Fort Stewart IWTP.

4.2 DATA QUALITY ASSESSMENT

Multiple activities were performed to achieve the desired data quality in this project. Data quality objectives (DQOs) were established to guide the implementation of the field sampling and laboratory analysis. A quality assurance (QA) program was established to standardize procedures and to document activities. Upon receipt by the project team, data were subjected to a verification and validation review that identified and qualified problems related to the analysis. The review steps contributed to a final Quality Control Summary Report (QCSR), Appendix C, which ensures that data used in the investigation met the criteria and were employed appropriately.

The QA program established requirements for both field and laboratory quality control (QC) procedures. In general, field QC duplicates and QA split samples were required for each environmental sample matrix collected at sites being investigated; VOC trip blanks were to accompany each cooler containing water samples for VOC determinations; equipment rinsate blanks were collected; and analytical laboratory QC duplicates, matrix spikes, laboratory control samples, and method blanks were required for every 20 samples or fewer of each matrix and analyte. The primary goal of the QA program was to ensure that the quality of results for all environmental measurements was appropriate for the data's intended use. To this end, a Quality Assurance Project Plan (QAPP) and standardized field procedures were compiled to guide the investigation. Through the process of readiness review, training, equipment calibration, QC implementation, and detailed documentation, the project successfully accomplished the goals set by the QA program.

Project data quality determines its usability. The evaluation was based on the interpretation of laboratory QC measures, field QC measures, and the project DQOs. Daily Quality Control Reports and other field-generated documents such as sampling logs, boring logs, daily health and safety summaries, daily safety inspections, equipment calibration and maintenance logs, and sample management logs were peer-reviewed on-site. Analytical data generated for this project have been subjected to a process of data verification, validation, and review. The project implemented the use of data validation checklists to facilitate laboratory data validation. These checklists were completed by the project-designated validation staff and were reviewed by the project laboratory coordinator.

The checklists are sent with the analytical laboratory data deliverable (if requested) to the USACE-Savannah District, and a copy is also maintained at SAIC's Central Records.

A total of 1,250 environmental soil, sediment, groundwater, surface water, and field QC samples were collected, with approximately 94,000 discrete analyses (i.e., analytes) being obtained, reviewed, and integrated into the assessment. (These totals do not include field measurements and field descriptions.) The project produced acceptable results for more than 98 percent of the sample analyses performed and successfully collected all required investigation samples. In response to GEPD comments, analyte values reported below the project reporting levels were qualified as nondetects based on poor accuracy and precision (SAIC 1999a). Estimated concentrations for compounds such as methylene chloride, acetone, and toluene were considered to represent systemic project blank contamination, with values within three times the reporting levels being qualified as estimated and nondetect ("UJ") at the level observed. The rationale and acceptance for this data interpretation are discussed in the QCSR (see Appendix C) and in the comment response table and the minutes of the September 14, 1999, meeting with GEPD (see Appendix L).

A subset of field duplicate analysis compared groundwater filtered and total metals values. This comparison is presented in Appendix C. Filtered and total metal values show good agreement from analytical data obtained from established monitoring well locations, which were sampled using low-flow sampling techniques; however, Geoprobe filtered and unfiltered metals analysis showed very poor agreement (Phase I sites SWMUs 19, 24B, and 27A through 27V). Geoprobe total metal values (i.e., lead) provided positive results greater than five times the reporting level, while filtered values were consistently below the reporting level. The higher total metal values were apparently the result of particulate material in the sample due to the Geoprobe sampling methodology and the resulting high sample turbidity. The dissolved metal concentration was used as the concentration representative of the groundwater metal characteristics at Phase I sites (SWMUs 19, 24B, and 27A through 27V). Appendix D presents a discussion comparing the Phase I groundwater metal concentrations and Phase II groundwater metal concentrations using low-flow techniques and justification for using the dissolved metal concentration as representative of the groundwater characteristics at Phase I sites.

The overall quality of this SWMU investigation information met or exceeded the established project objectives. Through proper implementation of the project data verification, validation, and assessment process, project information was determined to be acceptable for use. Data, as presented, have been qualified as usable but as estimated, when necessary. Data produced for this study demonstrates that they can withstand scientific scrutiny, are appropriate for their intended purposes, are technically defensible, and are of known and acceptable sensitivity, precision, and accuracy. A more detailed data quality assessment may be found in Appendix C.

5.0 CONTAMINANT NATURE AND EXTENT

This section summarizes the results of the chemical laboratory analyses for soil, groundwater, surface water, and sediment background samples collected during the Phase II RFI for 16 SWMUs. The nature and extent interpretation for each SWMU is presented under the respective SWMU section included in Chapters 9.0 and 10.0.

5.1 BACKGROUND DATA ANALYSIS

The reference background criteria for the 16 SWMUs have been developed based on data from background samples collected from SWMUs across the FSMR. In general, reference background samples were collected in each medium at locations upgradient or upstream of each site so as to be representative of naturally occurring conditions at sites under Phase II investigation. Upgradient or upstream samples were not collected at sites under a Phase I RFI (SWMUs 19, 24B, 27, and 37). Soil data collected during the Phase I RFI and from SWMUs that received an RFI but were not investigated under this investigation were included in the background data set if they were determined to be upgradient of the site and of sufficient quality to be representative of natural background conditions at the FSMR. Phase I RFI SWMUs whose soil data were included in the background data set were the Burn Pits (SWMUs 4A, 4B, 4D, 4E, and 4F) and the Active EOD Area (SWMU 12A). Other RFI SWMUs whose soil data were included in the background set were the South Central Landfill (SWMU 1), the Burn Pits (SWMUs 4A, 4B, 4D, 4E, and 4F), the 724th Tanker Purging Station (SWMU 26), and Wright Army Airfield (SWMU 35). A summary of the sample stations by SWMU and the source of the data (Phase I or Phase II RFI) are presented in Table 5-1. The locations of all reference background samples are shown in Appendix F on Figures F-1 and F-2.

U.S. Environmental Protection Agency (EPA) Region IV methodology (EPA 1995) was used as guidance for the development of the background data set for the screening of metals data. In cases in which enough samples (e.g., more than 20) to define background are collected, a background upper tolerance level can be calculated. In cases in which fewer samples (e.g., less than 20) are collected to define background, background can be calculated as two times the mean background concentration (EPA 1995). Given that fewer than 20 background samples were collected for the FSMR, the latter method was used for calculating reference background concentrations for metals.

Appendix F presents both the summary of background data and the two-times-mean background concentrations. Given the limited number of reference background samples, the mean concentrations for soils in the eastern United States are also presented for comparative purposes only.

The concentrations of organics detected in background samples were not used to calculate reference background criteria because all organic compounds are considered to be potentially man-made. Organic compounds were not screened against background. All detected organic compounds are considered to be site-related contaminants (SRCs). The following sections discuss the background data analysis for each medium.

5.2. SURFACE SOIL

Surface soil samples were taken from ground surface to a depth of 1 foot or 2 feet, depending on the amount of recovery from the sampling device. Thirteen surface soil samples were used to develop the background

Table 5-1. Background Media Summary

| SWMU Number | SWMU Name on Hazardous Waste Permit HW-045 | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment |
|-------------|---|---------------------------------|---------------------------------|-------------------|---------------|----------|
| 1 | South Central Landfill | SC-M17 ^a | SC-M17 ^a | MW10 ^a | SW/SED1 | SW/SED1 |
| 2 | Camp Oliver Landfill | MW5 ^b | MW5 ^b | MW5 ^b | SWS2 | SWS2 |
| 3 | TAC-X Landfill | MW5 ^b | MW5 ^b | MW5 ^b | NB | NB |
| 4A | Burn Pit A | | MW1 ^c (Phase I) | MW1 ^d | NP | NP |
| 4B | Burn Pit B | | MW3 ^c (Phase I) | MW3 ^d | NP | NP |
| 4C | Burn Pit C | MW7 ^d | MW7 ^d | MW7 ^d | NP | NP |
| 4D | Burn Pit D | | MW2 ^c (Phase I) | MW2 ^d | NP | NP |
| 4E | Burn Pit E | | MW3 ^c (Phase I) | MW3 ^d | NP | NP |
| 4F | Burn Pit F | | MW1 ^c (Phase I) | MW1 ^d | NP | NP |
| 10 | Inactive EOD Area | | | | SWS1 | SWS1 |
| 12A | Active EOD Containing Open Detonation Unit and Open Burn Unit | MW1 ^e | MW1 ^e | MW1 ^b | SWS1 | SWS1 |
| 14 | Old Fire Training Area | | | MW8 ^b | NP | NP |
| 17 | DRMO Hazardous Waste Storage Area | MW1 ^b | MW1 ^b | MW1 ^b | SWS1 | SWS1 |
| 18 | Industrial Wastewater Treatment Plant | MW1 ^b | MW1 ^b | MW1 ^b | SWS1 | SWS1 |
| 26 | Former 724th Tanker Purging Station | MW1 ^b | MW1 ^b | MW1 ^b | SWS1 | SWS1 |
| 29 | Evans Army Heliport POL Storage Facility | MW5 ^b | MW5 ^b | MW5 ^b | NP | NP |
| 31 | DEH Asphalt Tanks | MW1 ^b | MW1 ^b | MW1 ^b | NP | NP |
| 32 | Supply Diesel Tank | MW1 ^b | MW1 ^b | MW1 ^b | NP | NP |
| 34 | DEH Equipment Wash Rack | MW1 ^b | MW1 ^b | MW1 ^b | NP | NP |
| 35 | Wright Army Airfield Bulk Fuel System | HA-05 ^f (Phase I) | HA-05 ^f (Phase I) | | NA | NA |

^aSAIC 1998a.^bInformation provided in Chapters 9.0 and 10.0 under the respective Phase II RFI SWMUs.^cRust Environment and Infrastructure 1996.^dSAIC 1998b.^eRadian 1997.^fMetcalf and Eddy 1996b.

NA = Not applicable.

NB = No site-specific background sample available; results from Former 724th Tanker Purging Station (SWMU 26) used.

NP = No surface water/sediment pathway exists.

Bold indicates background groundwater sample collected from the same borehole as sample for soil (i.e., monitoring well was constructed in the borehole).

data set (Table 5-1). The reference background surface soil concentration was calculated as two times the mean concentration of these 13 locations. Phase I data from SWMU 12A and SWMU 35 were determined to be of sufficient quality to be included in the background data set. If a chemical was not detected at a site, then one-half the detection limit was used as the concentration when calculating the reference mean background concentration. The sample results included in the data set are presented in Table F-1, Appendix F. The reference background concentration for organics is also presented in Table F-1; however, all detected organic compounds are considered to be SRCs because organic constituents are considered to be potentially man-made. Inorganics were considered SRCs if their concentrations were above the calculated reference background

concentration, and organics were considered SRCs if they were detected, no matter what the concentrations. SRCs identified in the nature and extent of contamination section for each site were carried through to the HHPRE and EPRE and are presented in Chapters 9.0 and 10.0.

5.3 SUBSURFACE SOIL

Subsurface soil samples were taken from the interval between a depth of 2 feet bgs and the water table. Eighteen subsurface soil samples were used in the development of the subsurface soil background data set (Table 5-1). Phase I data from SWMUs 4A, 4B, 4D, 4E, and 4F, 12A, and 35 were determined to be of sufficient quality to be included in the subsurface background set. The reference background subsurface soil concentration was calculated as two times the mean of the chemical detected at the 18 locations. If a chemical was not detected in a sample, then one-half the detection limit was used in calculating the mean background concentration. The sample results included in the background data set are presented in Table F-2, Appendix F. When identifying SRCs at each SWMU, inorganics were considered SRCs if their concentrations were above the calculated reference background concentration, and organics were considered SRCs if they were detected because organic constituents are considered potentially man-made. SRCs identified in the nature and extent of contamination section for each site were carried through to the HHPRE and EPRE and are presented in Chapters 9.0 and 10.0.

5.4 GROUNDWATER

Only groundwater samples collected using low-flow techniques (Phase II RFI) were used in the development of the groundwater background data set. Groundwater samples from 18 SWMUs were used to develop the groundwater background composite (Table 5-1). The reference background groundwater concentration was calculated as two times the mean of these 19 samples. If a chemical was not detected at a site, then one-half the detection limit was used in calculating the mean background concentration. The sample results included in the background data set are presented in Table F-3, Appendix F. In addition to RCRA metals, groundwater samples at the Active EOD Area (SWMU 12A) were also analyzed for metals corresponding to Appendix D of the Subpart X Permit. These additional metals were compared to metal concentrations from MW1 (the site background well) at SWMU 12A collected during compliance monitoring using non-low-flow techniques. The site reference background concentration for these additional metals was determined to be two times the average concentration at MW1.

In addition, as indicated in Chapter 4.0, with the concurrence of GEPD some monitoring wells were installed at the SWMUs under the Phase I RFI (SWMUs 27F, Northeast of Building 1340; 27J, Building 10535; 27N, Block 9800; 27S; and 27U) only to confirm or deny potential contamination at selected Geoprobe locations. Groundwater at these SWMUs was collected using direct-push technology (DPT) and had a high level of turbidity from particulates in the groundwater. Total and dissolved metal samples were collected at the SWMUs under the Phase I RFI to differentiate the metals concentrations associated with the particulates/fines in the groundwater. The dissolved metals concentration was determined to be representative of the groundwater characteristics at the Phase I sites (SWMUs 19, 24B, and 27A through 27V) and was screened against reference groundwater background criteria (Appendix D).

Inorganics in groundwater were considered SRCs if their concentrations were above the calculated reference background concentration, and organics were considered SRCs if they were detected, no matter what the concentration, because organic constituents are considered to potentially be man-made. SRCs identified in the nature and extent of contamination section for each site were carried through to the HHPRE and EPRE and are presented in Chapters 9.0 and 10.0.

5.5 SURFACE WATER

Surface water background samples were collected during the Phase II RFI and applied to the SWMUs on a site-specific basis except at the TAC-X Landfill (SWMU 3) and motorpools (SWMUs 27A through 27V). At these sites, no upstream surface water was available, so the site-specific background for the Former 724th Tanker Purging Station (SWMU 26) was used. The SWMU 26 surface water sample was collected upstream in Mill Creek prior to the water's entering the FSMR. The locations by SWMU where site-specific background surface water samples were collected are presented in Table 5-1. No Phase I RFI data were used. Six locations had site-specific surface water samples collected (Table 5-1). The reference background surface water concentration was calculated as two times the mean of the data taken at the site-specific background location. If a chemical was not detected at a site, then one-half the detection limit was used as the mean background concentration. The sample results comprising the site-specific background data are presented in Table F-4, Appendix F. Inorganics were considered SRCs if their concentrations were above the site-specific reference background concentration, and organics were considered SRCs if they were detected, no matter what the concentration, because organic constituents are considered to potentially be man-made. SRCs identified in the nature and extent of contamination section for each site were carried through to the HHPRE and EPRE and are presented in Chapters 9.0 and 10.0.

5.6 SEDIMENT

Sediment background samples were collected during the Phase II RFI and applied to the SWMUs on a site-specific basis except at the TAC-X Landfill (SWMU 3) and motorpools (SWMUs 27A through 27V). At these sites no upstream sediment was available. The site-specific background for the Former 724th Tanker Purging Station (SWMU 26) was used for the TAC-X Landfill. The SWMU 26 background sediment sample was collected upstream in Mill Creek prior to the water's entering the FSMR. The reference background criteria for surface soil, as discussed in Section 5.2, was used for background screening of the sediment at the SWMU 27 sites because the SWMU 27 sediment samples were collected from dry ditch bottoms. The locations by SWMU where site-specific background sediment samples were collected are presented in Table 5-1. No Phase I RFI data were used. Six locations had site-specific sediment samples collected (Table 5-1). The reference background sediment concentration was calculated as two times the mean of the data taken at the site-specific background location. If a chemical was not detected at a site, then one-half the detection limit was used as the mean background concentration. The sample results comprising the site-specific background data are presented in Table F-5, Appendix F. Inorganics were considered SRCs if their concentrations were above the site-specific background concentration, and organics were considered SRCs if they were detected, no matter what the concentration, because organic constituents are considered to potentially be man-made. SRCs identified in the nature and extent of contamination section for each site were carried through to the HHPRE and EPRE and are presented in Chapters 9.0 and 10.0.

5.7 SITE-RELATED CONTAMINANTS

Inorganics for surface soil, subsurface soil, and groundwater were screened against the reference background criteria. Inorganics for surface water and sediment were screened against site-specific background values. As discussed in the preceding sections, all organics that were detected were considered to be potential SRCs because organic constituents are considered to potentially be man-made. Organic analytes that were detected at least once and those inorganic analytes for which at least one sample result exceeded background are considered SRCs. Only the SRCs were carried through for evaluation under fate and transport, HHPRE, and EPRE.

6.0 CONTAMINANT FATE AND TRANSPORT

6.1 INTRODUCTION

This chapter describes the potential migration pathways and mechanisms for transport of chemical substances found in surface and subsurface soils, sediment, surface water, and groundwater at the 16 SWMUs. Section 6.2 discusses the persistence, mobility, and other physical and chemical properties of the organics and metals found at the 16 SWMUs. Section 6.3 presents the components of a conceptual model developed for the SWMUs discussed in Chapters 9.0 and 10.0 and describes contaminant release mechanisms through the primary transport medium (groundwater). Section 6.4 discusses the fate and transport of the contaminants at the 16 SWMUs with respect to their leachability and natural attenuation.

6.2 PHYSICAL AND CHEMICAL PROPERTIES

The fate and transport of organic compounds and metals are functions of both site characteristics and the physical/chemical properties of the contaminants. Such properties include solubility in water, tendency to transform or degrade (usually described by a half-life or an environmental half-life in a given medium), and chemical affinity for solids or organic matter [usually described by a partitioning coefficient (K_d , K_{oc} , or K_{ow})]. These properties and how they affect inorganic and organic contaminant behavior are described below.

6.2.1 Metals

Inorganic SRCs at 16 SWMUs are subject to movement with soil moisture and may be transported through the vadose zone to groundwater. Metals do not degrade, although some metals can transform to other oxidation states in soil, reducing their mobility and toxicity. Metals also react with soils or other solid surfaces by ion exchange, sorption, precipitation, or complexation. Such reactions are affected by the pH; Redox conditions; and type and amount of organic matter, clay, and hydrous oxides present. In general, these reactions are reversible and cause an element's mobility to be retarded. The retardation factor (R_d) numerically describes the extent to which the velocity of the contaminant relative to water is slowed. The R_d is largely derived from the soil/water distribution (or partitioning) coefficient (K_d) expressed by the following relation:

$$R_d = 1 + K_d \times \rho_b / \theta ,$$

where

ρ_b = the soil bulk density (g/cm^3) and
 θ = volumetric soil moisture content.

K_d for the metals at this site may vary by large ranges. It has been found that K_d can vary even by orders of magnitude between samples from the same site. The range of K_d values [obtained from EPA (1996a) and Sheppard and Thibault (1990)] for the inorganic SRCs at the 16 SWMUs is presented in Table 6-1.

**Table 6-1. List of Distribution Coefficients for the
Inorganic SRCs at the 16 SWMUs**

| SRCs | K _d ^a (L/kg) |
|-----------|---------------------------------------|
| Aluminum | 1,500 ^b |
| Antimony | 45 |
| Arsenic | 29 |
| Barium | 41 |
| Beryllium | 790 |
| Cadmium | 75 |
| Chromium | 19 |
| Cobalt | 1,300 (100 to 9,700) ^c |
| Copper | 35 ^b |
| Iron | 800 (290 to 2,240) ^c |
| Lead | 270 (100 to 59,000) ^c |
| Manganese | 750 ^c |
| Mercury | 52 |
| Nickel | 65 |
| Selenium | 5 |
| Silver | 8 |
| Vanadium | 1,000 |
| Zinc | 62 |

^aThe K_d values correspond to pH=6.8 (EPA 1996a) unless otherwise indicated.

^bBaes et al. 1984.

^cSheppard and Thibault 1990.

K_d = Distribution coefficient.

6.2.2 Organic Compounds

The organic compounds detected in soils at the 16 SWMUs include VOCs, SVOCs, pesticides, and PCBs. These contaminants may be degraded in the environment by various processes, including hydrolysis, Redox, photolysis, or biodegradation. Half-lives of organic compounds in various media can vary from minutes to years, depending on the chemical and on the environmental conditions. Degradation may either enhance or reduce the toxicity of a chemical. The biodegradation rates for the organic compounds (SRCs) identified at the 16 SWMUs are presented in Table 6-2. These values are based on the biodegradation half-lives taken from the Handbook of Environmental Degradation Rates (Howard et al. 1991). Although a range of values is presented in that reference, only the lowest biodegradation rates corresponding to the highest half-lives are presented in this report to ensure conservatism in discussing contaminant loss through degradation/decay.

The mobility of an organic compound is affected by its volatility and its partitioning behavior between solids and water, water solubility, and concentration. The Henry's Law constant value (K_H) for a compound is a measurement of the ratio of the compound's vapor pressure to its aqueous solubility. The K_H value can be used to make general predictions about the compound's tendency to volatilize from water. Substances with K_H values less than 10⁻⁷ atmospheres/cubic meters/mole will generally volatilize slowly, while compounds with K_H values greater than 10⁻³ atmospheres/ cubic meters/mole will volatilize rapidly. Vapor pressure is a measurement of the pressure at which a compound and its vapor are in equilibrium. The value can be used to determine the extent to which a compound would travel in air, as well as the rate

Table 6-2. Physical and Chemical Properties of Organic Site-related Contaminants

| Constituents | Mol. Wt | Solubility S_w (mg/L) | S_w @ Temp. °C | K_{ow} (mL/mL) | Vapor Pressure (torr @ °C) | Henry's Constant (K_h) atm·m ³ /mol | K_h @ Temp. °C | Air Diff. Coeff. cm ² /s | K_{oc} mL/g | Biodegradation Rate λ 1/day | Log (K_{ow}) |
|-----------------------------------|---------|-------------------------|------------------|------------------|----------------------------|--|------------------|-------------------------------------|---------------|-------------------------------------|------------------|
| <i>Volatile Organic Compounds</i> | | | | | | | | | | | |
| 1,1-Dichloroethane | 99.0 | 5.06E+03 | 20 | 6.17E+01 | 234 @ 25 | 5.45E-03 | | 0.091 | 5.34E+01 | m | 1.13E-03 |
| 1,1-Dichloroethene | 96.9 | 2.25E+03 | e | 3.02E+01 | 591 @ 25 | 2.61E-02 | e | 0.009 | 6.50E+01 | m | 3.85E-03 |
| 1,1,1-Trichloroethane | 133.4 | 4.40E+03 | 20 | 2.95E+02 | 100 @ 20 | 4.08E-03 | 25 | 0.019 | 1.35E+02 | m | 6.35E-04 |
| 1,1,2-Trichloroethane | 133.4 | 4.42E+03 | e | 1.48E+02 | 30 @ 25 | 9.13E-04 | e | 0.078 | 7.50E+01 | m | 4.75E-04 |
| 1,1,2,2-Tetrachloroethane | 167.9 | 2.97E+03 | e | 1.10E+03 | 10 @ 19.3 | 3.40E-04 | e | 0.071 | 7.90E+01 | m | 9.62E-04 |
| 1,2-Dichloroethane | 99.0 | 8.69E+03 | 20 | 2.82E+01 | 8,690 @ 20 | 1.10E-03 | 25 | 0.091 | 3.80E+01 | m | 9.63E-04 |
| 1,2-Dichloroethene | 96.9 | 8.00E+02 | # | 1.23E+02 | 202 @ 25 | 6.60E-03 | | 0.114 | 7.75E+01 | | 2.41E-04 |
| 1,2-Dichloropropane | 113.0 | 2.70E+03 | 20 | 1.91E+02 | 42 @ 20 | 2.82E-03 | | 0.080 | 4.70E+01 | m | 1.34E-04 |
| 1,3-cis-Dichloropropene | 111 | 2.80E+03 | | 1.15E+02 | 43 @ - | 2.32E-03 | | 0.1 | 7.25E+01 | | 6.19E-03 |
| 1,3-trans-Dichloropropene | 111 | 2.80E+03 | | 1.12E+02 | 34 @ - | 1.80E-03 | | 0.08 | 7.08E+01 | | 2.05 |
| 2-Butanone | 72.1 | 2.75E+05 | | 1.82E+00 | 100 @ 25 | 6.61E-07 | 25 | # | 1.15E+00 | | 2.48E-02 |
| 2-Hexanone | 100.2 | 3.50E+04 | 20 | 2.40E+01 | 2 @ 20 | 7.53E-06 | 20 | 0.078 | 1.51E+01 | | |
| 4-Chloro-3-methylphenol | 142.6 | 3.85E+03 | 20 | 1.26E+03 | 10 @ 30 | 1.03E-06 | | 0.056 | 7.93E+03 | | 3.10 |
| 4-Methyl-2-pentanone | 100.2 | 1.91E+04 | | 5.25E+00 | 270 @ 30 | 3.88E-05 | e | 0.110 | 3.31E+00 | | 2.48E-02 |
| Acetone | 58.1 | 1.00E+06 | e | 5.75E-01 | 95 @ 25 | 5.55E-03 | e | 0.088 | 6.20E+01 | m | 2.48E-02 |
| Benzene | 78.1 | 1.75E+03 | e | 1.35E+02 | | 2.12E-03 | | 0.061 | 5.50E+01 | x | 2.13 |
| Bromodichloromethane | 163.8 | 4.50E+03 | | 7.59E+01 | | | | | | | 1.88 |
| Bromoform | 252.7 | 3.20E+03 | | 2.00E+02 | | 5.32E-04 | | 0.105 | 1.26E+02 | m | 2.30 |
| Bromomethane | 94.9 | 1.75E+04 | 20 | 1.26E+01 | 1,420 @ 20 | 1.53E-04 | 25 | # | 9.00E+00 | m | 9.62E-04 |
| Carbon disulfide | 76.1 | 2.90E+03 | 20 | 1.45E+02 | 298 @ 20 | 2.94E-04 | 25 | # | 4.57E+01 | x | 6.19E-03 |
| Carbon tetrachloride | 153.8 | 8.00E+02 | | 5.37E+02 | 113 @ 25 | 2.93E-02 | | 0.082 | 1.52E+02 | m | 1.93E-03 |
| Chlorobenzene | 112.6 | 4.88E+02 | 25 | 6.92E+02 | 11.8 @ 25 | 3.93E-03 | 25 | 0.073 | 2.24E+02 | m | 1.16E-03 |
| Chlorodibromomethane | 208.3 | 4.50E+03 | | 1.23E+02 | 15 @ 25 | 7.83E-04 | 25 | 0.054 | 6.31E+01 | x | 3.85E-03 |
| Chloroethane | 64.5 | 5.74E+03 | 20 | 2.69E+01 | 2,660 @ 25 | 1.10E-02 | 25 | 0.107 | 1.70E+01 | | 6.19E-03 |
| Chloroform | 119.4 | 9.30E+03 | 25 | 9.33E+01 | 160 @ 20 | 3.39E-03 | 25 | 0.091 | 5.30E+01 | m | 3.85E-04 |
| Chloromethane | 50.5 | 6.36E+03 | | 8.13E+00 | 3,800 @ 20 | 8.82E-03 | 25 | 0.110 | 6.00E+00 | m | 6.19E-03 |
| Dibromochloromethane | 208.3 | 4.50E+03 | | 1.23E+03 | | 7.83E-04 | | 0.054 | 6.31E+01 | x | 3.85E-03 |
| Ethylbenzene | 106.2 | 1.52E+02 | 20 | 1.41E+03 | 10 @ 25.9 | 7.88E-03 | e | 0.075 | 2.04E+02 | m | 3.04E-03 |
| Methylene chloride | 84.9 | 1.30E+04 | e | 1.78E+01 | 429 @ 25 | 2.19E-03 | e | 0.101 | 1.00E+01 | m | 6.19E-03 |
| Styrene | 104.1 | 3.00E+02 | 20 | 1.45E+03 | 5 @ 20 | 2.28E-03 | | 0.071 | 9.12E+02 | m | 3.30E-03 |

Note: Footnotes appear on page 6-5.

Table 6-2. Physical and Chemical Properties of Organic Site-related Contaminants (continued)

| Constituents | Mol. Wt | Solubility S_w (mg/L) | S_w @ Temp. °C | K_{ow} (mL/mL) | Vapor Pressure (torr @ °C) | Henry's Constant (K_h) atm·m ³ /mol | K_h @ Temp. °C | Air Diff. Coeff. cm ² /s | K_{oc} mL/g | Biodegradation Rate λ 1/day | Log (K_{ow}) | | | |
|--------------------------------|---------|-------------------------|------------------|------------------|----------------------------|--|------------------|-------------------------------------|---------------|-------------------------------------|------------------|---|----------|------|
| Semivolatile Organic Compounds | | | | | | | | | | | | | | |
| Tetrachloroethene | 165.8 | 1.50E+02 | 25 | 3.39E+02 | 19 @ 25 | 1.84E-02 | e | 25 | 0.077 | s | 2.65E+02 | m | 4.19E-04 | 2.53 |
| Toluene | 92.1 | 5.15E+02 | 20 | 4.90E+02 | 28 @ 25 | 6.64E-03 | e | 25 | 0.087 | t | 1.40E+02 | m | 3.30E-03 | 2.69 |
| Trichloroethene | 131.4 | 1.10E+03 | e | 5.13E+02 | 77 @ 25 | 1.03E-02 | e | 25 | 0.079 | e | 9.40E+01 | m | 4.19E-04 | 2.71 |
| Vinyl chloride | 62.5 | 1.10E+00 | e | 3.16E+01 | 2,580 @ 20 | 2.70E-02 | | 25 | 0.106 | s | 1.86E+01 | x | 2.41E-04 | 1.5 |
| Xylenes | 106.2 | 2.00E+02 | | 5.89E+02 | 5 @ 20 | 5.25E-03 | | 25 | 0.073 | s | 1.96E+02 | m | 1.93E-03 | 2.77 |
| Pesticides/PCBs | | | | | | | | | | | | | | |
| 1,2-Dichlorobenzene | 147.0 | 1.45E+02 | 25 | 2.40E+03 | 1.5 @ 25 | 1.90E-03 | | 25 | 0.063 | | 3.79E+02 | m | 9.63E-04 | 3.38 |
| 2,4-Dimethylphenol | 122.2 | 5.90E+02 | | # | 98 @ 104 | 2.00E-06 | | 25 | 0.074 | | 2.09E+02 | x | 2.48E-02 | 2.42 |
| 2-Methylnaphthalene | 142.2 | 2.60E+01 | 25 | 7.24E+03 | 10 @ 105 | 2.20E-02 | | | 0.056 | | 4.56E+03 | | | 3.86 |
| Acenaphthene | 154.2 | 3.42E+00 | 25 | 8.32E+03 | 10 @ 131 | 2.41E-04 | | 25 | 0.062 | | 4.90E+03 | m | 1.70E-03 | 3.92 |
| Acenaphthylene | 152.2 | 3.93E+00 | 25 | 1.17E+04 | 3.93 @ 25 | 1.14E-04 | | 25 | 0.062 | | 7.40E+03 | | 2.92E+03 | 4.07 |
| Anthracene | 178.2 | 1.29E+00 | 25 | 2.82E+04 | 1.95E-4 L | 6.50E-05 | e | 25 | 0.042 | | 2.35E+04 | m | 3.77E-04 | 4.45 |
| Benzo(a)anthracene | 228.3 | 1.00E-02 | 24 | 4.07E+05 | 5E-9 @ 20 | 3.35E-06 | e | 25 | # | 0.051 | 3.58E+05 | m | 2.55E-04 | 5.61 |
| Benzo(a)pyrene | 252.3 | 3.80E-03 | 25 | 9.55E+05 | 5E-9 @ 21 | 1.13E-06 | e | 25 | 0.043 | | 9.69E+05 | m | 3.27E-04 | 5.98 |
| Benzo(b)fluoranthene | 252.3 | 1.50E-03 | 25 | 3.72E+06 | 5E-7 L | 1.11E-04 | e | 25 | # | 0.044 ^a | 1230000 | x | 2.84E-04 | 6.57 |
| Benzo(k)fluoranthene | 252.3 | 3.00E-04 | | # | 1E-11 @ 20 | 8.29E-07 | e | 25 | # | 0.044 | 1.23E+06 | x | 8.10E-05 | 6.84 |
| Bis(2-ethylhexyl)phthalate | 390.6 | 1.30E+00 | 25 | 2.00E+05 | 1.2 @ 200 | 3.00E-07 | | 20 | 0.032 | s | 1.11E+05 | m | 1.78E-03 | 5.30 |
| Chrysene | 228.3 | 6.00E-03 | 25 | 4.07E+05 | 6.3E-9 @ 25 | 9.46E-05 | e | 25 | 0.046 | | 3.98E+05 | x | 1.73E-04 | 5.61 |
| Di-N-butylphthalate | 278.4 | 4.00E+02 | 25 | 1.58E+05 | 0.1 @ 115 | 9.38E-10 | e | 25 | 0.042 | | 1.57E+03 | m | 3.01E-02 | 5.20 |
| Di-N-octylphthalate | 390.6 | 3.00E+00 | 25 | 1.58E+09 | 0.2 @ 150 | 3.45E-14 | | 25 | 0.035 | | 9.98E+08 | | 1.90E-03 | 9.20 |
| Dibenz(o,a,h)anthracene | 278.4 | 5.00E-04 | 25 | 9.33E+05 | 1E-10 L | 7.30E-08 | | 25 | 0.042 | | 1.79E+06 | m | 1.84E-04 | 5.97 |
| Fluoranthene | 202.3 | 2.65E-01 | 25 | 2.14E+05 | 5E-6 L | 1.61E-05 | e | 25 | 0.069 | | 4.91E+04 | m | 3.94E-04 | 5.33 |
| Fluorene | 166.2 | 1.90E+00 | 25 | 1.51E+04 | 5.0E+5 @ 20 | 6.36E-05 | e | 25 | 0.055 | | 7.71E+03 | m | 2.89E-03 | 4.18 |
| Indeno(1,2,3-c,d)pyrene | 276.3 | 1.40E-04 | # | 4.57E+07 | 1E-10 L | 1.60E-06 | e | 25 | 0.044 | | 3.47E+06 | x | 2.37E-04 | 7.66 |
| N-nitrosodiphenylamine | 198.2 | 3.51E+01 | 25 | 6.17E+02 | 1.00E-01 | 5.00E-06 | e | 25 | 0.0312 | e | 1.29E+03 | x | 5.10E-03 | 2.79 |
| Naphthalene | 128.2 | 3.10E+01 | e | 2.29E+03 | 0.082 @ 25 | 4.83E-04 | | 25 | 0.059 | | 1.19E+03 | m | 2.69E-03 | 3.36 |
| Phenanthrene | 178.2 | 8.16E-01 | 21 | 2.88E+04 | 1 @ 118 | 3.93E-05 | | 25 | 0.054 | | 1.82E+04 | | 8.66E-04 | 4.46 |
| Phenol | 94.1 | 8.28E+04 | e | 3.02E+01 | 0.35 @ 25 | 3.97E-07 | e | 25 | 0.087 | | 2.88E+01 | x | 2.48E-02 | 1.48 |
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Note: Footnotes appear on page 6-5.

Table 6-2. Physical and Chemical Properties of Organic Site-Related Contaminants (continued)

| Constituents | Mol. Wt | Solubility S _w (mg/L) | S _w @ Temp. °C | K _{ow} (mL/mL) | Vapor Pressure (torr @ °C) | Henry's Constant (K _h) atm·m ³ /mol | K _h @ Temp. °C | Air Diff. Coeff. cm ² /s | K _{oc} mL/g | Biodegradation Rate λ 1/day | Log (K _{ow}) |
|-----------------------|---------|----------------------------------|---------------------------|-------------------------|----------------------------|--|---------------------------|-------------------------------------|----------------------|-----------------------------|------------------------|
| 4,4'-DDT | 354.5 | 2.50E-02 | e 25 | 3.39E+06 | e 1.5E-7 @ 20 | 8.10E-06 | e 25 | 0.039 | 6.78E+05 | m 6.16E-05 | 6.53 |
| Aldrin | 364.9 | 1.80E-01 | e 25 | 3.16E+06 | e 2.3E-5 @ 20 | 1.70E-04 | e 25 | 0.013 | 4.87E+04 | m 1.17E-03 | 6.50 |
| alpha-BHC | 290.8 | 2.00E+00 | e 25 | 6.46E+03 | 0.06 @ 40 | 1.41E-02 | | 0.0142 | e 1.76E+03 | m 2.57E-03 | 3.81 |
| alpha-Chlordane | 409.8 | 5.60E-02 | | 6.03E+02 | | 4.79E-05 | 25 | 0.012 | 3.80E+02 | | 2.78 |
| Arochlor-1254 | 328.4 | 5.70E-02 | 24 | 1.07E+06 | 7.71E-5 @ 25 | 8.37E-03 | 25 | 0.041 | 6.75E+05 | | 6.03 |
| beta-BHC | 290.8 | 7.00E-01 | 25 | 6.31E+03 | 0.7 @ 25 | 9.29E-02 | 40 | 0.018 | 2.14E+03 | m 2.80E-03 | 3.80 |
| delta-BHC | 290.8 | 2.13E+01 | 25 | 1.38E+04 | 0.02 @ 20 | 3.59E-04 | 20 | 0.018 | 8.7E+03 | | 4.14 |
| Dieldrin | 380.9 | 1.86E-01 | | 1.23E+04 | 1.8E-7 @ 25 | 5.84E-05 | | 0.013 | w 2.55E+04 | m 3.21E-04 | 4.09 |
| Endrin ketone | | | | | | | | | | | |
| Endosulfan II | 407.0 | 3.30E-01 | 22 | 4.17E+03 | 1E-5 @ 25 | 1.62E-05 | 25 | 0.012 | w 2.63E+03 | | 3.62 |
| Endosulfan sulfate | 422.9 | 2.20E-01 | | 4.57E+03 | | 2.53E+01 | | 0.011 | w 2.88E+03 | | 3.66 |
| gamma-BHC (Lindane) | 290.8 | 7.00E+00 | | 1.74E+03 | 9.4E-6 @ 20 | 4.93E-03 | | 0.018 | 1.35E+03 | m 1.68E-03 | 3.24 |
| gamma-Chlordane | 409.8 | 5.60E-02 | | 1.20E+05 | | 4.86E-05 | e | 0.012 | w 5.13E+04 | m | 5.08 |
| Heptachlor | 373.3 | 5.60E-02 | | 1.82E+06 | e 3E-4 @ 25 | 1.48E-03 | | 0.037 | 9.53E+03 | m 2.65E-03 | 6.26 |
| Heptachlor epoxide | 389.2 | 3.50E-01 | | 1.00E+05 | e | 3.16E-05 | 25 | 0.012 | w 8.32E+04 | x | 5.00 |
| Methoxychlor | 345.7 | 4.50E-02 | | 1.20E+05 | e 0.04 @ 24 | 1.58E-05 | e 25 | 0.016 | e 8.00E+04 | m 1.90E-03 | 5.08 |
| Explosives | | | | | | | | | | | |
| 2,4-Dinitrotoluene | 182.1 | 2.70E+02 | e 22 | 1.02E+02 | 0.0013 @ 59 | 9.26E-08 | e 25 | 0.203 | e 9.55E+01 | | 2.01 |
| 2,4,6-Trinitrotoluene | 227.13 | 1.00E+02 | 25 | 3.39E+05 | 0.046 @ 82 | 0.0000002 | | | 2.14E+05 | | |
| 2,6-Dinitrotoluene | 182.1 | 2.70E+02 | 22 | 1.12E+02 | 6 @ 150 | 0.00533 | 150 | 0.055 | 7.07E+01 | | 2.05 |
| HMX | 296.2 | | | | | | | | | | |
| RDX | 222.26 | 6.00E+01 | 23 | 7.41E+00 | | | | | 4.67E+00 | | 0.87 |

= Indicates RREL database (EPA 1994a) as the source.

* = Represents calculated values.

e = Soil Screening Guidance: Technical Background Document (EPA 1996a) is the source.

L = Source from EPA 1995.

m = Measured K_{oc} values.

s = Indicates Shen, Schmidt, and Card 1993 as the source.

w = EPA WATER7 database November 1990.

x = Calculated K_{oc} values from Soil Screening Guidance: Technical Background Document (EPA 1996a).

A blank indicates value not available.

Air diffusion coefficients were obtained from EPA 1987 unless otherwise indicated.

Biodegradation half-lives were taken from the Handbook of Environmental Degradation Rates (Howard et al. 1991) unless otherwise indicated.

Solubilities, Henry's Constant, and Log (K_{ow}) were taken from Risk Reduction Engineering Laboratory (RREL) Treatability Database (EPA 1994a) unless otherwise indicated.

of volatilization from soil and solution. In general, compounds with vapor pressures lower than 10^{-7} millimeters mercury will not be present in the atmosphere or soil vapor in significant amounts, while compounds with vapor pressures higher than 10^{-2} millimeters mercury will exist primarily in the soil vapor. Unless the soil is saturated, VOCs will exist primarily in the atmosphere and soil vapor. Polycyclic aromatic hydrocarbons (PAHs) and other SVOCs will exist in both the air and the soil. The air diffusion coefficient is a measurement of the rate of spontaneous mixing, presented in units of square centimeters per second, of one substance with another when in contact or separated by a permeable membrane. The rate of diffusion is proportional to the concentration gradient of a substance, increases with temperature, and is inversely related to density and pressure. In soil systems the principal type of diffusion is from a region of high concentration to a region of low concentration. Diffusion occurs most readily in gases, to a lesser extent in liquids, and least in solids.

Water solubility and the tendency to adsorb to particles or organic matter can correlate with retardation in groundwater transport. The adsorption coefficient/partitioning coefficient (K_d) of an organic compound is related to the organic carbon/water partitioning coefficient (K_{oc}) by

$$K_d = f_{oc} \times K_{oc}$$

where

f_{oc} = fraction of soil organic carbon content.

Chemical-specific K_{oc} values may be obtained from literature or may be calculated using empirical formulas relating the octanol-water partitioning coefficient (K_{ow}) to the K_{oc} . The K_{ow} (milliliters per milliliters) is the ratio of a contaminant's concentration in a system containing water and octanol. K_{ow} is used to estimate the tendency for a chemical to partition between environmental phases of different polarity. Organic compounds with log K_{ow} values less than one are highly hydrophilic, while organic compounds with log K_{ow} values greater than four are nearly insoluble in water and will partition to soil particles. Pesticides/PCBs and semivolatiles usually have higher log K_{ow} values. The most commonly used formula to relate K_{ow} to K_{oc} is

$$K_{oc} = 0.63 \times K_{ow} \text{ (Mills et al. 1985).}$$

Chemicals with relatively high water solubilities and low adsorption coefficients (e.g., acetone, methylene chloride) are expected to remain primarily as dissolved phases and be transported at the same rate as the groundwater flow. Chemicals with lower water solubilities and higher adsorption coefficients (e.g., SVOCs and pesticides) are expected to remain primarily adsorbed to the surface of the soils; their transportation with the groundwater would be very limited and at a much slower rate. Table 6-2 presents the solubility, Henry's Law constant (K_H), vapor pressure, air diffusion coefficients, and biodegradation rate constants for the organic compounds detected in soils and groundwater at the 16 SWMUs. Log K_{ow} , K_{ow} , K_{oc} , and K_d for these compounds are also presented in this table. Volatile organic SRCs at the 16 SWMUs generally have lower K_d s and are expected to move with the groundwater with little retardation. Although VOCs move faster in groundwater, they usually have a shorter half-life and degrade at a faster rate in the environment.

6.3 CONCEPTUAL SITE MODEL

The CSM is a statement of expected site conditions that serves as a paradigm with which observations can be compared and within which predictions can be made. The predictive function of the CSM, of primary importance to contaminant fate and transport analysis, relies on known information and informed

assumptions about the site. The better the information and the greater the accuracy of the assumptions, the more accurately the CSM describes the site and, therefore, the more reliable the predictions.

The CSM presented in this section summarizes the hydrogeologic components (presented in Chapter 2.0) and the distribution of contaminants in the subsurface soils and groundwater (presented in Chapters 9.0 and 10.0). Contaminant migration pathways and release mechanisms are also based on the information presented in Chapters 9.0 and 10.0. Site-specific CSMs for contaminant fate and transport are presented in Chapters 9.0 and 10.0. A summary of the model's elements follows.

6.3.1 Water Balance Components

The potential for contaminant transport begins with precipitation. The actual amount of rainwater available for flow is highly variable and dependent upon soil type and climatic conditions. A water balance calculation can be used as a tool to quantitatively account for all the components of the hydrologic cycle at the 16 SWMUs. A simple steady-state water balance model includes precipitation (P), evapotranspiration (ET), surface runoff (Sr), and groundwater recharge or percolation (Gr) and is defined as follows:

$$P = ET + Sr + Gr$$

or

$$\text{Rainwater available for flow} = Sr + Gr = P - ET.$$

The annual average components of water balance estimates for the individual SWMUs under Phase II RFI are presented under the SWMU-specific sections in Chapters 9.0 and 10.0. The water balance estimations were based on the Hydrologic Evaluation of Landfill Performance (HELP) model (EPA 1994b) calculations for an uncapped landfill cell using precipitation and temperature data for the years 1974 through 1978, which are the most current data available for Savannah, Georgia, in the HELP model.

6.3.2 Contaminant Release Mechanisms and Migration Pathways

6.3.2.1 Infiltration and surface runoff

The potential for contaminant transport begins with precipitation. The actual amount of rainwater available for flow is highly variable and dependent upon soil type and climatic condition. Local topography and geology influence both timing and quantity of runoff. Infiltration is affected by soil type, rainfall intensity, surface condition, and vegetation. The general topography of the 16 SWMUs is sloping from northwest to southeast. The regional topography is dominated by shallow terraces dissected by surface water drainage. The terraces are remnants of sea-level fluctuations. The principal surface water body accepting drainage from the 16 SWMUs is the Canoochee River, which joins the Ogeechee River (see Section 3.3)

6.3.2.2 Subsurface flow system

The infiltrated water that is not lost to evapotranspiration is integrated into the subsurface flow system. The subsurface flow system is comprised of the vadose zone and the saturated zone. The hydrology at the 16 SWMUs is dominated by two aquifers: the surficial aquifer and the Principal Artesian aquifer. The surficial aquifer consists of widely varying amounts of sand, silt, and clay ranging from 35 feet to 150 feet in thickness (Geraghty and Miller 1992). The top of the water table ranges from 2 feet to 15 feet bgs. The Principal Artesian aquifer is separated from the surficial aquifer by a confining layer.

6.3.2.3 Release mechanisms

The principal release mechanisms at the 16 SWMUs are infiltration and leaching to groundwater. Precipitation that does not leave the waste unit as surface runoff infiltrates into the subsurface. Some of the infiltrating water leaves this environment via evapotranspiration after little or no subsurface flow. The remainder of the water percolates into the subsurface flow system. The rate of percolation is controlled by soil cover, ground slope, saturated conductivity of the soil, and meteorological conditions.

Water infiltrating through contaminated surface and subsurface soils may leach contaminants into the groundwater. The factors that affect leaching rate include a contaminant's solubility, the partitioning coefficient, and the amount of infiltration. Whether it is a contaminant's partitioning coefficient or solubility that controls leaching depends on whether leaching is solubility-controlled or sorption-controlled. Insoluble compounds will precipitate out of solution in the subsurface or remain in their insoluble forms with little leaching. The contaminants detected at the 16 SWMUs generally do not form insoluble compounds in the natural environment, so sorption processes and the partitioning coefficient will have the greatest effect on leaching. Those contaminants with small partitioning coefficients will be leached more effectively than those with larger such coefficients.

Another factor that affects whether a contaminant will reach the water table through infiltration of rainwater is the contaminant's rate of decay. Most of the organic compounds decay at characteristic rates that are described by the substance's half-life. For a given percolation rate, those contaminants with long half-lives have a greater potential for contaminating groundwater than do those with shorter half-lives.

The water table at the 16 SWMUs may also cause direct leaching of waste that is submerged in water. Seasonal rises in the water table may increase this leaching during rainy seasons.

Contaminants that are sorbed onto surface soils at the 16 SWMUs can be released by desorption in surface runoff or captured with particulate matter by soil erosion during a rainstorm.

VOCs in surface soil are emitted to air via volatilization. The rate of emission is controlled by the vapor pressure of the organic compounds and decreases rapidly over a short period of time as the volatiles are depleted by release to the atmosphere. VOCs in the subsurface soils are emitted to the atmosphere via vertical diffusion through soil pores. Depending on how extensively diffusion has occurred, gaseous emissions from subsurface soils may be significant.

Particulate matter from contaminated surface soil can become airborne as a result of wind erosion. This process is controlled by vegetative cover, wind speed, moisture, and soil grain size in the surface soils.

6.4 FATE AND TRANSPORT ANALYSIS

6.4.1 Soil Leachability Analysis

Contaminant fate and transport analysis at each SWMU under Phase II RFI involves a series of screening steps to define the contaminant migration constituents of potential concern (CMCOPCs). The CMCOPCs are defined as the constituents that may pose the greatest problem if they migrate from the site source.

The first step of the screening process represents the development of the SRCs. The SRCs are selected by comparing the maximum detected concentrations of all the analytes measured in surface and subsurface soils with their respective FMSR reference background criteria. The FMSR reference background criteria

represent the average background concentration multiplied by a factor of two. If the maximum concentration of an analyte in the soil exceeds its reference background criterion, then that analyte is selected as an SRC.

The second step of the screening process involves comparing the maximum concentrations of the SRCs, developed in the previous step, with EPA generic soil screening levels (GSSLs). The GSSLs are set for Superfund sites for the migration to the groundwater pathway (EPA 1996a).

If an EPA-suggested GSSL for a constituent was not available, the GSSL was back-calculated from the target leachate concentration following EPA's soil screening guidance (EPA 1996a). The target leachate concentration was assumed to be equal to the MCL of the constituent, if available; otherwise, the EPA Region III-suggested risk-based concentration (RBC) for tap water corresponding to a 10^{-6} risk or hazard quotient (HQ) = 1 was used (SAIC 1999a).

The equation given in the soil screening guidance (EPA 1996a) is as follows:

$$\text{Screening Level in Soil} = C_w [K_d + (\theta_w + \theta_a H') / \rho_b]$$

where

C_w = target leachate concentration in (mg/L),

K_d = soil-water partition coefficient (L/kg) = $K_{oc} \times foc$ (for organics),

K_{oc} = soil organic carbon/water partition coefficient (L/kg),

foc = 0.002 (EPA 1996a),

θ_w (water-filled porosity) = 0.3 (EPA 1996a),

θ_a (air-filled porosity) = 0.13 (EPA 1996a),

H' = dimensionless Henry's Law constant, and

ρ_b (bulk density) = 1.5 gm/cubic-meter (EPA 1996a).

In the derivation of EPA GSSLs [dilution attenuation factor (DAF) = 1], direct partitioning is used, assuming groundwater is in contact with the analytes in soil; the groundwater concentration is assumed to be equal to the leachate concentration. However, as soil leachate moves through soil, contaminant concentrations are attenuated by adsorption and degradation. When the leachate reaches the water table, dilution by groundwater further reduces leachate concentrations. This reduction in concentration can be expressed by a DAF. A DAF of 20 is protective for sources up to 0.5 acre in size (EPA 1996a). Analyses presented in Appendix A of EPA's soil screening guidance (EPA 1996a) indicate that DAF of 20 can be protective of larger source areas as well. Considering the small soil contamination area of the majority of Fort Stewart's SWMUs, a DAF of 20 was used for organics. Although metals have higher adsorption factors, they do not biodegrade in the environments. The adsorption factor of a metal is a function of pH. Usually, adsorption of metals in soil is lower for a lower pH. For conservatism, a DAF of 20 was used for metals if the average pH of groundwater at the site was greater than or equal to 5 and the soil contamination area of the site was less than or equal to 0.5 acre in size. If the average pH of groundwater was less than 5 and the soil contamination area of the site was greater than 0.5 acre, then a DAF of 1 was used for metals.

The GSSL is defined as the concentration of a contaminant in soil that represents a level of contamination below which there is no concern under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA), provided conditions associated with soil screening levels (SSLs) are met. Generally, if contaminant concentrations in soil fall below the GSSL and there are no significant ecological receptors of concern, then no further study or action is warranted for that area. However, it should be noted

here that the purpose of this screening is not to identify the contaminants that may pose a risk at a downgradient location, but to target those contaminants that may pose the greatest problem if they migrate from the site. SRCs were identified as CMCOPCs if they exceeded the GSSL. To evaluate leaching of CMCOPCs at the 16 SWMUs from soil to groundwater, groundwater concentrations of CMCOPCs were compared to MCLs. If an MCL for the chemical was not available, the groundwater concentration was compared to the EPA Region III-suggested RBCs corresponding to a 10^{-6} risk or an HQ of 0.1.

6.4.2 Vadose Zone SESOIL Modeling of the Contaminant Migration Constituents of Potential Concern

Vadose zone modeling of CMCOPCs (if any) using the Seasonal Soil Compartment (SESOIL) model (Appendix K) was performed for the SWMUs that required a baseline risk assessment. SESOIL was used to predict the maximum groundwater concentration of the CMCOPCs in soil.

The input data for SESOIL can be grouped into four types: climatic, chemical, soil, and application data. There are a total of 61 separate parameters contained in these four data groups. Wherever possible, site-specific parameter values were used for modeling. Certain parameters, however, were not available and were estimated based on a pertinent scientific literature search, geochemical investigations, and consistency checks between model results and historical data. Conservative estimates were used when a range of values was indicated or when parameter values were not available.

6.4.2.1 Climate data

The climatic data file of SESOIL consists of an array of values for various climatic parameters (Appendix K, Table K-1). The climatic parameters were taken from the SESOIL database. The nearest rain gauge station to FSMR in the SESOIL database is Savannah, Georgia.

6.4.2.2 Chemical data

The pollutant fate cycle of SESOIL focuses on the various chemical transport and transformation processes that may occur in the soil zone. These processes include volatilization/diffusion, adsorption/desorption, cation exchange, biodegradation, hydrolysis, and metal complexation. The chemical's solubility in water, air diffusion coefficient, Henry's Law constant, and organic carbon partitioning coefficient are parameters required as input to the model. These chemical-specific values are presented in Table 6-2.

The lowest biodegradation rates from the Handbook of Environmental Degradation Rates (Howard et al. 1991), presented in Table 6-2, were used. If the biodegradation rate of a chemical was not found in the literature, it was assumed to be zero. The process of hydrolysis was not considered in this study because the rates of hydrolysis for certain organic chemicals may vary by more than 14 orders of magnitude. The use of such values in the model would place a high degree of uncertainty on the SESOIL results. Therefore, hydrolysis parameters were set to zero for this analysis, resulting in conservative output.

6.4.2.3 Soil data

The soil data file of SESOIL contains input parameters describing the physical characteristics of the subsurface soil. The parameters include: soil bulk density, intrinsic permeability, soil disconnectedness index, soil porosity, organic carbon content, soil moisture content, infiltration rate, depth to the water table, aquifer thickness, and area of the source. The infiltration rate was based on a water balance calculation using the HELP model (EPA 1994b) (see Section 6.3.1).

If a site-specific soil parameter was not available, a conservative default value was used (Appendix K, Tables K-2.1 through K-2.5). There is no measurement method for the Freundlich exponent (used in calculating the adsorbed contaminant concentration); therefore, the SESOIL default value was used for this parameter. The intrinsic permeability for the vadose zone was calibrated. The soil disconnectedness index replaces moisture retention curves (or characteristic curves) used by other unsaturated zone leaching models. The SESOIL User's Guide (General Sciences Corporation 1996) defines this parameter as being the exponent relating the "wetting" and "drying" time-dependent permeability of soil to its saturated permeability. This one-variable approach of using the soil disconnectedness index in SESOIL simplifies the data estimation process and reduces computation time.

6.4.2.4 Initial condition/source-term concentrations

Analytical data from soil samples collected during the RFI for 16 SWMUs were used as initial concentrations for SESOIL modeling. These data are presented in Chapter 9.0 or Chapter 10.0 for the individual SWMUs. The loading of initial concentrations as input to SESOIL was based on the soil sampling intervals. The initial condition/source-term concentrations used for the SWMUs are presented in Appendix K, Tables K-3.1 through K-3.5.

6.4.2.5 Model application

The SESOIL model used for leachate modeling in the RFI estimates pollutant concentrations introduced into the subsurface via direct application and/or interaction with other media. The model defines the soil compartment as a column extending from the ground surface through the unsaturated zone to the upper level of the saturated soil zone. Processes simulated in SESOIL are categorized in three cycles: hydrologic cycle, sediment cycle, and pollutant cycle. Each cycle is a separate submodule in the SESOIL code. The hydrologic cycle includes rainfall, surface runoff, infiltration, soil-water content, evapotranspiration, and groundwater recharge. The pollutant cycle includes advective transport, volatilization, adsorption/desorption, and degradation/decay. A contaminant in SESOIL can partition into up to four phases (aqueous, gaseous, adsorbed, and free liquid). Data requirements for SESOIL are not extensive because the model uses a minimum number of soil and chemical parameters and meteorological values as input. Output from the SESOIL model includes pollutant concentrations at various soil depths and pollutant loss through surface runoff, percolation to groundwater, volatilization, and degradation. The mathematical formulations used in the SESOIL code generally consider the rate at which the modeled processes occur, the interaction of these processes, and the initial conditions of the waste area and surrounding hydrogeologic formations. The models were arranged in four layers (Appendix K, Tables K-3.1 through K-3.5). Layers 1 through 3 were divided into sublayers to facilitate contaminant loading at intervals closely approximating the actual sampling points and the analytical results. The first three layers are constituent loading zones. The fourth layer represents the leaching zone. This layer was divided into sublayers to improve model precision. The fourth layer is very thin and lies just above the water table; it was used to record predicted leachate concentrations at the water table/vadose zone interface. The SESOIL simulations were continued until the maximum concentration in groundwater was attained.

6.4.3 Saturated Zone Groundwater Modeling

Saturated zone modeling using the Analytical Transient 1-, 2-, 3-Dimensional (AT123D) and One-dimensional Analytical Solute Transport (ODAST) (see Appendix K) was performed to support ecological and baseline risk assessments. Saturated zone modeling was performed if the CMCOPCs from the vadose zone or ecological contaminants of potential concern (ECOPCs)/human health contaminants of potential concern (HHCOPCs) in groundwater had the potential to migrate to the nearest surface water receptor.

AT123D was used for the modeling of organic contaminants, while ODAST was used for the modeling of inorganic contaminants. ODAST was selected over AT123D for the modeling of inorganic contaminants because the maximum simulation period in AT123D is limited to 100 years. Simulation for a period of 1,000 years was performed for inorganic chemicals using ODAST because inorganic chemicals usually do not biodegrade and move very slowly.

6.4.3.1 AT123D modeling

AT123D is a well-known and commonly used analytical groundwater pollutant fate and transport model (see Appendix K). The model computes the spatial-temporal concentration distribution of chemicals in the aquifer system and predicts the transient spread of a chemical plume through a groundwater aquifer. The fate and transport processes accounted for in AT123D are advection, dispersion, adsorption/retardation, and decay. This model can be used as a tool for estimating the dissolved concentration of a chemical in one, two, or three dimensions in the groundwater resulting from a mass release (either continuous or instant or depleting source) over a source area (i.e., point, line, area, or volume source).

Steady-state constant-source AT123D models were developed by calibrating the model against the maximum-observed or SESOIL-predicted maximum concentrations in groundwater beneath the SWMUs. Parameters needed for AT123D model simulations are listed in Appendix K, Tables K-7.1 and K-7.2. If a site-specific parameter was not available, the EPA default value was used. For conservatism, source concentrations were assumed to be constant in the model, and the biodegradation rate for a chemical was taken from the lowest biodegradation rate mentioned in the Handbook of Environmental Degradation Rates (Howard et al. 1991).

Conservative steady-state concentrations of organic chemicals predicted by the AT123D model at the ecological or human receptor locations were used in the respective baseline risk assessment.

6.4.3.2 ODAST modeling

The ODAST program evaluates the one-dimensional analytical solute transport solution considering convection, dispersion, decay, and adsorption in porous media. It uses an analytical equation (Appendix K) published by Van Genuchten and Alves (1982). It includes two simple function-type subroutines using FORTRAN computer code. One subroutine calculates the product of the exponential [$\exp(A)$] and the complementary error function [$\text{erfc}(B)$]; the FORTRAN code was written by Van Genuchten and Alves (1982). The input data for this program is very short and simple. The list of parameters required for ODAST modeling is also provided in Appendix K, Tables K-5.1 through K-5.4.

The ODAST model was used to predict the maximum concentration of inorganic chemicals at the ecological/human receptor location from the maximum-observed or SESOIL-predicted maximum groundwater concentration beneath the SWMUs. It was assumed that the source concentration remains at the maximum-observed/SESOIL-predicted concentration for a period of 70 years. Models developed using ODAST were simulated for a period of 1,000 years. The maximum concentration predicted by ODAST at the receptor location was used for the respective human/ecological risk assessment.

6.4.4 Natural Attenuation of the Contaminant Migration Constituents of Potential Concern

Natural attenuation refers to the observed reduction in contaminant concentrations as contaminants migrate from the source in environmental media. This reduction in concentration in groundwater is due to a number of fate and transport processes, including simple dilution, dispersion, sorption, volatilization, and biotic and abiotic transformations. Abiotic processes of natural attenuation, which include advection,

dispersion, sorption, dilution, volatilization, hydrolysis, dehydrohalogenation, and reduction reactions, usually occur universally. Intrinsic bioremediation of fuel hydrocarbon in groundwater occurs universally, while only some sites contaminated with chlorinated solvents exhibit intrinsic bioremediation. Biodegradation parameters collected during the Phase II RFI include dissolved oxygen, Redox potential, pH, sulfate (SO_4^{2-}), ferrous $[\text{Fe}(\text{II})]$, conductivity, temperature, and TOC.

Organic chemicals can be degraded in the environment through biotic and abiotic processes, which include hydrolysis, Redox, photolysis, biodegradation, or volatilization. As already discussed in Section 6.2, environmental half-lives of organic compounds in various media can vary from minutes to years, depending on the chemical and on the environmental conditions. Organic chemicals with differing chemical structures will biodegrade at different rates. Primary biodegradation consists of any biologically induced structural change in an organic chemical, while complete biodegradation is the biologically mediated degradation of an organic compound into carbon dioxide, water, oxygen, and other metabolic inorganic products. The biodegradation rate of an organic compound is proportional to the concentration:

$$-dC/dt = kC^n$$

where

- C = concentration,
- k = biodegradation rate constant = $1/t \ln (a/[a-x])$,
- t = time,
- a = initial concentration,
- x = change in concentration with time, and
- n = reaction order, $n=1$ for first-order kinetics.

The half-life ($t_{1/2} = \ln 2/k$) is the time necessary for half of the chemical concentration to react. The biodegradation rate of an organic chemical is generally dependent upon the presence and population size of soil microorganisms capable of degrading the chemical. Based on the above equation and the maximum concentrations of these constituents, a simple first-order correlation can be obtained between the constituent's half-life and the time required to degrade the contaminant to the concentration equal to its MCL/RBC.

Metals do not degrade in nature. However, as discussed in Section 6.2.1, metals may be naturally attenuated by ion exchange, Redox, precipitation, or complexation. Although these reactions may be reversible, they cause metals' mobility to be highly retarded.

6.4.5 Identification of Soil Remedial Levels

Remedial levels for soils based on leaching to groundwater are developed for the Phase II sites that require human health baseline risk assessments. An unsaturated zone contaminant transport model (SESOIL) is used to predict the concentration of contaminants in the percolating rainwater before reaching the water table. The SESOIL results are then converted into likely average groundwater concentrations at the site by using dilution factors (DFs). DFs are developed by using the hydraulic analysis method (EPA 1996a), which involves calculating the rate of flow through the aquifer system and the rate of rainwater percolating into the aquifer. Soil remedial levels are calculated based on the ratio of the MCL to the predicted maximum site groundwater concentration for a given analyte. If an MCL was not available for a constituent, the remedial level was based on the risk/hazard for the worst case scenario evaluated in the human health baseline risk assessment.

7.0 HUMAN HEALTH PRELIMINARY RISK EVALUATION METHODOLOGY

The HHPRE uses a Step 1 risk evaluation approach that is based on guidance from the GEPD (Figure 7-1). This evaluation is conducted to determine if there are potential risks to human health associated with contamination detected at the 16 SWMUs. Step 1 involves the following components:

- for inorganics, compare detected concentrations to naturally occurring statistical background levels (Appendix F) to determine if detected inorganics are naturally occurring or are associated with past activities at the site;
- identify potential migration and exposure pathways associated with the site and potential exposure scenarios to determine appropriate action levels;
- identify available risk-based action levels for each contaminant detected above background levels or develop levels if they do not exist; and
- compare sample concentrations to action levels to determine if site conditions warrant further evaluation.

Chemicals that exceed action levels will be identified as HHCOPCs.

7.1 DATA EVALUATION

The objective of the data evaluation step is to develop a set of chemical data that is suitable for use in the HHPRE. The data for the 16 SWMUs were evaluated to determine if they are of sufficient quality for use in the quantitative risk assessment.

The data used in the risk assessment were verified and validated using the methodology described in the QAPP [Part II of the SAP (SAIC 1997)]. Data qualified during the validation as rejected ("R") were not used in the risk assessment.

Detection limits achieved during sample analysis were reviewed to ensure that the required detection limits were met. Typically, detection limit requirements are established to ensure that characterization has occurred to levels that are low enough to determine if chemicals are present at hazardous levels. These levels are chemical-specific and related to each chemical's toxicity. Required detection limits are presented in the QAPP [Part II of the SAP (SAIC 1997)]. In some cases recommended detection limits cannot be achieved by a laboratory (e.g., if matrix or chemical interference requires that a sample be diluted). Samples with elevated detection limits that exceeded 10 times the required detection limit were excluded from the risk assessment data set.

With GEPD concurrence (see Appendix L), all volatile and semivolatile organics were removed from further consideration if the reported sample concentration was at or below 2 µg/L or 2 µg/kg. All acetone values at or below 30 µg/L or 30 µg/kg were treated as nondetects (see Chapter 4.0 and Appendix C). All methylene chloride and toluene values at or below 15 µg/L or 15 µg/kg were also treated as nondetects (see Chapter 4.0 and Appendix C).

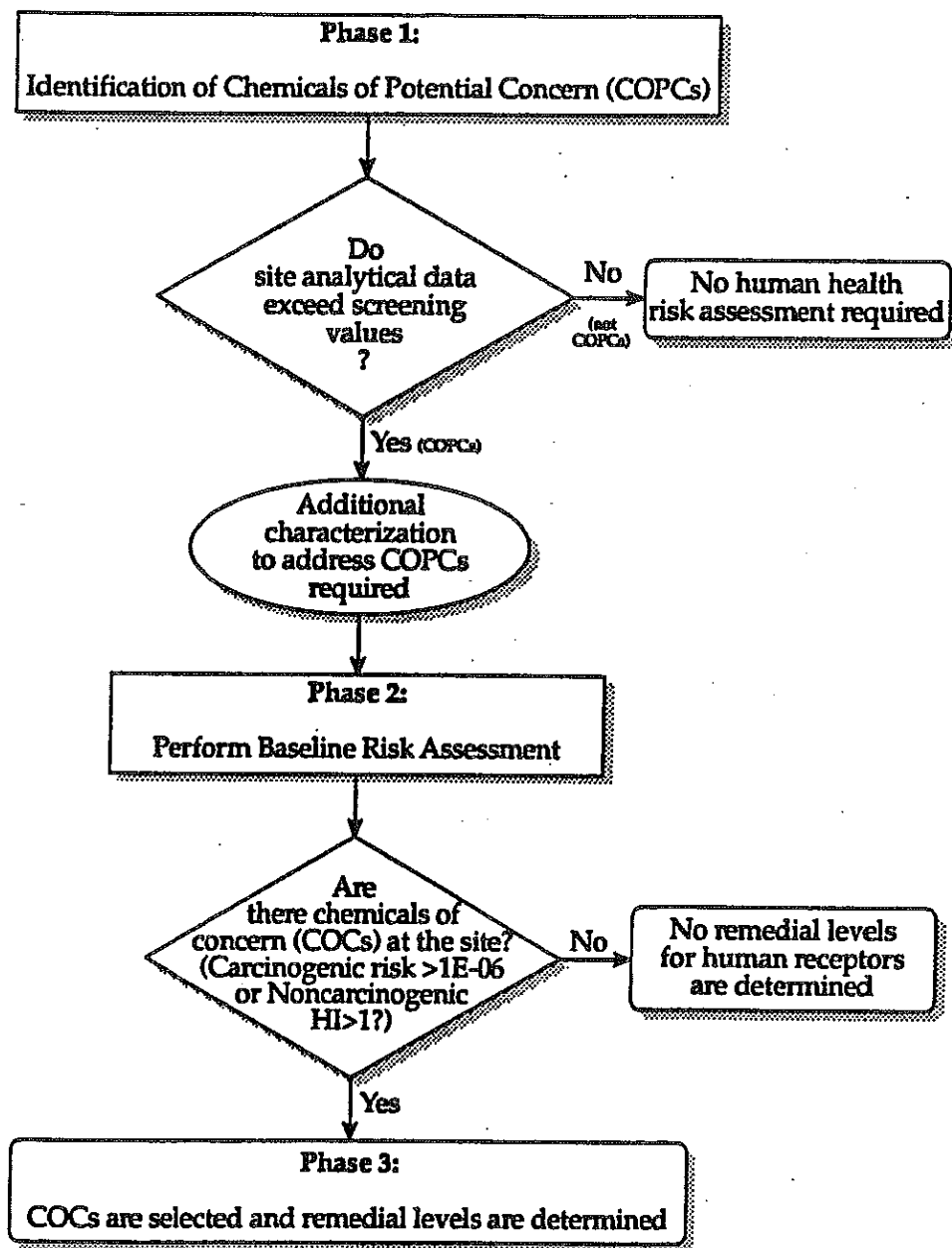


Figure 7-1. General Process for Assessing Risk and Selecting Remedial Levels for Human Receptors*

*Adapted from GEPD guidance for selecting media remediation levels at RCRA SWMUs

Inorganics that are essential nutrients were eliminated, unless they were found at exceptionally high concentrations (EPA 1989). Evidence suggests that there is little potential for toxicity resulting from overexposure to the essential nutrients calcium, magnesium, potassium, and sodium. The highly controlled physiological regulatory mechanisms of these inorganics suggest that there is little, if any, potential for bioaccumulation, and available toxicity data demonstrate that high dietary intakes of these nutrients are well tolerated (National Academy of Sciences 1977; National Research Council 1982; National Research Council 1984).

Background screening for inorganics has been discussed in Chapter 5.0; therefore, it will not be addressed in this section of the document.

7.2 EXPOSURE EVALUATION

The objective of this exposure evaluation is to identify potential exposure pathways that could result in human contact with SRCs. A complete exposure pathway consists of five elements: (1) a potential receptor population, (2) a source of contamination, (3) a transport or retention medium, (4) a point of contact for a receptor, and (5) a route of exposure (ingestion, dermal absorption, or inhalation) through which the chemical may be taken into the body. When all of these elements are present, human exposure to SRCs may take place. The assessment considers both on-site and off-site receptors and their relationship to the potential migration pathways, exposure pathways, and points of exposure for SRCs.

For the purposes of the HHPRE, the exposure assessment identifies potential exposure pathways for selection of pertinent risk-based action. Site-specific discussions are given in Chapters 9.0 and 10.0.

7.2.1 Receptor Assessment

This section identifies those populations that may be exposed to SRCs. These populations will vary among the different sites. For the purposes of this report, different types of sites (e.g., active versus those no longer in use or secured versus unsecured sites) and the general receptor populations that may be present are discussed separately. Site-specific descriptions of receptor populations are given in Chapters 9.0 and 10.0.

Generally, receptor populations are divided into two groups: on-site and off-site receptors. On-site receptors are those individuals who may be present within the site boundaries and come into direct contact with the contaminants present. The exposure of an off-site receptor requires that the contaminant be transported to an off-site exposure point.

The on-site receptors will vary from site to site depending upon the current land use. The sites can generally be divided into different categories depending on the current land use, with receptor populations common to each category. These categories are summarized in Table 7-1 and are discussed below.

Sites that are currently in use would be secured (i.e., the site would be surrounded by a fence or other deterrent and access to the site would be limited to personnel working at the site). The occupational receptor populations would include Installation personnel assigned to work at the site and contract workers (e.g., construction, building maintenance, and repair crews). However, the area of contamination at some of the active sites is not limited to the secured areas, and some soil contamination may exist in open areas adjacent to the site. At these sites juvenile trespassers and other nonoccupational populations may be exposed to the contamination present.

Table 7-1. Generalized Site Descriptions and Associated Receptor Populations

| Site Description | On-site Receptors | Off-site Receptors |
|--|---|--|
| Active; contamination limited to secured area. | Occupational receptor Construction worker | Occupational receptor |
| Active; contamination extends outside secured area. | Occupational receptor Construction worker Juvenile trespasser | Occupational receptor |
| Inactive; secured area. | Occupational receptor Construction worker | Occupational receptor |
| Inactive; unsecured areas within the garrison area. | Occupational receptor Construction worker Juvenile trespasser | Occupational receptor |
| Inactive; unsecured areas outside the garrison area. | Occupational receptor Construction worker Juvenile trespasser Sportsman (hunter) | Occupational receptor Recreational user |

Inactive sites can be divided into secured areas and unsecured or open sites. The secured, inactive sites are located within the garrison area and are represented by sites that are no longer in active use but have equipment or other items that are still present at the site. On-site receptors for these sites would include personnel who enter the site for specific purposes or tasks. Although juvenile trespassers may enter a secured, inactive site, these sites are located in the garrison area and, given the amount of activity in the surrounding area, it is unlikely that a juvenile would be able to enter the sites unnoticed.

The unsecured sites are located in open areas, where a juvenile trespasser may cross the site. Hunting is allowed on the FSMR, so hunters may also represent an on-site receptor population. Military personnel may be present on sites located within active training areas.

Although a receptor population may be identified under current conditions, potential changes in land use may result in the presence of more sensitive receptor populations in the future. The sites within the garrison area are located in developed industrial areas. Although the activities at the sites may change or the sites may become inactive, the sites are likely to remain secured. Therefore, on-site receptors would be limited to personnel working on the site. Inactive sites may be developed for various types of industrial operations; however, as previously discussed, the on-site populations are not likely to change.

None of the sites in the 16 SWMUs is likely to be used for residential purposes. However, to be sure that the first step of the risk assessment process does not exclude any potential future receptors, this assessment assumes that residential use of the site could occur in the future. Residential use of the site is highly unlikely, but is presented as a scenario in accordance with Risk-based Corrective Action (RBCA) guidance.

Off-site receptors include people living and working in the area of the sites as well as recreational users (hunting, fishing, or hiking). The off-site receptor population would change from occupational receptors to residential receptors if the area surrounding a site were to be developed into Installation housing or a residential area.

7.2.2 Migration Pathway Analysis

This section provides a general discussion of the potential chemical transport pathways that may lead to potential exposure points. In general, the major routes of migration are volatilization into air, wind

erosion resulting in fugitive dust, erosion of surface soils into nearby surface waters, leaching of contaminants into groundwater, migration through groundwater, and discharge of groundwater into a surface water body. Sites within the 16 SWMUs may have all or some of these potential migration pathways. The discussion below provides a description of potential migration pathways.

Air. SRCs in soils may be released via volatilization. This migration pathway is generally limited to VOCs found in the surface soils. Particulate-bound chemicals may also be transported to and through the air via generation of fugitive dust. This pathway is limited to compounds that have a high affinity for soils and a low vapor pressure, thus reducing the possibility of volatilization. This migration pathway is limited to chemicals found in surface soils at sites that lack sufficient vegetative cover.

Groundwater. Migration of soil contaminants to groundwater could occur as rainwater infiltrates and percolates through the soil to the groundwater table. The extent of contaminant migration depends primarily on the amount of rainfall, evaporation, solubility of the chemical in water, adsorption of the chemical to the soil, and distance to the groundwater. In general, VOCs [such as benzene, toluene, ethylbenzene, and xylenes (BTEX)] travel more easily through soils than do SVOCs because they are more soluble in water. Solubility of metals is dependent on the metal species and is difficult to generalize. The depth to groundwater at the sites varies from 2 feet to 15 feet bgs.

Contaminants in groundwater may be transported to off-site locations, including surface water bodies. The transportation of contaminants to surface water is generally limited to contaminants that are soluble in water and have a low affinity for soils.

Surface Water. Surface water may serve as a reservoir for contaminants discharged into the water from groundwater or from erosion of contaminated particulates in surface soils. Surface water may serve as a migration pathway as contaminants migrate downstream from their source.

Sediment. Sediment may act as a reservoir for chemicals with a high affinity of sorbing onto solid particles.

7.2.3 Identification of Exposure Pathways

Potential human exposure may occur by primary pathways (i.e., exposure pathways in which the receptor comes in direct contact with contaminated environmental media) or through secondary pathways involving the transfer of SRCs to food sources (i.e., crops, livestock, and game). Potential primary pathways for exposure of receptor populations include incidental ingestion, inhalation of volatile organics and airborne particulates, and dermal contact.

The potential primary and secondary exposure pathways for contaminants present in various environmental media (e.g., soils, groundwater) are discussed in the following sections.

Surface Soil. Potential primary pathways for exposure of receptor populations include ingestion of soils, inhalation of volatile organics and airborne particulates, and dermal contact with soils. Off-site receptors may be exposed via inhalation of fugitive dust. It is unlikely that the volatile compounds will migrate off-site in any significant concentrations; therefore, off-site receptor populations would not be exposed to volatilized compounds.

Indirect exposure pathways for soils would include uptake of contaminants into food sources. Hunting is allowed on the FSMR. Game species may bioaccumulate contaminants as a result of ingesting soils and contaminated vegetation. Current off-site receptors may be exposed as a result of consuming contaminated game. Fort Stewart does not currently lease agricultural lands and is unlikely to allow

agricultural practices within the Installation in the future; therefore, exposure via uptake into food crops is not a viable exposure pathway.

Subsurface Soil. Potential primary exposure pathways for subsurface soils include ingestion of soils, inhalation of volatile organics, and dermal contact with soils. These pathways exist for construction workers or other individuals who would be involved in an excavation. Off-site receptors are unlikely to be directly exposed to contaminants in subsurface soils; however, indirect exposures could occur if contaminants migrated to groundwater.

Groundwater. The surficial groundwater aquifer underlying Fort Stewart is 2 feet to 13 feet bgs. Below this aquifer is the Principal Artesian aquifer, which is hydrogeologically isolated from the surficial aquifer. Drinking water in the area is obtained from the Principal Artesian aquifer, not the surficial groundwater aquifer. The shallow aquifer is currently used in some areas of the region for irrigation or watering of lawns. Direct exposure to irrigation water could include dermal contact and inhalation of volatiles released from the groundwater.

Sediment. Potential direct exposure pathways for sediment would include incidental ingestion by children playing in the surface waters. Exposure via dermal contact is likely to be minimal given that the water is likely to remove the majority of the sediment before chemicals can be absorbed via the skin. Contaminants in sediment may bioaccumulate in benthic (bottom-dwelling) fish and invertebrates, which may be consumed by humans.

Surface Water. Potential direct exposure pathways for surface water include incidental ingestion of water and dermal contact by children playing in the creek. In addition, volatiles released from the water may result in exposure of children and sportsman fishing in the creek. Ingestion of fish may result in exposure to chemicals bioaccumulated into the fish tissue.

7.3 SELECTION OF SCREENING VALUES

Screening values generally represent risk-based action levels and applicable or relevant and appropriate requirements (ARARs) that are publicly available. Screening values inherently incorporate assumptions about land use. In identifying COPCs, it is generally accepted that screening levels will reflect any potential future land uses, and thus usually reflect a conservative residential use scenario (EPA 1991; EPA 1999b; ASTM 1995). Due to their conservative nature, screening values can be used with a high degree of confidence to indicate sites requiring NFA.

Step 1 screening levels generally reflect residential land uses; use of these levels in the first step of the risk process ensures that no chemical will be screened from consideration prematurely. EPA does provide guidance and default parameter values for developing screening levels that reflect industrial land-use assumptions. These levels are developed using equations and default values from EPA (1991). Residential land use is unlikely at any of the sites within the 16 SWMUs.

If risk-based screening values are not publicly available, it generally means that (1) the chemical is not considered to be toxic, except perhaps at extremely high concentrations (e.g., aluminum, sodium); (2) there are no dose-response data indicating a toxic effect; or (3) EPA is currently reviewing toxicity information, and no reference dose or cancer slope factor is currently available.

The following were used as sources of screening values for various media:

- RBCs developed by EPA Region III (EPA 1999b),

- RCRA-based cleanup standards for lead (EPA 1994c), and
- Safe Drinking Water Act (SDWA) technology action levels (EPA 1991).

The RBCs developed by EPA Region III were based on an incremental cancer risk of 1×10^{-6} for carcinogens and a hazard index (HI) of one for noncarcinogens. The screening values for noncarcinogens used in this assessment were based on an HI of 0.1, instead of an HQ of one.

In some cases a screening value was available for a specific compound but not its isomers. In these cases the screening value for the parent compound was used in the risk analysis. A list of these compounds is given below.

- Chlordane was used for alpha-chlordane and gamma-chlordane.
- Endosulfan was used for endosulfan II and endosulfan sulfate.
- Endrin was used for endrin ketone.
- 1,3-Dichloropropene was used for 1,3-*cis*-dichloropropene and 1,3-*trans*-dichloropropene.

For some compounds that did not have screening values, values were available for similar chemicals. If the chemistries of the two compounds were similar, then substitute values were used. There were no screening values for delta-BHC, although screening values were available for the other isomers and technical BHC. Technical BHC is the commercial product used and consists of a mixture of the various isomers. The screening value for technical BHC was used, resulting in a conservative assumption, given that this value takes into account the toxicity of delta-BHC and other more toxic isomers.

Some of the PAHs do not have screening values. The screening value for 2-methylnaphthalene was used for 1-methylnaphthalene based on the similarities in chemical structures. Pyrene was used as a surrogate for phenanthrene. Acenaphthene is similar to acenaphthylene, with acenaphthylene having an additional double bond; therefore the screening value for acenaphthylene was used for acenaphthene.

Benzo(*g,h,i*)perylene does not have a chemical structure that is similar to the PAHs that have toxicity values. Benzo(*g,h,i*)perylene is a questionable carcinogen. Numerous studies have failed to show an increase in the incidence of tumors (EPA 1999a). However, as a conservative measure, it was assumed that this PAH is carcinogenic. The cancer slope factor for carcinogenic PAHs is estimated using toxicity equivalence factors (TEFs), which adjust the slope factor based on the relative carcinogenic potency of the PAH as compared to benzo(*a*)pyrene. Given that benzo(*g,h,i*)perylene is a questionable carcinogen, it is assumed that this PAH would have a TEF of 0.01, which is equal to those of the weaker carcinogenic PAHs.

Screening values for soils, sediments, groundwater, and surface water are discussed below. A summary of the toxicity data for the SRCs is given in Appendix I.

7.3.1 Screening Values for Soils and Sediments

There are no separate risk screening values for sediments. The exposure pathways for sediments include ingestion; therefore, soil screening values for ingestion of soils were used, resulting in a conservative assumption given that the amount of sediment ingested is likely to be far less than that of incidental soil ingestion. Incidental soil ingestion results from soil adhering to the hand and being ingested as a result of

hand-to-mouth behavior. Sediments adhering to the skin are likely to be washed off as the hand is removed from the water; therefore, the amount of sediment adhering to the hand is likely to be much lower than would be the amount of soil. In addition, a person is not as likely to come in contact with sediment as compared to soils; therefore, the frequency of exposure will be lower.

Screening values were selected for residential land-use scenarios. The screening values for soils have been taken from the following sources:

- soil RBCs developed by EPA Region III (EPA 1999b), and
- RCRA-based cleanup standards for lead (EPA 1994c).

The default residential exposure assumptions for soil are described below.

- The assumptions for the soil ingestion pathways are twofold. For noncarcinogens, the receptor is a child (age 1 to 6) who ingests 200 milligrams of soil per day for 6 years. For carcinogens, the soil ingestion rate is age-adjusted over a time period of birth until age 30, assuming an adult ingests 114 milligrams per day (EPA 1996a; EPA 1999b).
- For inhalation of volatiles or fugitive dust, a resident is assumed to be exposed to airborne contaminants for 30 years (EPA 1996a).

The potential exposure pathways for soils present at the 16 SWMUs include ingestion of surface soils, ingestion of subsurface soils (construction worker), inhalation of volatiles, and inhalation of fugitive dust for future land-use scenarios. The value for soil ingestion was used instead of the value for exposure via air because exposure via inadvertent ingestion of soils is a more likely exposure pathway for the majority of the sites.

Toxicity values have been developed for both elemental mercury and mercuric chloride. Given that the exact nature of the mercury detected at the 16 SWMUs is unknown, elemental mercury would be more representative of the various types of mercury that may be present. However, elemental mercury does not have an oral reference dose; therefore, screening values could not be developed for all of the environmental media assessed. For the purposes of this study, the screening value for mercuric chloride was used.

Chromium may exist in two valence states, trivalent (Cr^{+3}) and hexavalent (Cr^{+6}) chromium. Hexavalent chromium is significantly more toxic than trivalent and is more mobile in the environment. However, hexavalent chromium is not naturally occurring and is unstable in the environment, oxidizing to the trivalent state. The risk-based screening values for residential soils include both trivalent and hexavalent chromium. It is unlikely that the chromium present is hexavalent chromium, given that there is no likely source for it. In addition, the value given represents the total chromium present, which includes naturally occurring trivalent chromium. As a conservative assumption, however, the hexavalent chromium value was used for the screening value.

Region III does not have screening levels for lead because lead does not have a reference dose. The EPA cleanup standard for lead represents the maximum concentration in soil that is not likely to have a significant impact upon the health of a child. The EPA screening level is based on a child consuming 200 milligrams of soil per day and estimates lead blood concentrations using biokinetic models.

7.3.2 Screening Values for Groundwater

The groundwater screening values reflect the use of groundwater as a source of drinking water (EPA 1999b). These values include the Region III screening values for tap water. As previously discussed, groundwater at this site is unlikely to be used as a drinking water source, but may be used as a source of water for future irrigation or watering. The drinking water screening values are considered to be health-protective values, given the conservative assumptions used in their derivation.

The default residential exposure assumptions for groundwater are described below.

- The assumptions for the groundwater pathway are twofold. For noncarcinogens, the receptor is an adult who ingests 2 liters of groundwater per day. For carcinogens, the water ingestion rate is age-adjusted over a time period of birth until age 30, assuming a child age 1 to 6 ingests 1 liter per day (EPA 1996a; EPA 1999b).
- Inhalation of volatiles is assumed to occur during showering.

There is no reference dose for lead; however, EPA has derived a technology action level for acceptable lead levels at the tap (EPA 1991). The 15 micrograms per liter action level was used for the lead screening value.

Toxicity values have been developed for both elemental mercury and mercuric chloride. Given that the exact nature of the mercury detected at the 16 SWMUs is unknown, elemental mercury would be more representative of the various types of mercury that may be present. However, elemental mercury does not have an oral reference dose; therefore, screening values could not be developed for all of the environmental media assessed. For the purposes of this study, the screening value for mercuric chloride was used.

7.3.3 Screening Values for Surface Water

Screening values surface water include: Georgia Ambient Water Quality Standards (WQSs) (GEPD 1999c), EPA Region IV WQSs for human health—water and organism ingestion, EPA Region III risk-based criteria for tap water (EPA 1999b), and EPA action levels for drinking water. The different criteria are designed to be protective of human health depending on the types of exposure. The Georgia Ambient WQSs (GEPD 1999c) and the EPA Region IV WQSs for human health—water and organism ingestion, represent the maximum concentrations of contaminants in water that will not present an unreasonable risk to human health if the waters are treated and used as a drinking water source or if aquatic life is harvested from the waters and consumed. The risk-based criteria for tap water and EPA action levels are values applied to water coming from the tap within a home where the water is used for drinking, bathing, cooking, etc.

The WQSs are applicable to many of the surface waters found within the 16 SWMUs. However, the numbers of chemicals addressed in these criteria are limited. Therefore, in the absence of a WQS value, the drinking water standards were applied. This is a conservative approach, given that the drinking water standards assume that the surface water is not treated and that the amount of water ingested is significantly greater for drinking water from a tap than for incidental ingestion from surface waters.

7.4 RISK EVALUATION

The risk evaluation compares the maximum value detected in each medium with the respective screening value for that chemical. If chemical concentrations exceed a screening value, then a risk may exist, and the chemical should be evaluated more carefully (see Figure 7-1).

The selection of COPCs for each environmental medium (surface soil, subsurface soil, groundwater, sediment, or surface water) at each of the sites is addressed in the respective SWMU chapters. The selection process involves two steps. The initial step is the comparison of the maximum concentrations to the appropriate screening values. (Selection of the screening values was discussed in Section 7.3.) Given the conservative nature of the screening values, a weight-of-evidence analysis of those chemicals passing the screening is done to determine if those chemicals selected should be analyzed further in a baseline risk assessment.

7.5 UNCERTAINTY

There are uncertainties associated with all phases of the Tier I risk analysis, including collection and laboratory analysis of the samples and selection of screening concentrations. For the purposes of this report, the general uncertainties are discussed in the following narrative. Site-specific uncertainties, if any, will be discussed as part of the human health risk assessment.

Uncertainties associated with the collection and laboratory analysis of the sampling data may impact the results of the selection process. These uncertainties result from the potential for contamination of samples during collection, preparation, or analysis and from normal error in the analytical techniques. The laboratory validation process minimizes these uncertainties.

The use of blank contamination data also contributes uncertainty to the analysis. Common laboratory contaminants may be excluded from the risk assessment because the associated blank samples were contaminated when these chemicals were actually present in the site-related samples. Conversely, a chemical may be included in the risk assessment because its corresponding blank was "clean" when, in fact, the chemical was a result of laboratory contamination. Site activities and the chemicals expected to result from these activities must be considered when interpreting the data. The data validation process minimizes the uncertainty associated with blank contamination.

Uncertainty is associated with the criteria used for the selection of the screening values. The use of conservative assumptions when selecting the screening values, coupled with the use of low toxicity assessment endpoints [i.e., the use of an HI of 0.1 and an incremental lifetime cancer risk (ILCR) of 1:1,000,000], ensures that those chemicals most likely to contribute significantly to potential risks are evaluated. This is likely to result in chemicals being included as COPCs when they do not present a significant risk to human health.

The use of surrogate values for screening chemicals that do not have screening values adds uncertainty concerning the potential risks associated with those chemicals. Small differences in the structure of a molecule can sometimes have significant effects upon the level of toxicity. This may result in chemicals being included or excluded from the list of COPCs, depending upon differences in the toxicity of the screening compound and the surrogate.

7.6 REMEDIAL LEVELS FOR PROTECTION OF HUMAN HEALTH

Some of the sites within 16 SWMUs (e.g., SWMU 31) will have Interim Removal Actions (IRAs) performed to reduce contamination associated with the site. Remedial levels were calculated for all COPCs at these sites. Remedial levels are based on ARARs or are RBCs designed to be protective of the most sensitive populations.

The risk-based remedial levels for carcinogenic COPCs are calculated using the following formula:

$$RL = \frac{(TR)(ST)}{(I_{oral} \times CSF_{oral}) + (I_{der} \times CSF_{der}) + (I_{inh} \times CSF_{inh})}$$

where

- RL = remedial level for a given COPC,
- ST = source-term concentration of the COPC in a given medium,
- TR = target risk,
- I_{oral} = intake via oral route (mg/kg/day),
- CSF_{oral} = oral cancer slope factor ([mg/kg/day]⁻¹),
- I_{der} = intake via dermal route (mg/kg/day),
- CSF_{der} = cancer slope factor based on absorbed dose of the chemical ([mg/kg/day]⁻¹),
- I_{inh} = intake via inhalation (mg/kg/day), and
- CSF_{inh} = inhalation cancer slope factor ([mg/kg/day]⁻¹).

The equations and exposure factors used to estimate the intake of carcinogens are given in Appendix I, Section I.2.4. The cancer slope factors are discussed in Appendix I, Sections I.3.1 and I.3.3 and listed in Table J-1 in Appendix J. The units for the remedial levels will be the same as the units for the source-term concentration.

The risk-based remedial levels based on systemic toxicity are calculated using the following formula:

$$RL = \frac{(THI)(ST)}{(I_{oral} / RfD_{oral}) + (I_{der} / RfD_{der}) + (I_{inh} / RfD_{inh})}$$

where

- RL = remedial level for a given COPC,
- ST = source-term concentration of the COPC in a given medium,
- THI = target hazard index,
- I_{oral} = intake via oral route (mg/kg/day),
- RfD_{oral} = oral reference dose (mg/kg/day),
- I_{der} = intake via dermal route (mg/kg/day),
- RfD_{der} = reference dose based on absorbed dose of the chemical (mg/kg/day),
- I_{inh} = intake via inhalation (mg/kg/day), and
- RfD_{inh} = inhalation reference dose (mg/kg/day).

The equations and exposure factors used to estimate the intake of carcinogens are given in Appendix I, Section I.2.4. The cancer slope factors are discussed in Appendix I, Sections I.3.2 and I.3.3. The reference doses are given in Table J-1 in Appendix J. The units for the remedial levels will be the same as the units for the source-term concentration.

doses are given in Table J-1 in Appendix J. The units for the remedial levels will be the same as the units for the source-term concentration.

Remedial levels will be estimated for 10^{-6} , 10^{-5} , and 10^{-4} cancer risk levels for each carcinogenic contaminant of concern (COC). Cleanup levels will be estimated for each noncarcinogenic COC at HQ levels of 0.1, 1, and 3. The remedial levels will be protective of current and possible future receptor populations identified in the receptor analysis. Constituents that have both reference doses and cancer slope factors will have remedial levels calculated using both types of toxicological values.

Remedial levels for groundwater may include both RBCs and regulatory levels such as MCLs. Given that MCLs take into consideration both human health and the limitations of technology in removing contaminants from water, these values will be used as remedial levels. In the absence of an MCL, an RBC will be calculated.

RBCs will be used as remedial levels for the other environmental media. The scenarios used to derive the remedial levels for a specific site are discussed in the site-specific sections.

8.0 ECOLOGICAL PRELIMINARY RISK EVALUATION

The state of Georgia allows RCRA facilities to set remediation levels based on an assessment of risk to human health and the environment. All RCRA facilities in Georgia that choose to set risk-based remediation levels, such as Fort Stewart, must prepare risk assessment documentation and propose remediation levels in accordance with the Guidance for Selecting Media Remediation Levels at RCRA Solid Waste Management Units (GEPD 1996). This guidance for ecological risk assessments (ERAs) is based on the guidance contained in EPA Region IV Bulletins, Supplemental Guidance to RAGS, Ecological Risk Assessment (EPA 1996d) and a 1994 draft of Ecological Risk Assessment for Superfund, Process for Designing and Conducting Ecological Risk Assessments (EPA 1994d; EPA 1997a). EPA has also proposed guidelines for conducting ERAs (EPA 1996b). In cases in which GEPD and EPA guidelines differ, the GEPD guidance document takes precedence.

Risk is the likelihood of experiencing adverse effects. The assessment of risk for ecological receptors at 16 SWMUs focuses on identifying and evaluating the potential for harmful effects on ecological receptors as a result of exposure to chemicals released to the environment.

The assessment of risk for ecological receptors at the 16 SWMUs is being conducted under a phased approach in accordance with GEPD (1996) and EPA Region IV (1996d) guidance. The two phases are

- the EPRE and
- the ERA.

The EPRE compares measured concentrations of detected substances to conservative ecological screening values (ESVs) to identify substances detected at the facility that pose a potential hazard to ecological receptors. An ERA is "a qualitative and/or quantitative appraisal of the actual or potential impacts of contaminants from a hazardous waste site on plants and animals other than humans or domesticated species" (EPA 1997a). EPA Region IV (EPA 1996d) and GEPD (1996) guidance emphasizes that ERAs are based on quantitative and site-specific data.

According to GEPD guidance, the risk assessment process terminates with the EPRE if there is no potential hazard or risk to ecological receptors. If contaminants are found to be potential hazards in the EPRE, then additional work may be required. Only those substances that are indicated to be potential hazards in the EPRE are evaluated as ECOPCs in an ERA, if one is required.

The need for an ERA is a risk management decision based on the nature and magnitude of risk to ecological receptors in the environmental setting of the 16 SWMUs. If risk managers decide an ERA is not required, then no further data are collected and ecological risk-based remedial levels are developed based on existing data from the individual SWMUs. Should an ERA be required for one or more of the 16 SWMUs, additional site-specific data will be collected to quantify exposure and evaluate effects (GEPD 1996). Appropriate site-specific data for ERAs include concentrations of contaminants in animals and plants (tissue residues) and toxicity tests (EPA 1997a). Remedial levels for protection of ecological resources are developed and proposed in the ERA for only those substances identified as environmental contaminants of concern in the ERA (GEPD 1996).

This section presents the methods for conducting the EPRE for the 16 SWMUs at Fort Stewart in accordance with GEPD guidance (GEPD 1996). The EPREs for SWMUs that have at least one substance

detected in an environmental medium at a concentration exceeding the respective background criterion are presented in Chapters 9.0 and 10.0.

According to GEPD (1996), the ecological risk evaluation process consists of five steps:

- i. ecological screening value comparison,
- ii. preliminary problem formulation,
- iii. preliminary ecological effects evaluation,
- iv. preliminary exposure estimate, and
- v. preliminary risk calculation.

These five steps correspond to the five steps of the EPA Region IV EPRE (EPA 1996d).

As shown in the flowchart of the GEPD ERA process (Figure 8-1), the first step of the EPRE (Step i) is to screen substances as ECOPCs by comparing the maximum detected concentration to the ESVs. Those substances detected in surface water, groundwater, or sediment at concentrations exceeding background criteria are screened at SWMUs where aquatic biota are potentially exposed to those media. This approach assumes that the most sensitive receptors are those that live in direct contact with the medium and are exposed to contaminants by multiple pathways. If no ECOPCs are identified based on the ESV comparison (Step i), then no further evaluation of risk to aquatic receptors is required. If ECOPCs are identified based on the screening, then they are evaluated further (Steps ii through v). Because there are no ESVs for surface soil, all substances in surface soil at a SWMU are evaluated further in EPRE Steps ii through v. These last four steps of the EPRE represent a preliminary evaluation of the risk to ecological receptors from exposure to ECOPCs occurring at a SWMU.

The preliminary problem formulation (Step ii) identifies categories of potential ecological receptors that occur at the 16 SWMUs and categories of ECOPCs—the substances in surface soil, sediment, surface water, and groundwater that might pose a risk to those receptors in the environmental setting. Preliminary assessment endpoints, ecological receptors, and surrogate species that represent the ecological receptors are selected for evaluation in the preliminary risk calculation (EPA 1996d).

The preliminary ecological effects evaluation (Step iii) identifies toxicity reference values (TRVs) for use in the preliminary risk calculation (EPA 1996d). For the 16 SWMUs, TRVs are average daily doses (ADDs) (milligram per kilogram of body weight per day) for the surrogate species. In accordance with GEPD guidance (GEPD 1996), TRVs are derived from published laboratory toxicity studies.

The preliminary exposure estimate (Step iv) evaluates the potential pathways of exposure for ecological receptors according to the preliminary assessment endpoints. The equations used to calculate ADDs for surrogate species from published values for exposure parameters and measured maximum concentrations of contaminants in surface soil, sediment, surface water, and groundwater at the 16 SWMUs are presented in Step iv (Section 8.4).

The preliminary risk calculation (Step v) calculates an HQ, the ratio of the estimated ADD for the surrogate species at the SWMU and the TRV for the surrogate species, for each ECOPC. HQs are used to evaluate the risk to ecological receptors; to identify ECOPCs in surface soil, sediment, surface water, and groundwater at the 16 SWMUs for ecological receptors; and to support risk management decisions about the need for further evaluation of SWMUs in an ERA and, ultimately, about the need for remediation.

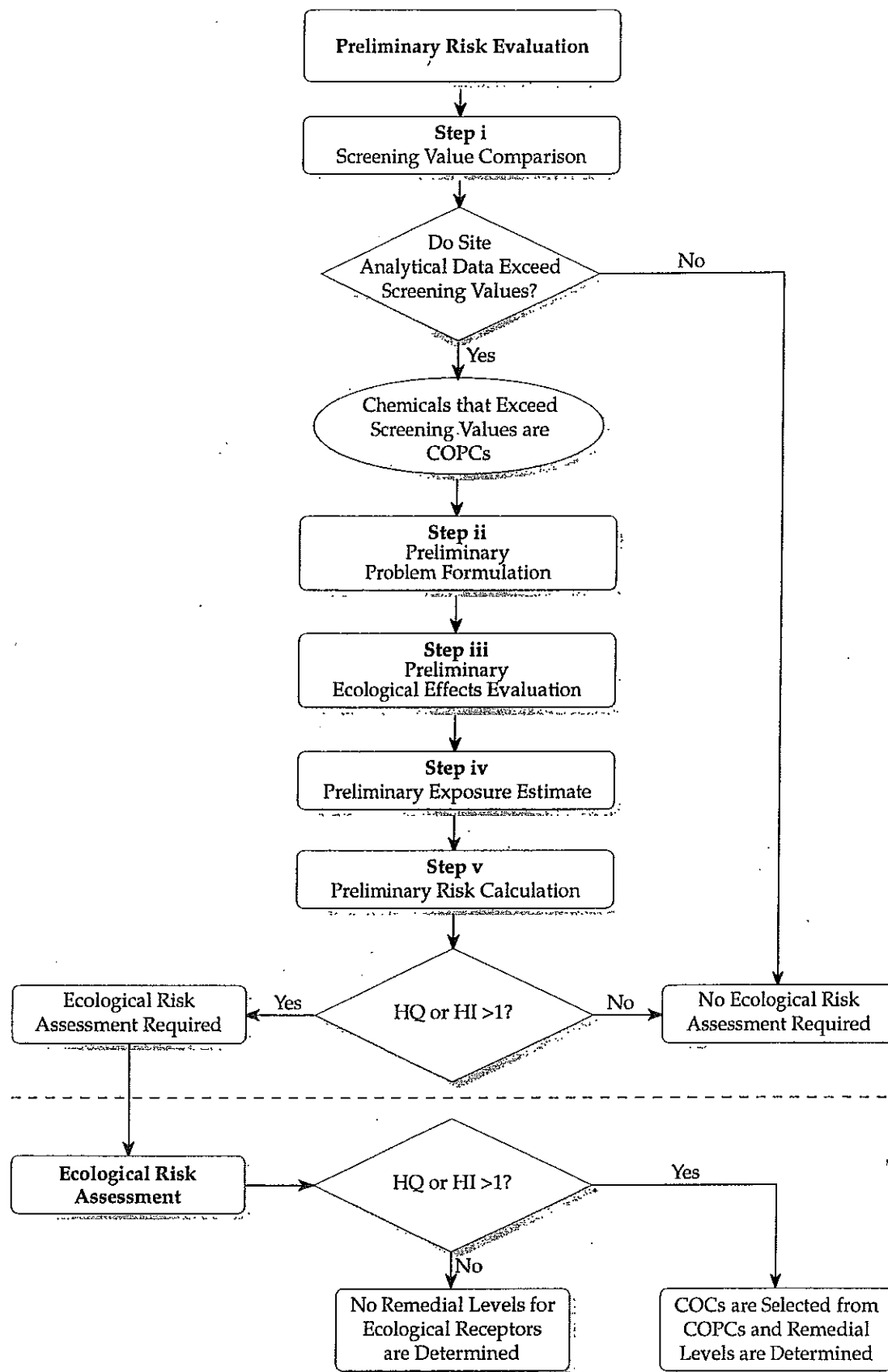


Figure 8-1. General Process for Assessing Risk and Selecting Remedial Levels for Ecological Receptors (adapted from GEPD 1996)

Both terrestrial and aquatic habitats are present at one or more of the 16 SWMUs. Media of concern to ecological receptors are surface soil, surface water, sediment, and groundwater. Surface soil, surface water, and sediment can be contacted or ingested directly by ecological receptors, or their presence in these media can result in the accumulation of contaminants in plants and animals, which can cause ecological receptors ingesting biota to be exposed. Groundwater is also evaluated because it can potentially discharge to springs, seeps, and surface water (EPA 1996d). Thus, up to four media are evaluated in the EPRE for the 16 SWMUs.

8.1 ECOLOGICAL SCREENING VALUE COMPARISON (Step i)

The ESVs used to identify ECOPCs at the 16 SWMUs are EPA Region IV screening values for hazardous waste sites. These are given in Table 8-1 for the substances detected in surface water or groundwater and those detected in sediment at the 16 SWMUs. For analytes without Region IV ESVs, screening values are proposed based on other methods and data obtained from published sources (e.g., Clayton and Clayton 1981) and toxicological databases such as the Hazardous Substances Data Bank and Integrated Risk Information System. Screening values are conservative to prevent elimination of any contaminant that might pose ecological risk. If no data are available to support the development of an ESV for an analyte, the analyte is an ECOPC by default (GEPD 1997a).

For the protection of aquatic biota potentially exposed to surface water and groundwater, such as fish and amphibians, EPA Region IV ESVs are the chronic Ambient Water Quality Criteria (AWQC) for the protection of aquatic life, such as aquatic plants, invertebrates, and fish, or similarly derived values (EPA 1996d). There are no EPA Region IV ESVs for some SRCs, so proposed ESVs for selected analytes are identified from published data sources (Suter and Tsao 1996; Clayton and Clayton 1981). The proposed ESVs are given in Table 8-1.

Sediment ESVs are based on observations of direct toxicity to sediment-dwelling organisms (EPA 1996d). EPA Region IV ESVs for sediment are not available for some SRCs. The proposed ESVs for sediment are given in Table 8-1.

The results of the screening value comparisons for surface water, sediment, and groundwater are presented in the individual sections for each SWMU along with substances detected above background concentrations (see Chapters 9.0 and 10.0). The maximum detected concentrations in samples from each SWMU are used to screen for ECOPCs for aquatic receptors.

A preliminary problem formulation (Step ii), preliminary ecological effects evaluation (Step iii), preliminary exposure estimate (Step iv), and preliminary risk calculation (Step v) are conducted for all SRCs in surface soil because there are no ESVs for terrestrial biota exposed to soil.

8.2 PRELIMINARY PROBLEM FORMULATION (Step ii)

The preliminary problem formulation (Step ii) identifies categories of potential ecological receptors and the substances that may pose a risk to those receptors in the environmental setting of the 16 SWMUs. Preliminary assessment endpoints, ecological receptors, and the surrogate species representative of ecological receptors are selected for evaluation in the preliminary risk calculation.

Table 8-1. Ecological Screening Values for Surface Water and Sediment

| Analyte | Surface Water ESV (µg/L) | Sediment ESV (µg/kg) |
|------------------------------------|-----------------------------|-------------------------|
| <i>RCRA Metals</i> | | |
| Antimony | 160 | 12 |
| Arsenic | 190 | 7.24 |
| Barium | 4.0 ^a | No ESV |
| Cadmium | 0.66 ^b | 1 |
| Calcium | No ESV | No ESV |
| Chromium (III) | 117.32 ^b | 52.3 |
| Chromium (VI) | 11.00 | 52.3 |
| Cobalt | 23 ^a | 50 ^c |
| Copper | 6.54 ^b | 18.7 |
| Iron | 1,000 ^a | 20,000 ^d |
| Lead | 1.32 ^b | 30.2 |
| Magnesium | No ESV | No ESV |
| Manganese | 120 ^a | No ESV |
| Mercury | 0.0123 | 0.13 |
| Nickel | 87.7 | 15.9 |
| Potassium | No ESV | No ESV |
| Selenium | .5 | No ESV |
| Silver | 0.012 ^b | 2 |
| Zinc | 58.9 | 124 |
| <i>Volatile Organic Compounds</i> | | |
| 1,1,1-Trichloroethane | 528 | 0.17 ^e |
| 1,1,2,2-Tetrachloroethane | 240 | 0.94 ^e |
| 1,1,2-Trichloroethane | 940 | No ESV |
| 1,1-Dichloroethane | 47 ^a | No ESV |
| 1,2-Dichloroethene | 590 ^a | No ESV |
| 1,3- <i>cis</i> -Dichloropropene | 0.055 ^a | No ESV |
| 1,3- <i>trans</i> -Dichloropropene | 0.055 ^a | No ESV |
| 2-Hexanone | 99 ^a | No ESV |
| Acetone | 1,500 ^a | 0.00863 ^f |
| Benzene | 53 | 0.057 ^e |
| Bromodichloromethane | No ESV | No ESV |
| Bromomethane | 48 | No ESV |
| Carbon disulfide | 0.92 ^a | No ESV |
| Chloroethane | No ESV | No ESV |
| Chloroform | 289 | 0.1 ^e |
| Chloromethane | No ESV | No ESV |
| Dibromochloromethane | 6,400 ^a | No ESV |
| Ethylbenzene | 453 | 3.5 ^e |
| Methyl ethyl ketone | 14,000 ^a | 0.273 ^f |
| 4-Methyl-2-pentanone | 170 ^a | No ESV |
| Methylene chloride | 1,930 | 0.385 ^f |
| Styrene | 10,000 ^c | No ESV |
| Tetrachloroethene | 84 | 0.53 ^e |
| Toluene | 175 | 0.877 ^f |
| Trichloroethene | 47 ^a | 1.6 ^e |

Note: Footnotes appear on pages 8-6 and 8-7.

Table 8-1. Ecological Screening Values for Surface Water and Sediment (continued)

| Analyte | Surface Water ESV (µg/L) | Sediment ESV (mg/kg) |
|---------------------------------------|-----------------------------|----------------------|
| Vinyl chloride | No ESV | No ESV |
| Xylenes, total | 1.8 ^a | 0.285 ^f |
| <i>Semivolatile Organic Compounds</i> | | |
| 1,2-Dichlorobenzene | 15.8 | 0.34 ^e |
| 2,4-Dimethylphenol | 21.2 | No ESV |
| 2-Methylnaphthalene | No ESV | 330 |
| 4-Chloro-3-methylphenol | No ESV | No ESV |
| Acenaphthene | 17 | 330 |
| Benzo(a)anthracene | 0.027 ^a | 330 |
| Benzo(a)pyrene | 0.014 ^a | 330 |
| Benzo(b)fluoranthene | No ESV | No ESV |
| Benzo(g,h,i)perylene | No ESV | 0.17 ^d |
| Bis(2-ethylhexyl)phthalate | 0.3 | 182 |
| Chrysene | No ESV | 330 |
| Di-N-butylphthalate | 9.4 | 11 ^e |
| Di-N-octylphthalate | No ESV | No ESV |
| Dibenzofuran | 3.7 ^a | 2 ^e |
| Fluoranthene | 39.8 | 330 |
| Indeno(1,2,3-cd)pyrene | No ESV | 0.2 ^d |
| Naphthalene | 62 | 330 |
| Phenanthrene | 6.3 ^a | 330 |
| Phenol | 256 | No ESV |
| Pyrene | No ESV | 330 |
| <i>Explosives</i> | | |
| 2,6-Dinitrotoluene | 100 ^g | No ESV |
| RDX | 970 ^h | 1.3 ^h |
| HMX | 330 ^h | 0.47 ^h |
| <i>Pesticides/PCBs</i> | | |
| beta-BHC | 5,000 | No ESV |
| delta-BHC | No ESV | No ESV |
| alpha-Chlordane | 0.0043 ⁱ | 1.7 |
| gamma-Chlordane | 0.0043 ⁱ | 1.7 ⁱ |
| 4,4'-DDD | 0.0064 | 3.3 |
| 4,4'-DDE | 10.5 | 3.3 |
| 4,4'-DDT | 0.001 | 3.3 |
| Heptachlor | 0.0038 ^a | No ESV |

ESV = EPA Region IV ESVs (EPA 1996d) and, where indicated, alternative values for analytes without ESVs.

^aChronic National Ambient Water Quality Criteria or Tier II values as reported in Suter and Tsao (1996), Table 1 or Table 3.

^bHardness dependent; assumes 50 mg/L CaCO₃.

^cFish LC₅₀ (Clayton and Clayton 1981).

^dOME-L: Ontario Ministry of Environment "Low" values (Persaud, Jaagumagi, and Hayton 1993).

^eSediment quality benchmark (SQB) = surface water ESV (mg/L) × K_{ow} (L/kg) × f_{oc}, where ESV values are from EPA Ecotox Thresholds (EPA 1996d) and fraction organic carbon (f_{oc}) assumed to be 1 percent.

Table 8-1. Ecological Screening Values for Surface Water and Sediment (continued)

^dFor calculation of site-specific SQBs calculated as in footnote d, assuming $f_{oc} = 0.01$.

| Compound | K_{ow}^* (L/kg) | Surface Water ESV (mg/L) | Source of Surface Water ESV |
|---------------------|-------------------|--------------------------|---|
| Acetone | 0.5754 | 1.5 | Chronic Tier II value (Suter and Tsao 1996) |
| Methyl ethyl ketone | 1.95 | 14 | Chronic Tier II value (Suter and Tsao 1996) |
| Methylene chloride | 19.95 | 1.93 | EPA Region IV screening value |
| Toluene | 501 | 0.175 | EPA Region IV screening value |
| Xylenes, total | 1585 | 0.0018 | Chronic Tier II value (Suter and Tsao 1996) |

* K_{ow} s from log K_{ow} s reported in HAZWRAP (1994).

^aEtnier 1987; calculated as one-half criterion continuous concentration for 2,4-dinitrotoluene.

^bTalmage et al. 1999.

^cEPA Region IV ESV value for chlordane.

GEPD (1996) specifies that the EPRE develop "risk characterization for a model ecological receptor." Development of risk characterization for multiple ecological receptors, such as mammals and birds, is allowable for sites at which more than one type of potentially hazardous chemical is detected (GEPD 1997b). Characterizing the risk to multiple receptors can make the EPRE more protective of ecological resources if each receptor type is more sensitive to one or more chemical contaminant. The risk characterization for surface soil, sediment, surface water, and groundwater at the 16 SWMUs considers both mammals and birds as ecological receptors.

8.2.1 Environmental Setting

Ten of the 16 SWMUs (SWMUs 14, 17, 18, 19, 24B, 27, 31, 32, 34, and 37) are located in the cantonment or garrison area of the FSMR (see Figure 2-4). The garrison area of the FSMR is almost entirely industrial in character. The remaining seven SWMUs (SWMUs 2, 3, 9, 10, 11, 12, and 29) are located in the generally forested land surrounding the cantonment area (see Figure 2-3). These areas are used for ranges and training areas or held as non-use areas. The open range and training areas comprise 11 percent of the Installation and consist of grasses, shrubs, and scrub oak.

Sixty-six percent of the approximately 367.2 square miles of forest in the FSMR is pine forest, with the major species being slash pine, loblolly pine (*P. taeda*), and longleaf pine (*P. palustris*). The remainder of the FSMR forest is composed of river bottomlands and swamps, whose major species include tupelo, other gum trees, water oak (*Quercus nigra*), and bald cypress trees (*Taxodium distidium*). The understory of the pine forest is saw-palmetto (*Serenoa repens*). The forests are managed by controlled burning, as evidenced by the presence of burn marks on the mature trees.

The principal surface water body receiving drainage from the FSMR is the Canoochee River, which joins the Ogeechee River (part of the northwestern boundary of the reservation). The individual 16 SWMU sites are located within different subwatersheds (see Figure 2-3).

The principal habitat types present at the 16 SWMUs are industrial, industrial with managed grass or neighboring forestland, forestland, managed grassland, unmanaged grassland, and aquatic. The habitat types present at the 16 SWMUs are presented in Table 8-2. More than one type of habitat may occur at a

Table 8-2. Habitat Types at 16 SWMUs

| SWMU | Site Name | Habitat Type | | | | | |
|-------------------------|--|----------------------|---|-------------|--------------------|----------------------|------------------|
| | | Industrialized Areas | Industrialized Areas with Habitats ^a | Forestlands | Managed Grasslands | Unmanaged Grasslands | Aquatic Habitats |
| 2 | Camp Oliver Landfill | - | - | - | - | • | • |
| 3 | TAC-X Landfill | - | - | • | - | - | • |
| 9 | Inactive EOD Area in Red Cloud Range, Hotel Area | NA | NA | NA | NA | NA | NA |
| 10 | Inactive EOD Area North of Garrison Area | - | - | - | - | • | • |
| 11 | Inactive EOD Area Located Approximately Three Miles Northeast of Garrison Area | - | - | - | - | • | - |
| 12A | Active EOD Containing Open Detonation Unit and Open Burn Unit | - | - | - | - | • | • |
| 14 | Old Fire Training Area | - | - | - | - | • | - |
| 17 | DRMO Hazardous Waste Storage Area | - | • | - | - | - | • |
| 18 | Industrial Wastewater Treatment Plant | - | - | - | • | - | • |
| 19 | Old Sludge Drying Beds | - | - | - | • | • | - |
| 24B | Old Radiator Shop/Paint Booth | • | - | - | - | - | - |
| 27A (Building 1339A) | 3d Squadron, 7th Cavalry Motorpool and Four Associated Oil/Water Separators | - | • | - | - | - | - |
| 27A (Building 1339B) | | - | • | - | - | - | - |
| 27A (Building 1322) | | - | • | - | - | - | - |
| 27B | 1st BN, 3d ADA Motorpool and Associated Oil/Water Separator | - | • | - | - | - | - |
| 27C | 92d ECB (H) Motorpool and Associated Oil/Water Separator | • | - | - | - | - | - |
| 27D | 26th SPT BN Motorpool and Associated Two Oil/Water Separators | • | - | - | - | - | - |

Note: Footnotes appear on page 8-10.

Table 8-2. Habitat Types at 16 SWMUs (continued)

| SWMU | Site Name | Habitat Type | | | | | |
|------------------------------|--|----------------------|---|-------------|--------------------|----------------------|------------------|
| | | Industrialized Areas | Industrialized Areas with Habitats ^a | Forestlands | Managed Grasslands | Unmanaged Grasslands | Aquatic Habitats |
| 27E (Wash Rack 1628) | 703d SPT BN (Main) Motorpool and Associated Two Oil/Water Separators | - | • | - | - | - | - |
| 27E (Building 1720) | | - | • | - | - | - | - |
| 27F (NW of Building 1340) | 3d Engineer Brigade Motorpool and Associated Two Oil/Water Separators | - | • | - | - | - | - |
| 27F (NE of Building 1340) | | - | • | - | - | - | - |
| 27G | DISCOM Motorpool and Associated Oil/Water Separator | - | • | - | - | - | - |
| 27H (Building 1071) | DOL Maintenance Motorpool and Associated Two Oil/Water Separators | • | - | - | - | - | - |
| 27H (Building 1056) | | • | - | - | - | - | - |
| 27I (Block 9900) | NGTC Block 9900, 10300 Motorpool and Associated Two Oil/Water Separators | - | • | - | - | - | - |
| 27I (Block 10300) | | - | • | - | - | - | - |
| 27J (Building 10535) | GANG MATES Motorpool and Associated Two Oil/Water Separators | - | • | - | - | - | - |
| 27J (Building 10531) | | - | • | - | - | - | - |
| 27K | 3d BN, 69th Armor Motorpool Wash Rack and Oil/Water Separator | - | • | - | - | - | - |
| 27L | NGTC Block 10200 Wash Rack and Oil/Water Separator | - | • | - | - | - | - |
| 27M | NGTC Block 10100 Wash Rack and Oil/Water Separator | - | • | - | - | - | - |
| 27N | NGTC Block 9800 Wash Rack and Oil/Water Separator | - | • | - | - | - | - |

Note: Footnotes appear on page 8-10.

Table 8-2. Habitat Types at 16 SWMUs (continued)

| SWMU | Site Name | Habitat Type | | | | |
|------|---|----------------------|---|-------------|--------------------|----------------------|
| | | Industrialized Areas | Industrialized Areas with Habitats ^a | Forestlands | Managed Grasslands | Unmanaged Grasslands |
| 27O | NGTC Block 9700 Wash Rack and Oil/Water Separator | - | • | - | - | - |
| 27P | NGTC Block 9500 Wash Rack and Oil/Water Separator | - | • | - | - | - |
| 27Q | NGTC Block 9400 Wash Rack and Oil/Water Separator | - | • | - | - | - |
| 27R | 396 Transportation Company Wash Rack and Oil/Water Separator | - | • | - | - | - |
| 27S | Two 103d MI BN Wash Racks and Associated Two Oil/Water Separators | - | • | - | - | - |
| 27T | 293 MP Company Wash Rack and Oil/Water Separator | - | • | - | - | - |
| 27U | Two Wright Army Airfield Wash Racks and Oil/Water Separator | - | • | - | • | - |
| 27V | Auto Craft Center Oil/Water Separator | - | • | - | - | - |
| 29 | Evans Army Helipoint POL Storage Facility | - | - | - | • | - |
| 31 | DEH Asphalt Tanks | - | - | - | • | - |
| 32 | Supply Diesel Tank | - | - | - | • | - |
| 34 | DEH Equipment Wash Rack | - | • | - | - | - |
| 37 | NGTC Equalization Basin | - | - | - | • | - |

^aIndustrialized areas with managed grass or neighboring habitats.

NA = Not applicable.

• = Indicates habitat type(s) identified at the SWMU by the on-site biologist.

given SWMU. Brief descriptions of the principal habitat types follow. These descriptions are based on observations made by SAIC personnel during the field sampling investigation conducted in January through March 1998.

8.2.1.1 Industrialized areas

Much of the garrison area consists of man-made structures and surfaces with little or no natural habitat. Buildings, paved roads, gravel or asphalt parking lots, and sidewalks cover the majority of the surface of these areas, many of which are surrounded by chain-link fence. Characteristic flora of industrialized areas consists of grasses and herbaceous weeds growing in spaces between man-made surfaces. Fauna consists mainly of birds that nest in man-made structures or feed upon refuse. Industrial SWMUs are isolated, and mammals, amphibians, and reptiles are unlikely to visit them. SWMUs 24B, 27C, 27D, and 27H are located in industrialized areas with no habitat for wildlife. No ecological risk evaluation was conducted for surface soil at these four SWMUs because of the lack of suitable habitat.

8.2.1.2 Industrialized areas with managed grass or neighboring habitats

The majority of the SWMUs within the garrison area are located adjacent to man-made structures and have small areas of managed grass and/or neighboring forest or grassland habitats. These SWMUs are similar to the industrialized areas described above, but they are bordered by habitats that probably contain animals that might visit them in search of food or water. The neighboring habitats vary from simple, small patches of managed grass to complex, mature forest. Characteristic flora and fauna of these habitats vary depending upon the neighboring habitat type and are described under the section related to that habitat type. The remaining SWMUs in the SWMU 27 complex (i.e., those not listed as having industrial habitat type) as well as SWMUs 17 and 34 are industrialized areas with managed grass or neighboring forest or grassland habitats. Of the SWMUs that are industrialized areas with managed grass or neighboring habitat, no ecological risk evaluation was conducted for surface soil at SWMUs 27G, 27S, and 27V because of the very small areas of open vegetated or bare soil surface.

8.2.1.3 Forestlands

The FSMR beyond the garrison area consists mainly of managed pine forests of two types. Palmetto-pine forest has a canopy of pine trees, such as long-leaf pine, loblolly pine, and slash pine, with an understory of palmetto. Fauna includes a wide variety of birds, mammals, and reptiles. Common species include white-tailed deer (*Odocoileus virginianus*), feral hogs (*Sus scrofa*), wild turkey (*Meleagris gallopavo*), and nine-banded armadillos (*Dasypus novemcinctus*). Palmetto-pine forests are typically managed by controlled burning of the understory.

The other common forest habitat type at the FSMR is pine-oak forest or mixed pine/hardwood forest. Characteristic flora of this habitat type includes slash pine, long-leaf pine, loblolly pine, sweetgum (*Liquidambar styraciflua*), blackgum (*Nyssa sylvatica*), live oak (*Quercus virginiana*), Southern red oak (*Q. falcata*), and white oak (*Q. alba*). Saw-palmetto is commonly found as one of several understory plants. Fauna of the pine-oak forest is similar to that of the palmetto-pine forest, with the addition of gray squirrels (*Sciurus carolinensis*), which feed heavily upon acorns. The habitat at SWMU 3 is predominately pine-oak forest.

8.2.1.4 Managed grasslands

Managed grasslands are found throughout the FSMR. Managed grasslands are typified by planted grass of one or more species maintained by mowing, application of fertilizers, etc. Many of the SWMUs that

exist on the border of or outside the garrison area contain large areas of managed grasses. Common fauna includes earthworms and other soil-dwelling invertebrates, birds such as robins, and mammals such as shrews and rabbits. Managed grasslands are found at SWMUs 18, 19, 27U, 29, 31, 32, and 37.

8.2.1.5 Unmanaged grasslands

Unmanaged grasslands are typically formerly managed grasslands that have undergone succession into meadows of native grasses and weeds because they are no longer mowed. Most of these areas are bordered on one or more sides by forest and are optimal animal foraging sites. Many of these areas have more sand on the surface than vegetation. Immature pine trees are commonly found growing sporadically throughout unmanaged grasslands along with sweetgum and blackgum. Unmanaged grasslands support a diverse fauna, including a large number of small mammals such as shrews, voles, and mice as well as birds and groundhogs (*Marmota monax*). Predators frequent these areas to prey upon the resident fauna. SWMUs 2, 10, 11, 12A, 14, 19, 27U, and 32 contain areas of unmanaged grassland.

8.2.1.6 Aquatic habitats

Aquatic environments are found throughout the FSMR and include streams, tributaries of the Canoochee River, and man-made ponds and basins. Aquatic environments characteristically contain gum trees and water oak, along with other common flora of the surrounding forest. Fauna consists of amphibians, reptiles, fish, aquatic invertebrates, and birds. Aquatic environments are present at SWMUs 2, 3, 10, 12A, 17, and 18. The equalization basin at SWMU 37 is not considered to be an aquatic habitat.

In addition to the above-mentioned aquatic habitat types, many of the OWSs contain ephemeral ponds with aquatic insects and other biota. Due to their isolation and small size (less than 0.1 hectare), ephemeral ponds at the OWSs are not considered to be aquatic habitats and have not been evaluated in the ecological risk evaluation.

Numerous SWMUs are located adjacent to shallow storm water drainage ditches that may contain ephemeral pools of water throughout various times of the year. Temporary pools in drainage ditches may serve as drinking holes for terrestrial animals and potentially as breeding locations for insects and amphibians. These shallow ditches are generally vegetated with grasses and are periodically mowed. In the ecological risk evaluation for the 16 SWMUs, sediment samples taken from drainage ditches at OWSs were evaluated as surface soil, and surface water samples were evaluated as a source of drinking water for terrestrial mammals only. Drainage ditches were not considered to be aquatic habitat in the ecological risk evaluation for the 16 SWMUs.

8.2.2 Surface Soil

The EPRE for surface soil (0 foot to 1 foot) and drainage ditch sediment at the 16 SWMUs evaluated the potential for risk to ecological receptors from ECOPCs. All analytes detected in surface soil and drainage ditch sediment are ECOPCs because there are no EPA Region IV ESVs for soil.

The categories of ecological receptors that are potentially directly exposed to substances in surface soil at the 16 SWMUs are soil bacteria and fungi, vegetation, and animals that come in direct contact with or ingest soil (e.g., soil-dwelling invertebrates). Other categories of receptors are potentially indirectly exposed to soil contaminants that are taken up and stored in the cells or tissues of those organisms that are directly exposed. Herbivorous invertebrates (e.g., insects) and vertebrates (e.g., birds and mammals) are potentially indirectly exposed when they ingest vegetation growing in contaminated soil. Carnivorous

animals are potentially exposed when they ingest animals directly or indirectly exposed to contaminated soil.

The proposed ecological receptors for the surface soil and drainage ditch sediment at the 16 SWMUs are carnivorous small mammals and birds that prey upon soil-dwelling invertebrates. These receptors are proposed because many of the substances detected in soil samples from one or more of the 16 SWMUs potentially biomagnify in soil-dwelling invertebrates (e.g., cadmium, lead, pesticides). In general, there are also greater amounts of published data on the effects of these substances on mammals and birds than there are for other vertebrate wildlife such as reptiles and amphibians.

The preliminary assessment endpoint for surface soil at the 16 SWMUs is protection of small mammals and bird populations from adverse effects. The surrogate species to represent the generic small mammal and bird receptors are the short-tailed shrew (*Blarina brevicauda*) and the American robin (*Turdus migratorius*). The home range of the shrew is small, and robins are territorial during the spring mating season. Earthworms and other soil-dwelling invertebrates represent a large percentage of both species' diets. The life history and behavior of these two surrogate species ensure a conservative estimate of risk.

8.2.3 Surface Water and Groundwater

The EPRE for the 16 SWMUs evaluated the potential for risk to ecological receptors from exposure to surface water or groundwater, which potentially emerges as surface water. For both surface water and groundwater, the same ecological receptor and surrogate species are used to evaluate the potential risk over the same exposure pathways.

The ecological receptors that are potentially directly exposed to substances in surface water or groundwater after it has emerged as surface water are aquatic plants and animals, terrestrial animals that come in direct contact with or ingest surface water, and those animals ingesting aquatic biota that live in the creeks. Amphibians potentially breed in standing water or natural wetlands. Other terrestrial animals potentially drink from creeks or wetland pools. Terrestrial predators of aquatic biota, such as fish-eating birds and mammals, are also likely to be indirectly exposed to contaminants in surface water, and potentially groundwater, through ingestion of aquatic prey. The types of ecological receptors exposed to surface water vary by location. Some SWMUs do not have aquatic habitat and thus do not have aquatic biota. Surface water at these SWMUs is a source of exposure to only those terrestrial animals that come in direct contact with or ingest surface water.

Based on the ECOPCs, the habitat, and the potential exposure pathways at the 16 SWMUs, the proposed ecological receptors for surface water and groundwater are aquatic biota, such as fish and amphibians, and terrestrial animals. Aquatic biota are directly exposed to ECOPCs in surface water. Terrestrial animals are potentially exposed by ingestion of surface water and of aquatic biota that have bioaccumulated substances in their tissues.

The preliminary assessment endpoints for surface water and groundwater at the 16 SWMUs are protection of:

- aquatic biota,
- terrestrial mammal populations from adverse effects from drinking surface water, and

- fish-eating mammals and bird populations from adverse effects from ingesting fish and other aquatic biota.

The aquatic biota assessment endpoint was not evaluated further in the EPRE. No additional evaluation of this assessment endpoint was included because EPA Region IV ESVs for surface water are the only reasonable published values that might serve as surface water TRVs, and there are no additional adjustments to exposure for aquatic biota. The results of the ESV comparison for surface water and groundwater, therefore, identify the ECOPCs for aquatic biota at the 16 SWMUs.

The surrogate species to represent the terrestrial ecological receptors exposed to surface water and groundwater are the raccoon (*Procyon lotor*), the mink (*Mustela vison*), and the green heron (*Butorides striatus*). These species are potentially found at Fort Stewart (GEPD 1997c) and potentially obtain food from creeks and other aquatic habitats. The raccoon is common to the coastal plain of Georgia. Raccoons drink water from shallow surface water bodies and ingest more water per unit body weight than do larger mammals such as the white-tailed deer (*Odocoileus virginianus*): 0.082 gram per gram per day (EPA 1993) versus 0.065 gram per gram per day (Sample and Suter 1994). Mink and herons ingest fish and other aquatic biota living in streams. Thus, the life history and behavior of these species ensure a conservative estimate of risk in accordance with GEPD (1996) and EPA (1997a) guidance.

8.2.4 Sediment

The potential for risk to ecological receptors from exposure to ECOPCs in sediment at the 16 SWMUs was evaluated at several SWMUs with aquatic habitat. The ecological receptors that are potentially exposed to ECOPCs in sediment at the 16 SWMUs are sediment-dwelling biota and terrestrial animals that come in contact with sediment or ingest sediment-dwelling biota living in creek sediments.

Based on the ECOPCs in sediment, the habitat, and the potential exposure pathways, the proposed ecological receptors for sediment are sediment-dwelling biota and terrestrial animals that prey upon sediment-dwelling invertebrates.

The preliminary assessment endpoints for sediment at the 16 SWMUs are protection of:

- sediment-dwelling biota and
- terrestrial animal populations from adverse effects from ingesting sediment-dwelling invertebrates.

The assessment endpoint for sediment-dwelling biota was not evaluated further in the EPRE. No additional evaluation of this assessment endpoint was included because EPA Region IV ESVs for sediment are the only reasonable published values that might serve as sediment TRVs, and there are no additional adjustments to exposure for sediment-dwelling invertebrates. The results of the ESV comparison for sediment, therefore, identify the ECOPCs for sediment-dwelling invertebrates at the 16 SWMUs.

The surrogate species to represent the terrestrial animals exposed to ECOPCs in sediment is the green heron. The green heron is potentially found at Fort Stewart (GEPD 1997c) and potentially uses creeks and ponds as sources of food. Herons ingest biota living in stream sediments. Thus, the life history and behavior of the heron ensure a conservative estimate of risk in accordance with GEPD (1996) and EPA (1997a) guidance.

8.3 PRELIMINARY ECOLOGICAL EFFECTS EVALUATION (Step iii)

The preliminary ecological effects evaluation (Step iii) identifies TRVs for use in the preliminary risk calculation (Step v). As described below, TRVs are derived from no observed adverse effect levels (NOAELs) from laboratory toxicity studies on test species. In the EPRE for the 16 SWMUs, TRVs were required for shrews and robins ingesting contaminated biota exposed to surface soil at the 16 SWMUs, raccoons ingesting contaminated water from drainage ditches, and fish-eating mammals (mink) and wading birds (green herons) ingesting contaminated biota exposed to surface water and sediment in creeks and ponds. The derivation of TRVs for the surrogate species is described below.

First, chronic NOAELs for test species were derived from published chronic or subchronic NOAEL or lowest observed adverse effect level (LOAEL) benchmarks for the test species exposed to the substance in controlled laboratory studies. The derivations of NOAELs for mammals and birds are shown in Tables 8-3 and 8-4, respectively. If a chronic NOAEL was not available for a contaminant, a chronic NOAEL was estimated from a subchronic NOAEL by dividing by an uncertainty factor of 10 (Sample, Opresko, and Suter 1996). Published LOAELs might be used to derive a NOAEL by dividing the LOAEL by a conservative uncertainty factor of 10 (EPA 1996d). Subchronic LOAELs were divided by an uncertainty factor of 100 to estimate a chronic NOAEL.

Most NOAELs and LOAELs for test species were those reported in Sample, Opresko, and Suter (1996). Some NOAELs were found in published toxicity studies or other risk assessments. In some cases, if neither a NOAEL or LOAEL was available for a contaminant, the benchmark for a related compound was used as a surrogate. The chronic NOAELs for PAHs for birds were derived from Shortelle et al. (1997), as cited in QST (1997).

The estimated bird NOAEL for pyrene reported in QST (1997) was used as the chronic NOAEL for the test species. Shortelle et al. (1997), as cited in QST (1997), used linear regression of NOAELs for chemicals for which there are benchmark values for both mammals and birds published in Opresko, Sample, and Suter (1995) to predict the bird NOAEL for SVOCs for which there were mammal data but no bird data. These predicted NOAELs are for a "composite" bird with a body weight equal to the average of all bird test species for the NOAELs used in the regression. These estimated bird NOAELs were used in ERAs for operable units at Fort Sheridan, Illinois (QST 1997; SAIC 1998c).

Once the published and estimated NOAELs for test species were identified or derived as described above, they were used to derive NOAELs for the 16 SWMUs surrogate species, as described below, and these derived NOAELs were used as the TRVs in the EPRE.

Chronic NOAELs for test species of the same taxonomic class as the surrogate species were adjusted for the body weight of the surrogate species to derive TRVs for the surrogate species. That is, mammal test species data were used for mammal surrogate species, and bird test species data were used for bird surrogate species. NOAELs for test species based on ADDs (milligrams per kilograms per day) were adjusted to the surrogate species based on body weight, according to the following equation:

$$\text{surrogate species NOAEL} = \text{test species NOAEL} \times (\text{bw}_{\text{ts}}/\text{bw}_{\text{ss}})^z,$$

where bw_{ts} and bw_{ss} are the body weights (kilograms) of the test species and surrogate species, respectively, and where $z = 0.25$ for mammals and $z = 0$ for birds (Sample, Opresko, and Suter 1996). For example, the published NOAEL for a chemical might be based on data for a 0.35-kilogram rat. The NOAEL for a 0.022-kilogram field mouse would be nearly two times larger than the rat NOAEL. The calculated NOAELs for the surrogate species were the TRVs used in the EPRE.

Table 8-3. Derivation of NOAELs for Mammal Test Species

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BW _i | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark × DCF × ECF |
|---|--------------|---|-----------------------|---------------|----------|-----------------------|--|----------------------------------|----------------------------------|---|
| INORGANICS | | | | | | | | | | |
| Aluminum | Mouse | 3.00E-02 | 1.93E+01 | Chronic | LOAEL | Reproduction | Ondreicka et al. (1966) in [1] | 1.0 | 0.1 | 1.93E+00 |
| Antimony | Mouse | 3.00E-02 | 1.25E+00 | Chronic | LOAEL | Longevity | Schroeder et al. (1968b) in [1] | 1.0 | 0.1 | 1.25E+01 |
| Arsenic | Mouse | 3.00E-02 | 1.26E+00 | Chronic | LOAEL | Reproduction | Schroeder and Mitchner (1971) in [1] | 1.0 | 0.1 | 1.26E+01 |
| Barium | Rat | 4.35E-01 | 5.06E+00 | Chronic | NOAEL | Growth | Perry et al. (1983) in [1] | 1.0 | 1.0 | 5.06E+00 |
| Beryllium | Rat | 3.50E-01 | 6.60E+01 | Chronic | NOAEL | Longevity | Schroeder and Mitchner (1975) in [1] | 1.0 | 1.0 | 6.60E+01 |
| Cadmium | Rat | 3.03E-01 | 1.00E+00 | Chronic | NOAEL | Reproduction | Sutou et al. (1980b) in [1] | 1.0 | 1.0 | 1.00E+00 |
| Calcium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chromium | Rat | 3.50E-01 | 2.74E+03 | Chronic | NOAEL | Reproduction | Ivankovic and Preussmann (1975) in [1] | 1.0 | 1.0 | 2.74E+03 |
| Cobalt | Rat | 3.50E-01 | 1.00E+00 | Chronic | NOAEL | Mortality | Underhill et al. (1931) in [2] | 1.0 | 1.0 | 1.00E+00 |
| Copper | Mink | 1.00E+00 | 1.17E+01 | Chronic | NOAEL | Reproduction | Aulerich et al. (1982) in [1] | 1.0 | 1.0 | 1.17E+01 |
| Iron | None | None | None | None | None | None | None | None | None | No NOAEL |
| Lead ^a | Rat | 3.50E-01 | 8.00E+00 | Chronic | NOAEL | Reproduction | Azar et al. (1973) in [1] | 1.0 | 1.0 | 8.00E+00 |
| Magnesium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Manganese | Rat | 3.50E-01 | 8.80E+01 | Chronic | NOAEL | Reproduction | Laskey et al. (1982) in [1] | 1.0 | 1.0 | 8.80E+01 |
| Mercury | Mink | 1.00E+00 | 1.01E+00 | Chronic | NOAEL | Reproduction | Aulerich et al. (1974) in [1] | 1.0 | 1.0 | 1.01E+00 |
| Nickel | Rat | 3.50E-01 | 4.00E+01 | Chronic | NOAEL | Reproduction | Ambrose et al. (1976) in [1] | 1.0 | 1.0 | 4.00E+01 |
| Potassium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Selenium | Rat | 3.50E-01 | 2.00E+01 | Chronic | NOAEL | Reproduction | Rosenfeld and Beath (1954) in [1] | 1.0 | 1.0 | 2.00E+01 |
| Silver | Rat | 3.50E-01 | 1.01E+02 | Chronic | NOAEL | None | Walker (1971) in [4] | 1.0 | 1.0 | 1.01E+02 |
| Sodium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Vanadium | Rat | 2.60E-01 | 2.10E+00 | Chronic | LOAEL | Reproduction | Domingo et al. (1986) in [1] | 1.0 | 0.1 | 2.10E+01 |
| Zinc | Rat | 3.50E-01 | 1.60E+02 | Chronic | NOAEL | Reproduction | Schlicker and Cox (1968) in [1] | 1.0 | 1.0 | 1.60E+02 |
| ORGANICS | | | | | | | | | | |
| <i>Volatile Organic Compounds</i> | | | | | | | | | | |
| Acetone | Rat | 3.50E-01 | 1.00E+02 | Subchronic | NOAEL | Reproduction | EPA (1986c) in [1] | 0.1 | 1.0 | 1.00E+01 |
| Benzene | Mouse | 3.00E-02 | 2.64E+02 | Chronic | LOAEL | Reproduction | Nawrot and Staples (1979) in [1] | 1.0 | 0.1 | 2.64E+01 |
| Bromodichloromethane | Mouse | 3.00E-02 | 1.79E+01 | Chronic | LOAEL | Gavage | NTP (1986) in [3] | 1.0 | 0.1 | 1.79E+00 |
| Bromoform | Rat | 3.50E-01 | 1.79E+01 | Subchronic | NOAEL | Hepatic lesions | NTP 1989 [3] | 0.1 | 1.0 | 1.79E+00 |
| Bromomethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Carbon disulfide | Rat | 3.50E-01 | 1.10E+01 | Chronic | NOAEL | Fetal toxicity | Hardin et al. (1981) in [3] | 1.0 | 1.0 | 1.10E+01 |
| Carbon tetrachloride | Rat | 3.50E-01 | 1.60E+01 | Chronic | NOAEL | Reproduction | Alumot et al. 1976a in [1] | 1.0 | 1.0 | 1.60E+01 |
| Chlorobenzene | Rat | 3.50E-01 | 5.00E+01 | Chronic | NOAEL | Increase liver/kidney | Knapp et al. 1971 [3] | 1.0 | 1.0 | 5.00E+01 |

Note: Footnotes appear on page 8-19.

Table 8-3. Derivation of NOAELs for Mammal Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BW _i | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark x DCF x ECF |
|---|--------------|---|-----------------------|---------------|----------|--------------------------|---|----------------------------------|----------------------------------|---|
| Chloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chloroform | Rat | 3.50E-01 | 1.50E+02 | Subchronic | NOAEL | Gonad atrophy | Palmer et al. (1979) in [1] | 0.1 | 1.0 | 1.50E+01 |
| Chloromethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Dibromochloromethane | Rat | 3.50E-01 | 2.14E+01 | Subchronic | NOAEL | Hepatic lesions | NTP (1985) in [3] | 0.1 | 1.0 | 2.14E+00 |
| 1,1-Dichloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1-Dichloroethene | Rat | 3.50E-01 | 3.00E+01 | Chronic | NOAEL | Mortality | Quast et al. (1983) in [1] | 1.0 | 1.0 | 3.00E+01 |
| 1,2-Dichloroethane | Mouse | 3.50E-02 | 5.00E+01 | Chronic | NOAEL | Reproduction | Lane et al. (1982) in [1] | 1.0 | 1.0 | 5.00E+01 |
| 1,2-Dichloroethene | Mouse | 3.00E-02 | 4.52E+02 | Subchronic | NOAEL | Blood chemistry | Palmer et al. (1979) in [1] | 0.1 | 1.0 | 4.52E+01 |
| 1,3-Dichloropropene | Rats | 3.50E-01 | 3.00E+00 | Subchronic | NOAEL | Increase in organ weight | Dow Chemical 1973 [3] | 0.1 | 1.0 | 3.00E+01 |
| Ethylbenzene | Rat | 3.50E-01 | 9.71E+01 | None | NOAEL | None | Wolfe et al. (1956) in [3] | 0.1 | 1.0 | 9.71E+00 |
| Methylene chloride | Rat | 3.50E-01 | 5.85E+00 | Chronic | NOAEL | Liver histology | NCA (1982) in [1] | 1.0 | 1.0 | 5.85E+00 |
| Methyl ethyl ketone | Rat | 3.50E-01 | 1.77E+03 | Chronic | NOAEL | Reproduction | Cox et al. (1975) in [1] | 1.0 | 1.0 | 1.77E+03 |
| 4-Methyl-2-pentanone | Rat | 3.50E-01 | 2.50E+02 | Subchronic | NOAEL | Liver/kidney | Microbiological Associates (1986) in [1] | 0.1 | 1.0 | 2.50E+01 |
| Styrene | Dog | 1.00E+01 | 2.00E+02 | Chronic | NOAEL | Unknown | Quast et al. (1979) | 1.0 | 1.0 | 2.00E+02 |
| Tetrachloroethene | Mouse | 3.00E-02 | 1.40E+01 | Subchronic | NOAEL | Hepatotoxicity | Buben and O'Flaherty (1985) in [1] | 0.1 | 1.0 | 1.40E+00 |
| 1,1,2,2-Tetrachloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Toluene | Mouse | 3.00E-02 | 2.60E+02 | Chronic | LOAEL | Reproduction | Nawrot and Staples (1979) in [1] | 1.0 | 0.1 | 2.60E+01 |
| 1,1,1-Trichloroethane | Mouse | 3.50E-02 | 1.00E+03 | Chronic | NOAEL | Reproduction | Lane et al. (1982) in [1] | 1.0 | 1.0 | 1.00E+03 |
| 1,1,2-Trichloroethane | Mouse | 3.50E-02 | 3.90E+00 | Subchronic | NOAEL | Clinical serum chemistry | White et al. 1985 and Sanders et al. 1985 [3] | 0.1 | 1.0 | 3.90E+01 |
| Trichloroethene | Mouse | 3.00E-02 | 7.00E+01 | Subchronic | LOAEL | Hepatotoxicity | Buben and O'Flaherty (1985) in [1] | 0.1 | 0.1 | 7.00E+01 |
| Vinyl chloride | Rat | 3.50E-01 | 1.70E+00 | Chronic | LOAEL | Mortality | Feron et al. (1981) in [1] | 1.0 | 0.1 | 1.70E+01 |
| Xylenes, total | Mouse | 3.00E-02 | 2.06E+00 | Chronic | NOAEL | Reproduction | Marks et al. (1982) in [1] | 1.0 | 1.0 | 2.06E+00 |
| <i>Semivolatile Organic Compounds</i> | | | | | | | | | | |
| Acenaphthene | Mouse | 3.00E-02 | 1.75E+02 | Chronic | NOAEL | None | ATSDR (1997) in [4] | 1.0 | 1.0 | 1.75E+02 |
| Benzo(a)anthracene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [4] | 1.0 | 1.0 | 1.33E+01 |
| Benzo(a)pyrene | Mouse | 3.00E-02 | 1.00E+01 | Chronic | NOAEL | Reproduction | Mackenzie and Angevine (1981) in [1] | 1.0 | 0.1 | 1.00E+00 |
| Benzo(b)fluoranthene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [4] | 1.0 | 1.0 | 1.33E+01 |
| Benzo(g,h,i)perylene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [4] | 1.0 | 1.0 | 1.33E+01 |
| Benzo(k)fluoranthene | Mouse | 3.00E-02 | 1.00E+01 | Chronic | LOAEL | Reproduction | Opresko (1995) in [4] | 1.0 | 0.1 | 1.00E+00 |
| Bis(2-ethylhexyl)phthalate | Mouse | 3.00E-02 | 1.83E+01 | Chronic | NOAEL | Reproduction | Lamb et al. (1987) in [1] | 1.0 | 1.0 | 1.83E+01 |
| Chlorobenzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chrysene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [4] | 1.0 | 1.0 | 1.33E+01 |
| Dibenzofuran | None | None | None | None | None | None | None | None | None | No NOAEL |

Note: Footnotes appear on page 8-19.

Table 8-3. Derivation of NOAELs for Mammal Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Species Body Weight (kg) BW _i | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark x DCF x ECF |
|---|--------------|--|-----------------------|---------------|----------|-----------------|-----------------------------------|----------------------------------|----------------------------------|---|
| 1,2-Dichlorobenzene | Rat | 3.50E-01 | 1.20E+02 | Chronic | NOAEL | Gavage | IRIS (EPA 1997b) | 1.0 | 1.0 | 1.20E+02 |
| 2,4-Dimethylphenol | None | None | None | None | None | None | None | None | None | No NOAEL |
| Di-N-butylphthalate | Mouse | 3.00E-02 | 5.50E+02 | Chronic | NOAEL | Reproduction | Lamb et al. (1987) in [1] | 1.0 | 1.0 | 5.50E+02 |
| Di-N-octylphthalate | None | None | None | None | None | None | None | None | None | No NOAEL |
| Fluoranthene | Mouse | 3.00E-02 | 5.00E+02 | Chronic | LOAEL | None | ATSDR (1997) in [4] | 1.0 | 0.1 | 5.00E+01 |
| 2-Hexanone | None | None | None | None | None | None | None | None | None | No NOAEL |
| Indeno(1,2,3-cd)pyrene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [4] | 1.0 | 1.0 | 1.33E+01 |
| 2-Methylnaphthalene | Rat | 3.50E-01 | 5.00E+01 | Chronic | LOAEL | None | ATSDR (1997) in [4] | 1.0 | 0.1 | 5.00E-00 |
| 4-Chloro-3-methylphenol | None | None | None | None | None | None | None | None | None | No NOAEL |
| Naphthalene | Rat | 3.50E-01 | 5.00E+01 | Chronic | LOAEL | None | ATSDR (1997) in [4] | 1.0 | 0.1 | 5.00E-00 |
| N-Nitrosodiphenylamine | None | None | None | None | None | None | None | None | None | No NOAEL |
| Phenanthrene | Mouse | 3.00E-02 | 1.00E+01 | Chronic | LOAEL | Reproduction | Opresko (1995) in [4] | 1.0 | 0.1 | 1.00E-00 |
| Phenol | Rat | 3.50E-01 | 6.00E+01 | Subchronic | NOAEL | Developmental | NTP (1983) in [3] | 0.1 | 1.0 | 6.00E-00 |
| Pyrene | Mouse | 3.00E-02 | 1.00E+01 | Chronic | LOAEL | Reproduction | Opresko (1995) in [4] | 1.0 | 0.1 | 1.00E-00 |
| <i>Pesticides/PCBs</i> | | | | | | | | | | |
| Aldrin | Rat | 3.50E-01 | 2.00E-01 | Chronic | NOAEL | Reproduction | EPA (1988a) in [1] | 1.0 | 1.0 | 2.00E-01 |
| alpha-BHC | Rat | 3.50E-01 | 1.60E-00 | Chronic | NOAEL | Reproduction | Grant et al. (1977) in [1] | 1.0 | 1.0 | 1.60E-00 |
| beta-BHC | Rat | 3.50E-01 | 4.00E-00 | Subchronic | NOAEL | Organ histology | Van Velsen et al. (1986) in [1] | 0.1 | 1.0 | 4.00E-01 |
| delta-BHC | Rat | 3.50E-01 | 1.60E-00 | Chronic | NOAEL | Reproduction | [alpha-BHC surrogate] | 1.0 | 1.0 | 1.60E-00 |
| gamma-BHC (Lindane) | Rat | 3.50E-01 | 8.00E-00 | Chronic | NOAEL | Reproduction | Palmer et al. (1978) in [1] | 1.0 | 1.0 | 8.00E-00 |
| alpha-Chlordane | Mouse | 3.00E-02 | 4.58E-00 | Chronic | NOAEL | Reproduction | [Chlordane surrogate] | 1.0 | 1.0 | 4.58E-00 |
| gamma-Chlordane | Mouse | 3.00E-02 | 4.58E-00 | Chronic | NOAEL | Reproduction | [Chlordane surrogate] | 1.0 | 1.0 | 4.58E-00 |
| 4,4'-DDD | Rat | 3.50E-01 | 8.00E-01 | Chronic | NOAEL | Reproduction | Fitzhugh (1948) in [1] | 1.0 | 1.0 | 8.00E-01 |
| 4,4'-DDE | Rat | 3.50E-01 | 8.00E-01 | Chronic | NOAEL | Reproduction | Fitzhugh (1948) in [1] | 1.0 | 1.0 | 8.00E-01 |
| 4,4'-DDT | Rat | 3.50E-01 | 8.00E-01 | Chronic | NOAEL | Reproduction | Fitzhugh (1948) in [1] | 1.0 | 1.0 | 8.00E-01 |
| Dieldrin | Rat | 3.50E-01 | 2.00E-01 | Chronic | LOAEL | Reproduction | Treon and Cleveland (1955) in [1] | 1.0 | 0.1 | 2.00E-02 |
| Endosulfan II | Rat | 3.50E-01 | 1.50E-00 | Subchronic | NOAEL | Reproduction | [Endosulfan surrogate] | 0.1 | 1.0 | 1.50E-01 |
| Endosulfan sulfate | Rat | 3.50E-01 | 1.50E-00 | Subchronic | NOAEL | Reproduction | [Endosulfan surrogate] | 0.1 | 1.0 | 1.50E-01 |
| Endrin ketone | Mouse | 3.00E-02 | 9.20E-01 | Chronic | LOAEL | Reproduction | [Endrin surrogate] | 1.0 | 0.1 | 9.20E-02 |
| Heptachlor | Mink | 1.00E-00 | 1.00E-00 | Chronic | LOAEL | Reproduction | Crum et al. (1993) in [1] | 1.0 | 0.1 | 1.00E-01 |
| Heptachlor epoxide | Mink | 1.00E-00 | 1.00E-00 | Chronic | LOAEL | Reproduction | [Heptachlor surrogate] | 1.0 | 0.1 | 1.00E-01 |
| Methoxychlor | Rat | 3.50E-01 | 4.00E-00 | Chronic | NOAEL | Reproduction | Gray et al. (1988) in [1] | 1.0 | 1.0 | 4.00E-00 |

Note: Footnotes appear on page 8-19.

Table 8-3. Derivation of NOAELs for Mammal Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BW ₁ | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark x DCF x ECF |
|---|-----------------|---|--------------------------|------------------|----------|--------------|----------------------|---|--|--|
| | | | | | | | | | | |
| Explosives | | | | | | | | | | |
| 2,4-Dinitrotoluene | Mouse | 3.00E-02 | 1.35E+01 | Chronic | NOAEL | Reproduction | Ellis et al. (1979) | 1.0 | 1.0 | 1.35E+01 |
| 2,6-Dinitrotoluene | Rat | 3.50E-01 | 7.00E-00 | Subchronic | NOAEL | Reproduction | ATSDR (1989) | 0.1 | 1.0 | 7.00E-01 |
| 2,4,6-Trinitrotoluene | Rat | 3.50E-01 | 1.60E+02 | Subchronic | LOAEL | Reproduction | Dilley et al. (1982) | 0.1 | 0.1 | 1.60E-00 |
| HMX | None | None | None | None | None | None | None | None | None | No NOAEL |
| RDX | None | None | None | None | None | None | None | None | None | No NOAEL |

^a = Lead acetate.

ATSDR = Agency for Toxic Substances and Disease Registry.

DCF = 1 if chronic, 0.1 if subchronic (Sample, Opreko, and Suter 1996).

ECF = 1 if NOAEL, 0.1 if LOAEL (Sample, Opreko, and Suter 1996).

IRIS = Integrated Risk Information System.

[1] = Sample, Opreko, and Suter (1996).

[2] = Clayton and Clayton (1981).

[3] = IRIS (EPA 1997b).

[4] = QST (1997); all values assumed to be chronic.

Table 8-4. Derivation of NOAELs for Bird Test Species

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg/ BW _i) | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark × DCF × ECF |
|---|---------------------|---|--------------------------|------------------|----------|--------------|-----------------------------------|---|---|--|
| INORGANICS | | | | | | | | | | |
| Aluminum | Ringed dove | 1.55E-01 | 1.10E+02 | Chronic | NOAEL | Reproduction | Carriere et al. (1986) in [1] | 1.0 | 1.0 | 1.10E+02 |
| Antimony | Composite bird | 8.50E-01 | 3.57E-01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 3.57E-01 |
| Arsenic | Mallard duck | 1.00E+00 | 5.14E+00 | Chronic | NOAEL | Mortality | USFWS (1979) in [1] | 1.0 | 1.0 | 5.14E+00 |
| Barium | Chick (14 days old) | 1.21E-01 | 2.08E+02 | Subchronic | NOAEL | Mortality | Johnson et al. (1960) in [1] | 0.1 | 1.0 | 2.08E+01 |
| Beryllium | Composite bird | 8.50E-01 | 1.67E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.67E+00 |
| Cadmium | Mallard duck | 1.15E+00 | 1.45E+00 | Chronic | NOAEL | Reproduction | White and Finley (1978) in [1] | 1.0 | 1.0 | 1.45E+00 |
| Calcium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chromium | Black duck | 1.25E+00 | 1.00E+00 | Chronic | NOAEL | Reproduction | Haseltine et al. (unpubl.) in [1] | 1.0 | 1.0 | 1.00E+00 |
| Cobalt | None | None | None | None | None | None | None | None | None | No NOAEL |
| Copper | Chick (5 weeks old) | 5.34E-01 | 4.70E+01 | Chronic | NOAEL | Mortality | Mehring et al. (1960) in [1] | 1.0 | 1.0 | 4.70E+01 |
| Iron | None | None | None | None | None | None | None | None | None | No NOAEL |
| Lead ^a | Quail | 1.50E-01 | 1.13E+00 | Chronic | NOAEL | Reproduction | Edens et al. (1976) in [1] | 1.0 | 1.0 | 1.13E+00 |
| Magnesium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Manganese | Quail | 7.20E-02 | 9.77E+02 | Chronic | NOAEL | Growth | Laskey and Edens (1985) in [1] | 1.0 | 1.0 | 9.77E+02 |
| Mercury | Quail | 1.50E-01 | 4.50E-01 | Chronic | NOAEL | Reproduction | Hill and Schaffner (1976) in [1] | 1.0 | 1.0 | 4.50E-01 |
| Nickel | Mallard duckling | 7.82E-01 | 7.74E+01 | Chronic | NOAEL | Growth | Cain and Pafford (1981) in [1] | 1.0 | 1.0 | 7.74E+01 |
| Potassium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Selenium | Mallard duck | 1.00E+00 | 5.00E-01 | Chronic | NOAEL | Reproduction | Heinz et al. (1989) in [1] | 1.0 | 1.0 | 5.00E-01 |
| Silver | Composite bird | 8.50E-01 | 5.79E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 5.79E+01 |
| Sodium | None | None | None | None | None | None | None | None | None | No NOAEL |
| Vanadium | Mallard duck | 1.17E+00 | 1.14E+01 | Chronic | NOAEL | Mortality | White and Dieter (1978) in [1] | 1.0 | 1.0 | 1.14E+01 |
| Zinc | Leghorn chicken | 1.94E+00 | 1.45E+01 | Chronic | NOAEL | Reproduction | Stahl et al. (1990) in [1] | 1.0 | 1.0 | 1.45E+01 |
| ORGANICS | | | | | | | | | | |
| <i>Volatile Organic Compounds</i> | | | | | | | | | | |
| Acetone | None | None | None | None | None | None | None | None | None | No NOAEL |
| Benzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Bromodichloromethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Bromoform | None | None | None | None | None | None | None | None | None | No NOAEL |
| Bromomethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Carbon disulfide | None | None | None | None | None | None | None | None | None | No NOAEL |
| Carbon tetrachloride | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chlorobenzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chloroform | None | None | None | None | None | None | None | None | None | No NOAEL |

Note: Footnotes appear on page 8-23.

Table 8-4. Derivation of NOAELs for Bird Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BW _i | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversi on Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark × DCF × ECF |
|---|----------------|---|--------------------------|------------------|----------|--------------|--------------------------------|--|---|--|
| Chloromethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Dibromochloromethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1-Dichloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1-Dichloroethene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,2-Dichloroethane | Chicken | 1.60E+00 | 1.72E+01 | Chronic | NOAEL | Reproduction | Alumot et al. (1976b) in [1] | 1.0 | 1.0 | 1.72E+01 |
| 1,2-Dichloroethene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,3-Dichloropropene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Ethylbenzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Methylene chloride | None | None | None | None | None | None | None | None | None | No NOAEL |
| Methyl ethyl ketone | None | None | None | None | None | None | None | None | None | No NOAEL |
| 4-Methyl-2-pentanone | None | None | None | None | None | None | None | None | None | No NOAEL |
| Styrene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Tetrachloroethene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1,2,2-Tetrachloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Toluene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1,1-Trichloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,1,2-Trichloroethane | None | None | None | None | None | None | None | None | None | No NOAEL |
| Trichloroethene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Vinyl chloride | None | None | None | None | None | None | None | None | None | No NOAEL |
| Xylenes, total | None | None | None | None | None | None | None | None | None | No NOAEL |
| <i>Semivolatile Organic Compounds</i> | | | | | | | | | | |
| Acenaphthene | Composite bird | 8.50E-01 | 8.78E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 8.78E+01 |
| Benzo(a)anthracene | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.24E+01 |
| Benzo(a)pyrene | Composite bird | 8.50E-01 | 9.97E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 9.97E+00 |
| Benzo(b)fluoranthene | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.24E+01 |
| Benzo(g,h,i)perylene | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.24E+01 |
| Benzo(k)fluoranthene | Composite bird | 8.50E-01 | 9.97E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 9.97E+00 |
| Bis(2-ethylhexyl)phthalate | Ringed dove | 1.55E-01 | 1.10E+00 | Chronic | NOAEL | Reproduction | Peakall (1974) in [1] | 1.0 | 1.0 | 1.10E+00 |
| Chlorobenzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| Chrysene | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.24E+01 |
| Dibenzofuran | None | None | None | None | None | None | None | None | None | No NOAEL |
| 1,2-Dichlorobenzene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 2,4-Dimethylphenol | None | None | None | None | None | None | None | None | None | No NOAEL |
| Di-N-butylphthalate | Ringed dove | 1.55E-01 | 1.11E+00 | Chronic | LOAEL | Reproduction | Peakall (1974) in [1] | 1.0 | 0.1 | 1.11E-01 |

Note: Footnotes appear on page 8-23.

Table 8-4. Derivation of NOAELs for Bird Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Species Body Weight (kg) BW _t | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark × DCF × ECF |
|---|----------------------|--|-----------------------|---------------|----------|--------------|----------------------------------|----------------------------------|----------------------------------|---|
| Di-N-octylphthalate | None | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 1.95E+02 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.95E+02 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.24E+01 |
| | Composite bird | 8.50E-01 | 3.39E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 3.39E+01 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 3.39E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 3.39E+01 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 9.97E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 9.97E+00 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| Pyrene | Composite bird | 8.50E-01 | 9.97E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 9.97E+00 |
| Pesticides/PCBs | | | | | | | | | | |
| Aldrin | Composite bird | 8.50E-01 | 1.73E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 1.73E+00 |
| | Japanese quail | 1.50E-01 | 5.63E-01 | Chronic | NOAEL | Reproduction | Vos et al. (1971) in [1] | 1.0 | 1.0 | 5.63E-01 |
| | Japanese quail | 1.50E-01 | 5.63E-01 | Chronic | NOAEL | Reproduction | [alpha-BHC surrogate] | 1.0 | 1.0 | 5.63E-01 |
| | Japanese quail | 1.50E-01 | 5.63E-01 | Chronic | NOAEL | Reproduction | [alpha-BHC surrogate] | 1.0 | 1.0 | 5.63E-01 |
| | Mallard duck | 1.00E+00 | 2.00E+00 | Chronic | NOAEL | Reproduction | Chakravarty et al. (1986) in [1] | 1.0 | 1.0 | 2.00E+00 |
| | Red-winged blackbird | 6.40E-02 | 2.14E+00 | Chronic | NOAEL | Mortality | [Chlordane surrogate] | 1.0 | 1.0 | 2.14E+00 |
| | Red-winged blackbird | 6.40E-02 | 2.14E+00 | Chronic | NOAEL | Mortality | [Chlordane surrogate] | 1.0 | 1.0 | 2.14E+00 |
| | Brown pelican | 3.50E+00 | 2.80E-02 | Chronic | LOAEL | Reproduction | Anderson et al. (1975) in [1] | 1.0 | 0.1 | 2.80E-03 |
| | Brown pelican | 3.50E+00 | 2.80E-02 | Chronic | LOAEL | Reproduction | Anderson et al. (1975) in [1] | 1.0 | 0.1 | 2.80E-03 |
| | Barn owl | 4.66E-01 | 7.70E-02 | Chronic | NOAEL | Reproduction | Mendenhall et al. (1983) in [1] | 1.0 | 1.0 | 7.70E-02 |
| Endosulfan II | Gray partridge | 4.00E-01 | 1.00E+01 | Chronic | NOAEL | Reproduction | [Endosulfan surrogate] | 1.0 | 1.0 | 1.00E+01 |
| | Gray partridge | 4.00E-01 | 1.00E+01 | Chronic | NOAEL | Reproduction | [Endosulfan surrogate] | 1.0 | 1.0 | 1.00E+01 |
| | Screech owl | 1.81E-02 | 1.04E-01 | Chronic | LOAEL | Reproduction | [Endrin surrogate] | 1.0 | 0.1 | 1.04E-02 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| | None | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 8.42E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 8.42E+00 |
| | Heptachlor epoxide | None | None | None | None | None | None | None | None | No NOAEL |
| | Methoxychlor | None | None | None | None | None | None | None | None | No NOAEL |
| | Composite bird | 8.50E-01 | 8.42E+00 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 1.0 | 8.42E+00 |
| | None | None | None | None | None | None | None | None | None | No NOAEL |

Note: Footnotes appear on page 8-23.

Table 8-4. Derivation of NOAELs for Bird Test Species (continued)

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BW _i | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | NOAEL (mg/kgBW/d) Benchmark × DCF × ECF |
|---|--------------|---|--------------------------|------------------|----------|--------|--------|---|---|--|
| | | | | | | | | | | |
| Explosives | | | | | | | | | | |
| 2,4-Dinitrotoluene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 2,6-Dinitrotoluene | None | None | None | None | None | None | None | None | None | No NOAEL |
| 2,4,6-Trinitrotoluene | None | None | None | None | None | None | None | None | None | No NOAEL |
| HMX | None | None | None | None | None | None | None | None | None | No NOAEL |
| RDX | None | None | None | None | None | None | None | None | None | No NOAEL |

^a = Lead acetate.
DCF = 1 if chronic, 0.1 if subchronic (Sample, Opreko, and Suter 1996).
ECF = 1 if NOAEL, 0.1 if LOAEL (Sample, Opreko, and Suter 1996).
[1] = Sample, Opreko, and Suter 1996.
[2] = QST (1997).

The TRVs for ECOPCs derived for shrews, raccoons, and mink are shown in Table 8-5 and those for robins and green heron in Table 8-6.

The EPA Region IV ESVs for surface water used to identify ECOPCs for surface water and groundwater at the 16 SWMUs were considered to be protective of aquatic life; therefore, the preliminary risk calculations for aquatic biota exposed to surface water and groundwater at the SWMUs with aquatic habitat were not required to evaluate the preliminary assessment endpoints for aquatic receptors. The EPA Region IV ESVs for sediment used to identify ECOPCs were considered to be protective of sediment-dwelling biota; therefore, preliminary risk calculations for sediment-dwelling invertebrates exposed to sediment in creeks and ponds were not required to evaluate the assessment endpoint for sediment-dwelling invertebrates.

8.4 PRELIMINARY EXPOSURE ESTIMATE (Step iv)

For shrews and robins, which are indirectly exposed by ingestion of biota, the maximum detected concentration of each analyte in the 0-foot to-1-foot interval of the 16 SWMUs surface soil samples and drainage ditch sediment samples was used as the exposure point concentration to calculate the maximum ADDs.

For raccoons, which are directly exposed only by ingestion of surface water, and mink and herons, which are indirectly exposed only by ingestion of aquatic biota, the maximum detected concentration of each analyte in surface water grab samples was used as the exposure point concentration to calculate the maximum ADDs. The maximum concentrations of ECOPCs in sediment from SWMUs with aquatic habitats were used as the exposure point concentrations for calculation of ADDs for green herons ingesting sediment-dwelling invertebrates. The maximum detected concentration of each analyte in samples from the 16 SWMUs monitoring wells, Geoprobe, and vertical-profile locations was used as the exposure point concentration to calculate the maximum ADDs for raccoons, mink, or green herons directly or indirectly exposed to groundwater potentially discharging to surface water.

The ADD to shrews and robins from substances in surface soil was calculated as the product of the maximum detected concentration, the unitless soil-to-invertebrate bioaccumulation factor (BAF_i), and the daily specific food ingestion rate (IR) of the receptor. That is,

$$ADD \text{ (mg/kg/d)} = \text{max. soil concentration (mg/kg)} \times BAF_i \times IR \text{ (kg/kg/d)}.$$

The ADD to raccoons by ingestion of substances in surface water and groundwater was calculated as the product of the maximum detected concentration, the unit conversion factor (0.001 microgram per milligram), and the daily specific water ingestion rate (IR_w) of the receptor. That is,

$$ADD \text{ (mg/kg/d)} = \text{max. water concentration (}\mu\text{g/L)} \times 0.001 \text{ (}\mu\text{g/mg)} \times IR_w \text{ (L/kg/d)}.$$

The ADD to mink and green herons from ingestion of biota exposed to substances in surface water and groundwater was calculated as the product of the maximum detected concentration, the unitless bioconcentration factor (BCF) for the contaminant in fish tissue, and the daily specific food ingestion rate (IR) of the receptor. That is,

$$ADD \text{ (mg/kg/d)} = \text{max. water concentration (}\mu\text{g/L)} \times 0.001 \text{ (}\mu\text{g/mg)} \times BCF \times IR \text{ (kg/kg/d)}.$$

Table 8-5. Derivation of NOAELs and Screening Toxicity Reference Values for Mammal Receptors

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | Raccoon | | Short-tailed Shrew | | Mink | |
|----------------------------|--------------|---|---|--|---|--|---|--|---|
| | | | | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} |
| INORGANICS | | | | | | | | | |
| Aluminum | Mouse | 3.00E-02 | 1.93E+00 | 2.66E-01 | 5.14E-01 | 1.19E+00 | 2.30E+00 | 4.16E-01 | 8.03E-01 |
| Antimony | Mouse | 3.00E-02 | 1.25E-01 | 2.66E-01 | 3.33E-02 | 1.19E+00 | 1.49E-01 | 4.16E-01 | 5.20E-02 |
| Arsenic | Mouse | 3.00E-02 | 1.26E-01 | 2.66E-01 | 3.35E-02 | 1.19E+00 | 1.50E-01 | 4.16E-01 | 5.24E-02 |
| Barium | Rat | 4.35E-01 | 5.06E+00 | 5.19E-01 | 2.63E+00 | 2.32E+00 | 1.17E+01 | 8.12E-01 | 4.11E+00 |
| Beryllium | Rat | 3.50E-01 | 6.60E-01 | 4.92E-01 | 3.25E-01 | 2.20E+00 | 1.45E+00 | 7.69E-01 | 5.08E-01 |
| Cadmium | Rat | 3.03E-01 | 1.00E+00 | 4.74E-01 | 4.74E-01 | 2.12E+00 | 2.12E+00 | 7.42E-01 | 7.42E-01 |
| Calcium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chromium | Rat | 3.50E-01 | 2.74E+03 | 4.92E-01 | 1.35E+03 | 2.20E+00 | 6.02E+03 | 7.69E-01 | 2.11E+03 |
| Cobalt | Rat | 3.50E-01 | 1.00E+00 | 4.92E-01 | 4.92E-01 | 2.20E+00 | 2.20E+00 | 7.69E-01 | 7.69E-01 |
| Copper | Mink | 1.00E+00 | 1.17E+01 | 6.39E-01 | No NOAEL | 2.86E+00 | 3.35E+01 | 1.00E+00 | 1.17E+01 |
| Iron | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Lead | Rat | 3.50E-01 | 8.00E+00 | 4.92E-01 | 3.93E+00 | 2.20E+00 | 1.76E+01 | 7.69E-01 | 6.15E+00 |
| Magnesium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Manganese | Rat | 3.50E-01 | 8.80E+01 | 4.92E-01 | 4.33E+01 | 2.20E+00 | 1.93E+02 | 7.69E-01 | 6.77E+01 |
| Mercury | Mink | 1.00E+00 | 1.01E+00 | 6.39E-01 | 6.46E-01 | 2.86E+00 | 2.89E+00 | 1.00E+00 | 1.01E+00 |
| Nickel | Rat | 3.50E-01 | 4.00E+01 | 4.92E-01 | 1.97E+01 | 2.20E+00 | 8.79E+01 | 7.69E-01 | 3.08E+01 |
| Potassium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Selenium | Rat | 3.50E-01 | 2.00E-01 | 4.92E-01 | 9.84E-02 | 2.20E+00 | 4.40E-01 | 7.69E-01 | 1.54E-01 |
| Silver | Rat | 3.50E-01 | 1.01E+02 | 4.92E-01 | 4.98E+01 | 2.20E+00 | 2.22E+02 | 7.69E-01 | 7.78E+01 |
| Sodium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Vanadium | Rat | 2.60E-01 | 2.10E-01 | 4.57E-01 | 9.59E-02 | 2.04E+00 | 4.28E-01 | 7.14E-01 | 1.50E-01 |
| Zinc | Rat | 3.50E-01 | 1.60E+02 | 4.92E-01 | 7.87E+01 | 2.20E+00 | 3.52E+02 | 7.69E-01 | 1.23E+02 |
| ORGANICS | | | | | | | | | |
| Volatile Organic Compounds | | | | | | | | | |
| Acetone | Rat | 3.50E-01 | 1.00E+01 | 4.92E-01 | 4.92E+00 | 2.20E+00 | 2.20E+01 | 7.69E-01 | 7.69E+00 |
| Benzene | Mouse | 3.00E-02 | 2.64E+01 | 2.66E-01 | 7.02E+00 | 1.19E+00 | 3.13E+01 | 4.16E-01 | 1.10E+01 |
| Bromodichloromethane | Mouse | 3.00E-02 | 1.79E+00 | 2.66E-01 | 4.76E-01 | 1.19E+00 | 2.13E+00 | 4.16E-01 | 7.45E-01 |

Note: Footnotes appear on page 8-28.

Table 8-5. Derivation of NOAELs and Screening Toxicity Reference Values for Mammal Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | Raccoon | | Short-tailed Shrew | | Mink | |
|---------------------------|--------------|---|---|---|--|---|--|---|--|
| | | | | Body-weight Conversion Factor BW _t ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _t ^{conv} | Body-weight Conversion Factor BW _t ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _t ^{conv} | Body-weight Conversion Factor BW _t ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _t ^{conv} |
| Bromoform | Rat | 3.50E-01 | 1.79E+00 | 4.92E-01 | 8.80E-01 | 2.20E+00 | 3.93E+00 | 7.69E-01 | 1.38E+00 |
| Bromomethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Carbon disulfide | Rat | 3.50E-01 | 1.10E+01 | 4.92E-01 | 5.41E+00 | 2.20E+00 | 2.42E+01 | 7.69E-01 | 8.46E+00 |
| Carbon tetrachloride | Rat | 3.50E-01 | 1.60E+01 | 4.92E-01 | 7.87E+00 | 2.20E+00 | 3.52E+01 | 7.69E-01 | 1.23E+01 |
| Chlorobenzene | Rat | 3.50E-01 | 5.00E+01 | 4.92E-01 | 2.46E+01 | 2.20E+00 | 1.10E+02 | 7.69E-01 | 3.85E+01 |
| Chloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chloroform | Rat | 3.50E-01 | 1.50E+01 | 4.92E-01 | 7.38E+00 | 2.20E+00 | 3.30E+01 | 7.69E-01 | 1.15E+01 |
| Chloromethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Dibromochloromethane | Rat | 3.50E-01 | 2.14E+00 | 4.92E-01 | 1.05E+00 | 2.20E+00 | 4.70E+00 | 7.69E-01 | 1.65E+00 |
| 1,1-Dichloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,1-Dichloroethene | Rat | 3.50E-01 | 3.00E+01 | 4.92E-01 | 1.48E+01 | 2.20E+00 | 6.59E+01 | 7.69E-01 | 2.31E+01 |
| 1,2-Dichloroethane | Mouse | 3.50E-02 | 5.00E+01 | 2.77E-01 | 1.38E+01 | 1.24E+00 | 6.18E+01 | 4.33E-01 | 2.16E+01 |
| 1,2-Dichloroethene | Mouse | 3.00E-02 | 4.52E+01 | 2.66E-01 | 1.20E+01 | 1.19E+00 | 5.38E+01 | 4.16E-01 | 1.88E+01 |
| 1,3-Dichloropropene | Rat | 3.50E-01 | 3.00E-01 | 4.92E-01 | 1.48E-01 | 2.20E+00 | 6.59E-01 | 7.69E-01 | 2.31E-01 |
| Ethylbenzene | Rat | 3.50E-01 | 9.71E+00 | 4.92E-01 | 4.78E+00 | 2.20E+00 | 2.13E+01 | 7.69E-01 | 7.47E+00 |
| Methylene chloride | Rat | 3.50E-01 | 5.85E+00 | 4.92E-01 | 2.88E+00 | 2.20E+00 | 1.29E+01 | 7.69E-01 | 4.50E+00 |
| Methyl ethyl ketone | Rat | 3.50E-01 | 1.77E+03 | 4.92E-01 | 8.71E+02 | 2.20E+00 | 3.89E+03 | 7.69E-01 | 1.36E+03 |
| 4-Methyl-2-pentanone | Rat | 3.50E-01 | 2.50E+01 | 4.92E-01 | 1.23E+01 | 2.20E+00 | 5.49E+01 | 7.69E-01 | 1.92E+01 |
| Styrene | Dog | 1.00E+01 | 2.00E+02 | 1.14E+00 | 2.27E+02 | 5.08E+00 | 1.02E+03 | 1.78E+00 | 3.56E+02 |
| Tetrachloroethene | Mouse | 3.00E-02 | 1.40E+00 | 2.66E-01 | 3.73E-01 | 1.19E+00 | 1.66E+00 | 4.16E-01 | 5.83E-01 |
| 1,1,2,2-Tetrachloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Toluene | Mouse | 3.00E-02 | 2.60E+01 | 2.66E-01 | 6.91E+00 | 1.19E+00 | 3.09E+01 | 4.16E-01 | 1.08E+01 |
| 1,1,1-Trichloroethane | Mouse | 3.50E-02 | 1.00E+03 | 2.77E-01 | 2.77E+02 | 1.24E+00 | 1.24E+03 | 4.33E-01 | 4.33E+02 |
| 1,1,2-Trichloroethane | Mouse | 3.50E-02 | 3.90E-01 | 2.77E-01 | 1.08E-01 | 1.24E+00 | 4.82E-01 | 4.33E-01 | 1.69E-01 |
| Trichloroethene | Mouse | 3.00E-02 | 7.00E-01 | 2.66E-01 | 1.86E-01 | 1.19E+00 | 8.32E-01 | 4.16E-01 | 2.91E-01 |
| Vinyl chloride | Rat | 3.50E-01 | 1.70E-01 | 4.92E-01 | 8.36E-02 | 2.20E+00 | 3.74E-01 | 7.69E-01 | 1.31E-01 |
| Xylenes, total | Mouse | 3.00E-02 | 2.06E+00 | 2.66E-01 | 5.48E-01 | 1.19E+00 | 2.45E+00 | 4.16E-01 | 8.57E-01 |

Note: Footnotes appear on page 8-28.

Table 8-5. Derivation of NOAELs and Screening Toxicity Reference Values for Mammal Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | Raccoon | | Short-tailed Shrew | | Mink | |
|----------------------------------|--------------|---|---|--|---|--|---|--|---|
| | | | | Body-weight Conversion Factor BW ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW ^{conv} | Body-weight Conversion Factor BW ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW ^{conv} | Body-weight Conversion Factor BW ^{conv} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW ^{conv} |
| -Semi-volatile Organic Compounds | | | | | | | | | |
| Acenaphthene | Mouse | 3.00E-02 | 1.75E+02 | 2.66E-01 | 4.66E+01 | 1.19E+00 | 2.08E+02 | 4.16E-01 | 7.28E+01 |
| Benzo(a)anthracene | Mouse | 3.00E-02 | 1.33E+01 | 2.66E-01 | 3.54E+00 | 1.19E+00 | 1.58E+01 | 4.16E-01 | 5.54E+00 |
| Benzo(a)pyrene | Mouse | 3.00E-02 | 1.00E+00 | 2.66E-01 | 2.66E-01 | 1.19E+00 | 1.19E+00 | 4.16E-01 | 4.16E-01 |
| Benzo(b)fluoranthene | Mouse | 3.00E-02 | 1.33E+01 | 2.66E-01 | 3.54E+00 | 1.19E+00 | 1.58E+01 | 4.16E-01 | 5.54E+00 |
| Benzo(g,h,i)perylene | Mouse | 3.00E-02 | 1.33E+01 | 2.66E-01 | 3.54E+00 | 1.19E+00 | 1.58E+01 | 4.16E-01 | 5.54E+00 |
| Benzo(k)fluoranthene | Mouse | 3.00E-02 | 1.00E+00 | 2.66E-01 | 2.66E-01 | 1.19E+00 | 1.19E+00 | 4.16E-01 | 4.16E-01 |
| Bis(2-ethylhexyl)phthalate | Mouse | 3.00E-02 | 1.83E+01 | 2.66E-01 | 4.87E+00 | 1.19E+00 | 2.18E+01 | 4.16E-01 | 7.61E+00 |
| Chlorobenzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chrysene | Mouse | 3.00E-02 | 1.33E+01 | 2.66E-01 | 3.54E+00 | 1.19E+00 | 1.58E+01 | 4.16E-01 | 5.54E+00 |
| Dibenzofuran | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,2-Dichlorobenzene | Rat | 3.50E-01 | 1.20E+02 | 4.92E-01 | 5.90E+01 | 2.20E+00 | 2.64E+02 | 7.69E-01 | 9.23E+01 |
| 2,4-Dimethylphenol | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Di-N-butylphthalate | Mouse | 3.00E-02 | 5.50E+02 | 2.66E-01 | 1.46E+02 | 1.19E+00 | 6.54E+02 | 4.16E-01 | 2.29E+02 |
| Di-N-octylphthalate | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Fluoranthene | Mouse | 3.00E-02 | 5.00E+01 | 2.66E-01 | 1.33E+01 | 1.19E+00 | 5.95E+01 | 4.16E-01 | 2.08E+01 |
| 2-Hexanone | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Indeno(1,2,3-cd)pyrene | Mouse | 3.00E-02 | 1.33E+01 | 2.66E-01 | 3.54E+00 | 1.19E+00 | 1.58E+01 | 4.16E-01 | 5.54E+00 |
| 2-Methylnaphthalene | Rat | 3.50E-01 | 5.00E+00 | 4.92E-01 | 2.46E+00 | 2.20E+00 | 1.10E+01 | 7.69E-01 | 3.85E+00 |
| 4-Chloro-3-methylphenol | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Naphthalene | Rat | 3.50E-01 | 5.00E+00 | 4.92E-01 | 2.46E+00 | 2.20E+00 | 1.10E+01 | 7.69E-01 | 3.85E+00 |
| N-Nitrosodiphenylamine | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Phenanthrene | Mouse | 3.00E-02 | 1.00E+00 | 2.66E-01 | 2.66E-01 | 1.19E+00 | 1.19E+00 | 4.16E-01 | 4.16E-01 |
| Phenol | Rat | 3.50E-01 | 6.00E+00 | 4.92E-01 | 2.95E+00 | 2.20E+00 | 1.32E+01 | 7.69E-01 | 4.61E+00 |
| Pyrene | Mouse | 3.00E-02 | 1.00E+00 | 2.66E-01 | 2.66E-01 | 1.19E+00 | 1.19E+00 | 4.16E-01 | 4.16E-01 |

Note: Footnotes appear on page 8-28.

Table 8-5. Derivation of NOAELs and Screening Toxicity Reference Values for Mammal Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | Raccoon | | Short-tailed Shrew | | Mink | |
|----------------------|--------------|---|---|--|---|--|---|--|---|
| | | | | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t /BW) ^{0.25} | NOAEL (mg/kgBW/d) NOAEL _t × BW _{conv} |
| Pesticides/PCBs | | | | | | | | | |
| Aldrin | Rat | 3.50E-01 | 2.00E-01 | 4.92E-01 | 9.84E-02 | 2.20E+00 | 4.40E-01 | 7.69E-01 | 1.54E-01 |
| alpha-BHC | Rat | 3.50E-01 | 1.60E+00 | 4.92E-01 | 7.87E-01 | 2.20E+00 | 3.52E+00 | 7.69E-01 | 1.23E+00 |
| beta-BHC | Rat | 3.50E-01 | 4.00E-01 | 4.92E-01 | 1.97E-01 | 2.20E+00 | 8.80E-01 | 7.69E-01 | 3.08E-01 |
| delta-BHC | Rat | 3.50E-01 | 1.60E+00 | 4.92E-01 | 7.87E-01 | 2.20E+00 | 3.52E+00 | 7.69E-01 | 1.23E+00 |
| gamma-BHC (Lindane) | Rat | 3.50E-01 | 8.00E+00 | 4.92E-01 | 3.94E+00 | 2.20E+00 | 1.76E+01 | 7.69E-01 | 6.15E+00 |
| alpha-Chlordane | Mouse | 3.00E-02 | 4.58E+00 | 2.66E-01 | 1.22E+00 | 1.19E+00 | 5.45E+00 | 4.16E-01 | 1.91E+00 |
| gamma-Chlordane | Mouse | 3.00E-02 | 4.58E+00 | 2.66E-01 | 1.22E+00 | 1.19E+00 | 5.45E+00 | 4.16E-01 | 1.91E+00 |
| 4,4'-DDD | Rat | 3.50E-01 | 8.00E-01 | 4.92E-01 | 3.93E-01 | 2.20E+00 | 1.76E+00 | 7.69E-01 | 6.15E-01 |
| 4,4'-DDE | Rat | 3.50E-01 | 8.00E-01 | 4.92E-01 | 3.93E-01 | 2.20E+00 | 1.76E+00 | 7.69E-01 | 6.15E-01 |
| 4,4'-DDT | Rat | 3.50E-01 | 8.00E-01 | 4.92E-01 | 3.93E-01 | 2.20E+00 | 1.76E+00 | 7.69E-01 | 6.15E-01 |
| Dieldrin | Rat | 3.50E-01 | 2.00E-02 | 4.92E-01 | 9.84E-03 | 2.20E+00 | 4.40E-02 | 7.69E-01 | 1.54E-02 |
| Endosulfan II | Rat | 3.50E-01 | 1.50E-01 | 4.92E-01 | 7.38E-02 | 2.20E+00 | 3.30E-01 | 7.69E-01 | 1.15E-01 |
| Endosulfan sulfate | Rat | 3.50E-01 | 1.50E-01 | 4.92E-01 | 7.38E-02 | 2.20E+00 | 3.30E-01 | 7.69E-01 | 1.15E-01 |
| Endrin ketone | Mouse | 3.00E-02 | 9.20E-02 | 2.66E-01 | 2.45E-02 | 1.19E+00 | 1.09E-01 | 4.16E-01 | 3.83E-02 |
| Heptachlor | Mink | 1.00E+00 | 1.00E-01 | 6.39E-01 | 6.39E-02 | 2.86E+00 | 2.86E-01 | 1.00E+00 | 1.00E-01 |
| Heptachlor epoxide | Mink | 1.00E+00 | 1.00E-01 | 6.39E-01 | 6.39E-02 | 2.86E+00 | 2.86E-01 | 1.00E+00 | 1.00E-01 |
| Methoxychlor | Rat | 3.50E-01 | 4.00E+00 | 4.92E-01 | 1.97E+00 | 2.20E+00 | 8.79E+00 | 7.69E-01 | 3.08E+00 |
| 2,4-Dinitrotoluene | Mouse | 3.00E-02 | 1.35E+01 | 2.66E-01 | 3.59E+00 | 1.19E+00 | 1.61E+01 | 4.16E-01 | 5.62E+00 |
| 2,6-Dinitrotoluene | Rat | 3.50E-01 | 7.00E-01 | 4.92E-01 | 3.44E-01 | 2.20E+00 | 1.54E+00 | 7.69E-01 | 5.38E-01 |
| 2,4,6-Dinitrotoluene | Rat | 3.50E-01 | 1.60E+00 | 4.92E-01 | 7.87E-01 | 2.20E+00 | 3.52E+00 | 7.69E-01 | 1.23E+00 |
| HMX | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| RDX | None | None | No NOAEL | None | No NOAEL | None | No NOAEL | None | No NOAEL |

BW(kg) Raccoon = 5.98;

BW (kg) Shrew = 0.015.

BW (kg) Mink = 1.0.

Table 8-6. Derivation of NOAELs and Screening Toxicity Reference Values for Bird Receptors

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | American Robin | | Green Heron | |
|------------|---------------------|--|---|---|--|---|--|
| | | | | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW ₀) | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} ⁰ | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW ₀) | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} ⁰ |
| INORGANICS | | | | | | | |
| Aluminum | Ringed dove | 1.55E-01 | 1.10E+02 | 1.00E+00 | 1.10E+02 | 1.00E+00 | 1.10E+02 |
| Antimony | Composite bird | 8.50E-01 | 3.57E-01 | 1.00E+00 | 3.57E-01 | 1.00E+00 | 3.57E-01 |
| Arsenic | Mallard duck | 1.00E+00 | 5.14E+00 | 1.00E+00 | 5.14E+00 | 1.00E+00 | 5.14E+00 |
| Barium | Chick (14 days old) | 1.21E-01 | 2.08E+01 | 1.00E+00 | 2.08E+01 | 1.00E+00 | 2.08E+01 |
| Beryllium | Composite Bird | 8.50E-01 | 1.67E+00 | 1.00E+00 | 1.67E+00 | 1.00E+00 | 1.67E+00 |
| Cadmium | Mallard duck | 1.15E+00 | 1.45E+00 | 1.00E+00 | 1.45E+00 | 1.00E+00 | 1.45E+00 |
| Calcium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chromium | Black duck | 1.25E+00 | 1.00E+00 | 1.00E+00 | 1.00E+00 | 1.00E+00 | 1.00E+00 |
| Cobalt | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Copper | Chick (5 weeks old) | 5.34E-01 | 4.70E+01 | 1.00E+00 | 4.70E+01 | 1.00E+00 | 4.70E+01 |
| Iron | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Lead | Quail | 1.50E-01 | 1.13E+00 | 1.00E+00 | 1.13E+00 | 1.00E+00 | 1.13E+00 |
| Magnesium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Manganese | Quail | 7.20E-02 | 9.77E+02 | 1.00E+00 | 9.77E+02 | 1.00E+00 | 9.77E+02 |
| Mercury | Quail | 1.50E-01 | 4.50E-01 | 1.00E+00 | 4.50E-01 | 1.00E+00 | 4.50E-01 |
| Nickel | Mallard duckling | 7.82E-01 | 7.74E+01 | 1.00E+00 | 7.74E+01 | 1.00E+00 | 7.74E+01 |
| Potassium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Selenium | Mallard duck | 1.00E+00 | 5.00E-01 | 1.00E+00 | 5.00E-01 | 1.00E+00 | 5.00E-01 |
| Silver | Composite bird | 8.50E-01 | 5.79E+01 | 1.00E+00 | 5.79E+01 | 1.00E+00 | 5.79E+01 |
| Sodium | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Vanadium | Mallard duck | 1.17E+00 | 1.14E+01 | 1.00E+00 | 1.14E+01 | 1.00E+00 | 1.14E+01 |
| Zinc | Leghorn chicken | 1.94E+00 | 1.45E+01 | 1.00E+00 | 1.45E+01 | 1.00E+00 | 1.45E+01 |

Note: Footnotes appear on page 8-32.

Table 8-6. Derivation of NOAELs and Screening Toxicity Reference Values for Bird Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | American Robin | | Green Heron | |
|----------------------------|--------------|---|---|---|---|---|---|
| | | | | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW) ⁰ | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW) ⁰ | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} |
| ORGANICS | | | | | | | |
| Volatile Organic Compounds | | | | | | | |
| Acetone | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Benzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Bromodichloromethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Bromoform | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Bromomethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Carbon disulfide | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Carbon tetrachloride | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chlorobenzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chloroform | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chloromethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Dibromochloromethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,1-Dichloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,1-Dichloroethene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,2-Dichloroethane | Chicken | 1.60E+00 | 1.72E+01 | 1.00E+00 | 1.72E+01 | 1.00E+00 | 1.72E+01 |
| 1,2-Dichloroethene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,3-Dichloropropene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Ethylbenzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Methylene chloride | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Methyl ethyl ketone | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 4-Methyl-2-pentanone | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Styrene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Tetrachloroethene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,1,2,2-Tetrachloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Toluene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,1,1-Trichloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |

Note: Footnotes appear on pages 8-22

Note: Footnotes appear on page 8-32.

Table 8-6. Derivation of NOAELs and Screening Toxicity Reference Values for Bird Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | American Robin | | Green Heron | |
|---------------------------------------|----------------|---|---|--|--|--|--|
| | | | | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW ₀) | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} ⁰ | Body-weight Conversion Factor BW _{conv} ⁰ (BW _t / BW ₀) | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} ⁰ |
| 1,1,2-Trichloroethane | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Trichloroethene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Vinyl chloride | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Xylenes, total | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| <i>Semivolatile Organic Compounds</i> | | | | | | | |
| Acenaphthene | Composite bird | 8.50E-01 | 8.78E+01 | 1.00E+00 | 8.78E+01 | 1.00E+00 | 8.78E+01 |
| Benzo(a)anthracene | Composite bird | 8.50E-01 | 1.24E+01 | 1.00E+00 | 1.24E+01 | 1.00E+00 | 1.24E+01 |
| Benzo(a)pyrene | Composite bird | 8.50E-01 | 9.97E+00 | 1.00E+00 | 9.97E+00 | 1.00E+00 | 9.97E+00 |
| Benzo(b)fluoranthene | Composite bird | 8.50E-01 | 1.24E+01 | 1.00E+00 | 1.24E+01 | 1.00E+00 | 1.24E+01 |
| Benzo(g,h,i)perylene | Composite bird | 8.50E-01 | 1.24E+01 | 1.00E+00 | 1.24E+01 | 1.00E+00 | 1.24E+01 |
| Benzo(k)fluoranthene | Composite bird | 8.50E-01 | 9.97E+00 | 1.00E+00 | 9.97E+00 | 1.00E+00 | 9.97E+00 |
| Bis(2-ethylhexyl)phthalate | Ringed dove | 1.55E-01 | 1.10E+00 | 1.00E+00 | 1.10E+00 | 1.00E+00 | 1.10E+00 |
| Chlorobenzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Chrysene | Composite bird | 8.50E-01 | 1.24E+01 | 1.00E+00 | 1.24E+01 | 1.00E+00 | 1.24E+01 |
| Dibenzofuran | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 1,2-Dichlorobenzene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 2,4-Dimethylphenol | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Di-N-butylphthalate | Ringed dove | 1.55E-01 | 1.11E-01 | 1.00E+00 | 1.11E-01 | 1.00E+00 | 1.11E-01 |
| Di-N-octylphthalate | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Fluoranthene | Composite bird | 8.50E-01 | 1.95E+02 | 1.00E+00 | 1.95E+02 | 1.00E+00 | 1.95E+02 |
| 2-Hexanone | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Indeno(1,2,3-cd)pyrene | Composite bird | 8.50E-01 | 1.24E+01 | 1.00E+00 | 1.24E+01 | 1.00E+00 | 1.24E+01 |
| 2-Methylnaphthalene | Composite bird | 8.50E-01 | 3.39E+01 | 1.00E+00 | 3.39E+01 | 1.00E+00 | 3.39E+01 |
| 4-Chloro-3-methylphenol | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Naphthalene | Composite bird | 8.50E-01 | 3.39E+01 | 1.00E+00 | 3.39E+01 | 1.00E+00 | 3.39E+01 |
| N-Nitrosodiphenylamine | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Phenanthrene | Composite bird | 8.50E-01 | 9.97E+00 | 1.00E+00 | 9.97E+00 | 1.00E+00 | 9.97E+00 |
| Phenol | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Pyrene | Composite bird | 8.50E-01 | 9.97E+00 | 1.00E+00 | 9.97E+00 | 1.00E+00 | 9.97E+00 |

Note: Footnotes appear on page 8-32.

Table 8-6. Derivation of NOAELs and Screening Toxicity Reference Values for Bird Receptors (continued)

| Analyte | Test Species | Test Species Body Weight (kg) BW _t | Test Species NOAEL _t (mg/kgBW/d) | American Robin | | Green Heron | |
|-----------------------|----------------------|--|---|---|---|---|---|
| | | | | Body-weight Conversion Factor BW _{conv} (BW _t / BW) ⁰ | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} | Body-weight Conversion Factor BW _{conv} (BW _t / BW) ⁰ | NOAEL (mg/kgBW/d) NOAEL _t x BW _{conv} |
| Pesticides/PCBs | | | | | | | |
| Aldrin | Composite bird | 8.50E-01 | 1.73E+00 | 1.00E+00 | 1.73E+00 | 1.00E+00 | 1.73E+00 |
| alpha-BHC | Japanese quail | 1.50E-01 | 5.63E-01 | 1.00E+00 | 5.63E-01 | 1.00E+00 | 5.63E-01 |
| beta-BHC | Japanese quail | 1.50E-01 | 5.63E-01 | 1.00E+00 | 5.63E-01 | 1.00E+00 | 5.63E-01 |
| delta-BHC | Japanese quail | 1.50E-01 | 5.63E-01 | 1.00E+00 | 5.63E-01 | 1.00E+00 | 5.63E-01 |
| gamma-BHC (Lindane) | Mallard duck | 1.00E+00 | 2.00E+00 | 1.00E+00 | 2.00E+00 | 1.00E+00 | 2.00E+00 |
| alpha-Chlordane | Red-winged blackbird | 6.40E-02 | 2.14E+00 | 1.00E+00 | 2.14E+00 | 1.00E+00 | 2.14E+00 |
| gamma-Chlordane | Red-winged blackbird | 6.40E-02 | 2.14E+00 | 1.00E+00 | 2.14E+00 | 1.00E+00 | 2.14E+00 |
| 4,4'-DDD | Brown pelican | 3.50E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 |
| 4,4'-DDE | Brown pelican | 3.50E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 |
| 4,4'-DDT | Brown pelican | 3.50E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 | 1.00E+00 | 2.80E-03 |
| Dieldrin | Barn owl | 4.66E-01 | 7.70E-02 | 1.00E+00 | 7.70E-02 | 1.00E+00 | 7.70E-02 |
| Endosulfan II | Gray partridge | 4.00E-01 | 1.00E+01 | 1.00E+00 | 1.00E+01 | 1.00E+00 | 1.00E+01 |
| Endosulfan sulfate | Gray partridge | 4.00E-01 | 1.00E+01 | 1.00E+00 | 1.00E+01 | 1.00E+00 | 1.00E+01 |
| Endrin ketone | Screech owl | 1.81E-02 | 1.04E-02 | 1.00E+00 | 1.04E-02 | 1.00E+00 | 1.04E-02 |
| Heptachlor | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Heptachlor epoxide | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| Methoxychlor | Composite bird | 8.50E-01 | 8.42E+00 | 1.00E+00 | 8.42E+00 | 1.00E+00 | 8.42E+00 |
| Explosives | | | | | | | |
| 2,4-Dinitrotoluene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 2,6-Dinitrotoluene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| 2,4,6-Trinitrotoluene | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| HMX | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |
| RDX | None | None | No NOAEL | None | No NOAEL | None | No NOAEL |

BW (kg) Robin = 0.08

BW (kg) Robin = 0.08.

BW (kg) Green heron = 0.25.

The ADD to green herons from ingestion of invertebrates exposed to substances in sediment was calculated as the product of the maximum detected concentration, the unitless bioaccumulation factor (BAF_i) for the contaminant in invertebrate tissue, and the daily specific water ingestion rate (IR_w) of the receptor. That is,

$$\text{ADD (mg/kg/d)} = \text{max. sediment concentration (mg/kg)} \times \text{BAF}_i \times \text{IR (kg/kg/d)}.$$

The exposure parameters for shrews and robins exposed to substances in surface soil and raccoons, mink, and green herons exposed to ECOPCs in surface water, sediment, or groundwater are presented in Table 8-7.

Table 8-7. Exposure Parameters for Surrogate Species

| Parameter | Surrogate Species | | | | |
|-------------------------------|-----------------------|-----------------------|----------------------------|----------------------------|---|
| | Shrew | Robin | Raccoon | Mink | Green Heron |
| Body weight (kg) | 0.015 ^a | 0.077 ^a | 4.31 ^b | 1 ^a | 0.25 ^c |
| Food ingestion rate (kg/kg/d) | 0.6 ^d | 1.2 ^d | — | 0.137 ^d | 0.192 ^c |
| Water ingestion rate (L/kg/d) | — | — | 0.08 ^b | — | — |
| Area use factor | 1 | 1 | 1 | 1 | 1 |
| Relative bioavailability | 100 percent | 100 percent | 100 percent | 100 percent | 100 percent |
| Diet | 100 percent earthworm | 100 percent earthworm | — | 100 percent fish | 100 percent fish (for surface water and groundwater); 100 percent sediment-dwelling invertebrates (for sediment) |
| Source medium | Surface soil | Surface soil | Surface water; groundwater | Surface water; groundwater | Surface water; groundwater; sediment |

^aSample, Opresko, and Suter (1996), Table B.1.

^bEPA 1993.

^cEPA Region IV Supplemental Guidance to RAGS (EPA 1996d), Table A.

^dConverted from values reported as kilograms per day in Sample, Opresko, and Suter (1996) by dividing by body weight (kilograms).

— = Not required for preliminary risk calculation.

The exposures of surrogate species were estimated using conservative assumptions (Table 8-7). It was assumed that the receptors spend their entire lives and obtain 100 percent of their diet or drinking water at the facility [i.e., the area use factor (AUF) equals one]. Shrews and robins were assumed to eat only soil-dwelling invertebrates such as worms that bioaccumulate contaminants from soil, in accordance with EPA Region IV requirements that the screening be based on exposure through two trophic transfers (EPA 1997a). Raccoons were assumed to drink only water from the creeks or ponds at the individual SWMUs but were assumed to eat no solid food from the SWMU. Mink were assumed to eat only fish from creeks and ponds. Herons were assumed to eat only fish when evaluating surface water and groundwater, and only sediment-dwelling invertebrates when evaluating sediment. Chemicals in surface soil and sediment were assumed to bioaccumulate in the soil- and sediment-dwelling invertebrate prey of ecological receptors at levels equal to published bioaccumulation factors (BAFs) for earthworms and other invertebrates as reported in Hazardous Waste Remedial Actions Program (HAZWRAP) (1994).

Chemicals in surface water and groundwater were assumed to bioaccumulate in aquatic biota at levels equal to published BCFs for fish (HAZWRAP 1994).

8.5 PRELIMINARY RISK CALCULATION (Step v)

The preliminary risk calculation (Step v) uses HQs, the ratio of the ADD calculated using the measured maximum concentration and the TRV, to evaluate the potential for risk. The HQs of ECOPCs with consistent modes of toxicity and effects endpoints are added to produce an HI. An HI greater than one for a category of COPCs is a useful indicator of potential risk when no individual COPC in that category has an HQ greater than one. An HI assumes that the effects of the individual COPCs in the category are additive. Metals are assumed to have distinct modes of toxicity and effects endpoints. Therefore, HIs are calculated only for VOCs, SVOCs, and pesticides/PCBs when no individual ECOPC has an HQ greater than one and when HQs are calculated for more than one chemical.

Because of the uncertainties in quantifying exposure and effects, the exposure and effects assessments for each of the SWMUs were designed to produce HQs that minimized the probability of falsely concluding that there was no risk when in fact there was. Therefore, ECOPCs with HQs and HIs less than one indicated little to no likelihood of risk to the ecological receptors. To minimize the probability of falsely concluding there was risk when there was none, an ERA using site-specific data for those ECOPCs with calculated HQs or HIs exceeding one was recommended (GEPD 1996).

The HQs and HIs for ECOPCs in the relevant media at each of the 16 SWMUs are presented in the Chapters 9.0 and 10.0. Conclusions and recommendations are presented in Chapter 11.0.

8.6 UNCERTAINTIES

The EPRE for the 16 SWMUs was designed to minimize the probability of falsely concluding that there was no risk when in fact there was risk (GEPD 1996). The EPA Region IV ESV comparison was designed to be conservative and to screen out only those substances for which there was little probability that they would pose a hazard to ecological receptors. The preliminary ecological effects and exposure assessments were designed to produce preliminary risk calculations that overestimated a risk. Using conservative exposure and effects assumptions (e.g., AUF equal to one, maximum detected concentrations, and TRVs based on NOAELs), as required by the guidance documents (GEPD 1996; EPA 1997a), overestimates risk to ecological receptors. Therefore, contaminants with HQs and HIs less than or equal to one indicate little to no likelihood of hazard to the ecological receptors.

Because of the conservatism of the TRVs and exposure estimates, HQs and HIs exceeding one do not necessarily mean that the ecological receptors are at risk of ecologically significant adverse effect. ECOPCs with small HQs may not be potential hazards unless the receptors obtain all of their food from the individual SWMU and the TRVs closely approximate the actual toxicity threshold. ECOPCs with large HQs may not be potential hazards if the TRVs for those contaminants greatly overestimate the toxicity to the receptors at the 16 SWMUs. TRVs can overestimate toxicity because of differences in the form of the chemical tested, the means by which the chemical was administered to the test species, or if the test endpoint is not expected to cause a significant ecological effect. Most ECOPCs at the 16 SWMUs that are judged not to be potential hazards have small HQs for ecological receptors that are unlikely to obtain 100 percent of their food from the SWMUs. Aluminum and lead are ECOPCs that have TRVs that probably overestimate toxicity to ecological receptors at the 16 SWMUs; therefore, aluminum and lead are not considered to be potential hazards where HQs for terrestrial receptors are less than 10.

The published NOAEL for aluminum is based on a laboratory study that orally administered aluminum salts dissolved in water to the test species (Sample, Opresko, and Suter 1996). The uptake of aluminum dissolved in water overestimates the uptake from ingested tissue and especially soil. In addition, only one dose was administered, and the observed effect on the test species was of questionable ecological significance to wildlife populations. The effect endpoint for aluminum was growth rates (Ondreika, Ginter, and Kortus 1966). The NOAEL, estimated as one-tenth the nonlethal LOAEL, overestimates the potential for adverse effects to endpoint receptor populations because individual growth rates are not directly linked to reduced population sizes. The bird TRV for aluminum is based on a NOAEL for aluminum sulfate, $Al_2(SO_4)_3$ (Carriere et al. 1986). No LOAEL was observed, and aluminum is unlikely to occur as a sulfate in natural soil with clay minerals. The TRV for aluminum probably overestimates risk to ecological receptors by several orders of magnitude.

The TRV for lead is the observed NOAEL for rats exposed to lead acetate over three generations, resulting in sublethal effects on offspring, namely reduced offspring weights and kidney damage (Azar, Trochimowicz, and Maxwell 1973). These are not necessarily ecologically significant, so a TRV based on a NOAEL is perhaps an unreasonable basis for characterizing risk. The bird TRV for lead is based on a NOAEL for lead acetate; the observed LOAEL was 10-fold larger (Edens et al. 1976). Furthermore, it is unlikely that 100 percent of the lead in surface soil is in the form of lead acetate. The conservative default BAF value of 0.4 for lead probably overestimates bioaccumulation by a factor of five or more.

In addition to conservative exposure and effects parameters used in the preliminary risk calculations, groundwater was treated as surface water in the EPRE in accordance with EPA Region IV guidance (EPA 1996d), which is a conservative treatment of groundwater. Groundwater is not expected to rapidly migrate away from some SWMUs because of soil characteristics and hydraulic gradients. The concentration of ECOPCs in groundwater might, therefore, decline to safe levels by adsorption or biodegradation before the groundwater discharges to the nearest aquatic habitat with biota. ECOPCs in groundwater at some SWMUs, therefore, might not pose a hazard to aquatic and terrestrial receptors. At some Phase II SWMUs, the likelihood that groundwater discharges to nearby aquatic habitats was considered when evaluating the potential for ECOPCs in groundwater to pose a hazard to aquatic and terrestrial receptors. If the data indicated that groundwater discharge was unlikely, then groundwater ECOPCs were judged not to be potential hazards.

To evaluate the uncertainty associated with ECOPCs that are judged to pose a potential risk to wildlife receptors, supplemental risk calculations are made using realistic diets, site-specific AUFs, mean sample concentrations, and LOAEL-based TRVs. Supplemental risk calculations are made for shrews and robins exposed to ECOPCs in surface soil; raccoons, mink, and green herons exposed to ECOPCs in surface water or groundwater potentially discharging to surface water; and green herons or raccoons exposed to ECOPCs in sediment. Raccoons are evaluated instead of green herons when an ECOPC has no TRV for birds but does have a TRV for mammals. Supplemental risk estimates are calculated for only those ECOPCs that are present at maximum concentrations resulting in maximum exposures to a wildlife receptor that exceed the NOAEL-based TRVs (HQ of greater than 1) or for which there are no TRVs for either mammals or birds.

Supplemental risk calculations for shrews and robins are made using published dietary fractions of plant tissue, animal tissue, and incidental soil (Table 8-8) instead of 100 percent earthworms. Evaluating the exposure from ingested plants requires uptake factors, similar to BAFs for animal tissue. Uptake factors for vegetative parts of plants are required for shrews, and factors for reproductive parts of plants are required for robins. Plant uptake factors for inorganic substances are those reported in Baes et al. (1984) converted to a wet-weight basis assuming 80 percent water content and for organic compounds are those reported in HAZWRAP (1994), which are already reported on a wet-weight basis. Means of measured

concentrations instead of maximum concentrations are used to estimate exposure for the supplemental risk calculations. Modeled maximum groundwater concentrations predicted to occur at a distance equal to the distance to the nearest surface water body are also considered in the uncertainty evaluation.

In addition to the adjustments to the exposure estimates, TRVs based on observed or estimated LOAELs instead of NOAELs are used to calculate supplemental HQs. LOAELs reported in Sample, Opresko, and Suter (1996) and other sources (Tables 8-9 and 8-10) are used to derive body-weight-adjusted TRVs (Tables 8-11 and 8-12) for the supplemental evaluations. For those ECOPCs for which there are no NOAEL-based TRVs for any receptor, toxicity data for surrogate chemicals are used to derive LOAEL-based TRVs for use in the supplemental risk calculations. The supplemental risk calculations are presented in the uncertainty discussions in the subsections for the individual SWMUs.

Table 8-9. Derivation of LOAEL Toxicity Reference Values for Mammal Test Species

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BWt | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | TRV (mg/kgBW/d) Benchmark \times DCF \times ECF |
|---|--------------|-----------------------------------|-----------------------|---------------|----------|-----------------|--|----------------------------------|----------------------------------|---|
| INORGANICS | | | | | | | | | | |
| Cadmium | Rat | 3.03E-01 | 1.00E+01 | Chronic | LOAEL | Reproduction | Sutou et al. (1980b) in [1] | 1.0 | 1.0 | 1.00E+01 |
| Chromium | Rat | 3.50E-01 | 2.74E+03 | Chronic | NOAEL | Reproduction | Ivankovic and Preussmann (1975) in [1] | 1.0 | 10.0 | 2.74E+04 |
| Lead | Rat | 3.50E-01 | 8.00E+01 | Chronic | LOAEL | Reproduction | Azar et al. (1973) in [1] | 1.0 | 1.0 | 8.00E+01 |
| Mercury | Mink | 1.00E-00 | 1.01E-00 | Chronic | NOAEL | Reproduction | Aulerich et al. (1974) in [1] | 1.0 | 10.0 | 1.01E+01 |
| ORGANICS | | | | | | | | | | |
| <i>Volatile Organic Compounds</i> | | | | | | | | | | |
| Acetone | Rat | 3.50E-01 | 5.00E+01 | Subchronic | LOAEL | Hepatotoxicity | EPA (1986e) in [1] | 0.1 | 1.0 | 5.00E-00 |
| Chloroethane | Rat | 3.50E-01 | 1.80E+02 | Subchronic | LOAEL | Mortality | Value for chloromethane in [2] | 0.1 | 1.0 | 1.80E+01 |
| Chloromethane | Rat | 3.50E-01 | 1.80E+02 | Subchronic | LOAEL | Mortality | LD ₅₀ reported in [2] | 0.1 | 1.0 | 1.80E+01 |
| Methylene chloride | Rat | 3.50E-01 | 5.00E+01 | Chronic | LOAEL | Liver histology | NCA (1982) in [1] | 1.0 | 1.0 | 5.00E+01 |
| Vinyl chloride | Rat | 3.50E-01 | 1.70E-00 | Chronic | LOAEL | Mortality | Feron et al. (1981) in [1] | 1.0 | 1.0 | 1.70E-00 |
| <i>Semivolatile Organic Compounds</i> | | | | | | | | | | |
| Benzo(b)fluoranthene | Mouse | 3.00E-02 | 1.33E+01 | Chronic | NOAEL | None | Neal and Rigdon (1967) in [3] | 1.0 | 10.0 | 1.33E+02 |
| <i>Pesticides/PCBs</i> | | | | | | | | | | |
| 4,4'-DDE | Rat | 3.50E-01 | 4.00E-00 | Chronic | LOAEL | Reproduction | Value from 4,4'-DDT in [1] | 1.0 | 1.0 | 4.00E-00 |

DCF = 1 if chronic, 0.1 if subchronic (Sample, Opreko, and Suter 1996).

ECF = 10 if NOAEL, 1.0 if LOAEL (Sample, Opreko, and Suter 1996).

[1] = Sample, Opreko, and Suter (1996).

[2] = Registry of Toxic Effects of Chemical Substances, downloaded in January 1999.

[3] = QST (1997).

Table 8-10. Derivation of LOAEL Toxicity Reference Values for Bird Test Species

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight (kg) BWt | Benchmark (mg/kgBW/d) | Test Duration | Endpoint | Effect | Source | Duration Conversion Factor (DCF) | Endpoint Conversion Factor (ECF) | TRV (mg/kgBW/d) Benchmark × DCF × ECF |
|---|----------------|-----------------------------------|-----------------------|---------------|----------|--------------|-----------------------------------|----------------------------------|----------------------------------|---------------------------------------|
| INORGANICS | | | | | | | | | | |
| Cadmium | Mallard duck | 1.15E-00 | 2.00E+01 | Chronic | LOAEL | Reproduction | White and Finley (1978) in [1] | 1.0 | 1.0 | 2.00E+01 |
| Chromium | Black duck | 1.25E-00 | 5.00E-00 | Chronic | LOAEL | Reproduction | Haseltine et al. (unpubl.) in [1] | 1.0 | 1.0 | 5.00E-00 |
| Lead | Quail | 1.50E-01 | 1.13E+01 | Chronic | LOAEL | Reproduction | Edens et al. (1976) in [1] | 1.0 | 1.0 | 1.13E+01 |
| Mercury | Quail | 1.50E-01 | 9.00E-01 | Chronic | LOAEL | Reproduction | Hill and Schaffner (1976) in [1] | 1.0 | 1.0 | 9.00E-01 |
| ORGANICS | | | | | | | | | | |
| <i>Volatile Organic Compounds</i> | | | | | | | | | | |
| Acetone | None | None | None | None | None | None | None | None | None | No TRV |
| Chloroethane | None | None | None | None | None | None | None | None | None | No TRV |
| Chloromethane | None | None | None | None | None | None | None | None | None | No TRV |
| Methylene chloride | None | None | None | None | None | None | None | None | None | No TRV |
| Vinyl chloride | None | None | None | None | None | None | None | None | None | No TRV |
| <i>Semivolatile Organic Compounds</i> | | | | | | | | | | |
| Benzo(b)fluoranthene | Composite bird | 8.50E-01 | 1.24E+01 | Chronic | NOAEL | None | Shortelle et al. (1997) in [2] | 1.0 | 10.0 | 1.24E+02 |
| <i>Pesticides/PCBs</i> | | | | | | | | | | |
| 4,4'-DDE | Brown pelican | 3.50E-00 | 2.80E-02 | Chronic | LOAEL | Reproduction | Value from 4,4'-DDT in [1] | 1.0 | 1.0 | 2.80E-02 |

DCF = Duration conversion factor; i if chronic, 0.1 if subchronic (Sample, Opreko, and Suter 1996).

ECF = Endpoint conversion factor; 10 if NOAEL, 1.0 if LOAEL (Sample, Opreko, and Suter 1996).

[1] = Sample, Opreko, and Suter (1996).

[2] = QST (1997).

Table 8-11. Derivation of LOAEL Toxicity Reference Values for Mammal Receptors

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight BWt (kg) | TRV _t (mg/kgBW/d) | Raccoon | | Short-tailed Shrew | | Mink | |
|---|-----------------|---|---------------------------------|---|---|---|---|---|---|
| | | | | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t / BW) ^{0.25} | TRV (mg/kgBW/d) TRV _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t / BW) ^{0.25} | TRV (mg/kgBW/d) TRV _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} ^{0.25} (BW _t / BW) ^{0.25} | TRV (mg/kgBW/d) TRV _t × BW _{conv} |
| | | | | | | | | | |
| INORGANICS | | | | | | | | | |
| Cadmium | Rat | 3.03E-01 | 1.00E+01 | 5.64E-01 | 5.64E-00 | 2.12E-00 | 2.12E+01 | 7.42E-01 | 7.42E-00 |
| Chromium | Rat | 3.50E-01 | 2.74E+04 | 5.84E-01 | 1.60E+04 | 2.20E-00 | 6.02E+04 | 7.69E-01 | 2.11E+04 |
| Lead | Rat | 3.50E-01 | 8.00E+01 | 5.84E-01 | 4.68E+01 | 2.20E-00 | 1.76E+02 | 7.69E-01 | 6.15E+01 |
| Mercury | Mink | 1.00E-00 | 1.01E+01 | 7.60E-01 | 7.67E-00 | 2.86E-00 | 2.89E+01 | 1.00E-00 | 1.01E+01 |
| ORGANICS | | | | | | | | | |
| Volatile Organic Compounds | | | | | | | | | |
| Acetone | Rat | 3.50E-01 | 5.00E-00 | 5.84E-01 | 2.92E-00 | 2.20E-00 | 1.10E+01 | 7.69E-01 | 3.85E-00 |
| Chloroethane | Rat | 3.50E-01 | 1.80E+01 | 5.84E-01 | 1.05E+01 | 2.20E-00 | 3.96E+01 | 7.69E-01 | 1.38E+01 |
| Chloromethane | Rat | 3.50E-01 | 1.80E+01 | 5.84E-01 | 1.05E+01 | 2.20E-00 | 3.96E+01 | 7.69E-01 | 1.38E+01 |
| Methylene chloride | Rat | 3.50E-01 | 5.00E+01 | 5.84E-01 | 2.92E+01 | 2.20E-00 | 1.10E+02 | 7.69E-01 | 3.85E+01 |
| Vinyl chloride | Rat | 3.50E-01 | 1.70E-00 | 5.84E-01 | 9.94E-01 | 2.20E-00 | 3.74E-00 | 7.69E-01 | 1.31E-00 |
| Semivolatile Organic Compounds | | | | | | | | | |
| Benzo(b)fluoranthene | Mouse | 3.00E-02 | 1.33E+02 | 3.16E-01 | 4.21E+01 | 1.19E-00 | 1.58E+02 | 4.16E-01 | 5.54E+01 |
| Pesticides/PCBs | | | | | | | | | |
| 4,4'-DDE | Rat | 3.50E-01 | 4.00E-00 | 5.84E-01 | 2.34E-00 | 2.20E-00 | 8.79E-00 | 7.69E-01 | 3.08E-00 |

BW(kg) Raccoon = 3 per Rod Stafford (OEPA), September 1999.

BW(kg) Short-tailed shrew = 0.015 per Sample, Opreko, and Suter (1996), Table B.1.

BW(kg) Mink = 1 per Sample, Opreko, and Suter (1996), Table B.1.

Table 8-12. Derivation of LOAEL Toxicity Reference Values for Bird Receptors

| Ecological Constituent of Potential Concern | Test Species | Test Species Body Weight BW _t (kg) | TRV _t (mg/kgBW/d) | American Robin | | Green Heron | |
|---|-----------------|---|---------------------------------|--|---|--|---|
| | | | | Body-weight Conversion Factor BW _{conv} (BW _t / BW) ⁰ | TRV (mg/kgBW/d) TRV _t × BW _{conv} | Body-weight Conversion Factor BW _{conv} (BW _t / BW) ⁰ | TRV (mg/kgBW/d) TRV _t × BW _{conv} |
| INORGANICS | | | | | | | |
| Cadmium | Mallard duck | 1.15E-00 | 2.00E+01 | 1.00E-00 | 2.00E+01 | 1.00E-00 | 2.00E+01 |
| Chromium | Black duck | 1.25E-00 | 5.00E-00 | 1.00E-00 | 5.00E-00 | 1.00E-00 | 5.00E-00 |
| Lead | Quail | 1.50E-01 | 1.13E+01 | 1.00E-00 | 1.13E+01 | 1.00E-00 | 1.13E+01 |
| Mercury | Quail | 1.50E-01 | 9.00E-01 | 1.00E-00 | 9.00E-01 | 1.00E-00 | 9.00E-01 |
| ORGANICS | | | | | | | |
| Volatile Organic Compounds | | | | | | | |
| Acetone | None | None | No TRV | None | None | None | None |
| Chloroethane | None | None | No TRV | None | None | None | None |
| Chloromethane | None | None | No TRV | None | None | None | None |
| Methylene chloride | None | None | No TRV | None | None | None | None |
| Vinyl chloride | None | None | No TRV | None | None | None | None |
| Semivolatile Organic Compounds | | | | | | | |
| Benzo(b)fluoranthene | Composite bird | 8.50E-01 | 1.24E+02 | 1.00E-00 | 1.24E+02 | 1.00E-00 | 1.24E+02 |
| Pesticides/PCBs | | | | | | | |
| 4,4'-DDE | Brown pelican | 3.50E-00 | 2.80E-02 | 1.00E-00 | 2.80E-02 | 1.00E-00 | 2.80E-02 |

BW(kg) American robin = 0.077 (Sample, Opresko, and Suter 1996; Table B.1).
BW(kg) Green heron = 0.241 (Birds of North America, No. 129, 1994).

BW(kg) American robin = 0.077 (Sample, Opreko, and Suter 1996; Table B.1).

BW(kg) Green heron = 0.241 (Birds of North America, No. 129, 1994).

10.0. RESULTS OF THE RCRA FACILITY INVESTIGATION AT THE 16 SWMUS RESULTING IN RECOMMENDATION OF FURTHER INVESTIGATION OR A CAP

INDEX OF SITES RECOMMENDED FOR FURTHER INVESTIGATION OR A CAP

The following table provides an index of the sites contained in the 16 SWMUs for which further investigation or a CAP is recommended. Fort Stewart respectfully requests that the Installation's RCRA Subpart B permit be amended to annotate the revised status of these SWMUs, if approved by GEPD.

| Section Number | SWMU Number and Designation | SWMU Name on Hazardous Waste Permit HW-045 |
|----------------|----------------------------------|--|
| 10.1 | 2 | Camp Oliver Landfill |
| 10.2 | 3 | TAC-X Landfill |
| 10.3 | 9 | Inactive EOD Area in Red Cloud Range, Hotel Area |
| 10.4 | 10 | Inactive EOD Area North of Garrison Area |
| 10.5 | 11 | Inactive EOD Area Located Approximately Three Miles Northeast of Garrison Area |
| 10.6 | 12A | Active EOD Containing Open Detonation Unit and Open Burn Unit |
| 10.7 | 18 | Industrial Wastewater Treatment Plant |
| 10.8 | 24B | Old Radiator Shop/Paint Booth |
| 10.9 | 27F (Northwest of Building 1340) | 3d Engineer Brigade Motorpool and Associated Two Oil/Water Separators |
| 10.10 | 27H (Building 1056) | DOL Maintenance Motorpool and Associated Two Oil/Water Separators |
| 10.11 | 27H (Building 1071) | DOL Maintenance Motorpool and Associated Two Oil/Water Separators |
| 10.12 | 27J (Building 10531) | GANG MATES Motorpool and Associated Two Oil/Water Separators |
| 10.13 | 27L | NGTC Block 10200 Wash Rack and Oil/Water Separator |
| 10.14 | 27T | 293 MP Company Wash Rack and Oil/Water Separator |
| 10.15 | 29 | Evans Army Heliport POL Storage Facility |

10.1 SWMU 2: CAMP OLIVER LANDFILL

10.1.1 History and Description of SWMU 2, Camp Oliver Landfill

The Camp Oliver Landfill is located approximately 17 miles northwest of the Fort Stewart garrison area along Fort Stewart Tank Trail 129. It is just north of the bivouac area on a northern side of a small hill and is approximately 2 acres in size. From the 1960s to 1979, the area was used for disposal of refuse from troop training activities and nearby residents via open-pit burning. The landfill was officially closed in 1970; however, the trench method of solid waste disposal was reported to have continued. General refuse from ground maintenance activities and construction debris was placed in the landfill from 1979 to 1984 during the annual 3- to 4-month period of training activities. The landfill is reported to be 15 feet wide by 300 feet long by 5 feet to 6 feet deep (Geraghty and Miller 1992). The waste disposed of at the landfill included garbage and refuse, grass clippings, tree branches, root stumps, and chunks of asphalt and concrete. There was no evidence of disposal of toxic or hazardous wastes in the records searched by Environmental Science and Engineering (1982). There is little obvious surface evidence that a landfill or open dumping area existed. During a site reconnaissance in November 1995, small soil piles, some roofing tin, and wooden construction-type debris were observed. Also, spent small weapons cartridges were observed in the ditch along the site's southwestern and southeastern boundaries. A site reconnaissance in September 1996 indicated no evidence of any landfill operations. Grass, small trees, and bushes now cover the area.

10.1.1.1 1980 Investigation

USACE installed four monitoring wells (CO-M1 through CO-M4) in 1980. Groundwater and surface water samples were collected in 1980 and 1981.

Groundwater. Iron was detected at concentrations that exceeded the drinking water standard of 0.3 mg/L.

Surface Water. Fecal coliform was detected at the two surface water sampling sites. Iron concentrations in the surface water near the landfill were reported as high; however, concentrations near background values for iron were reported a short distance downstream from the landfill.

10.1.1.2 1982 Investigation

Four soil borings (CO-B2 through CO-B5) were installed during a 1982 Environmental Science and Engineering study. Subsurface soil samples were collected for analysis of geotechnical parameters. No samples were submitted for analysis of analytical parameters.

10.1.1.3 1993 Phase I RCRA Facility Investigation

As part of the Phase I RFI conducted in 1993, subsurface soil samples were collected during the drilling of two replacement monitoring wells (CO-M1 and CO-M4) and analyzed for VOCs and total RCRA metals. Subsurface soil samples were collected from 10 feet to 12 feet bgs in CO-M1 and from 6 feet to 8 feet bgs in CO-M4. Groundwater samples were collected from the four site monitoring wells and analyzed for VOCs, total RCRA metals, and pesticides/PCBs. Upstream and downstream surface water samples were collected from Canoochee Creek and analyzed for VOCs, total RCRA metals, and pesticides/PCBs. Analytical results for the Phase I RFI are presented in Table 10.1-1. Groundwater and surface water sampling locations are presented in Figure 10.1-1.

Subsurface Soil

VOCs. Concentrations of VOCs were not reported above the detection limit in the subsurface soil samples.

Pesticides/PCBs. Concentrations of pesticides/PCBs were not reported above the detection limit in the subsurface soil samples.

RCRA Metals. Arsenic, barium, chromium, and lead were detected in subsurface soil; however, none of the metals were detected above the respective reference background criteria.

Groundwater

Barium, chromium, lead, and mercury were detected above their respective reference background criteria in monitoring well CO-M4. Concentrations of chromium and lead also exceeded their respective MCLs. Concentrations of VOCs and pesticides/PCBs were not reported above the detection limit in the groundwater samples.

Surface Water

Concentrations of VOCs were not reported above the detection limit in the surface water samples; however, toluene was detected in the duplicate sample at the upgradient location. The concentration for toluene at that location was below the GEPD guideline of 2,000 µg/L. Barium and lead were detected in the downgradient surface water sample. The lead concentration was above the GEPD guideline value of 1.3 µg/L. Concentrations of pesticides/PCBs were not reported above the detection limit in the groundwater samples.

10.1.2 Summary of Phase II RCRA Facility Investigation Activities

Initial screening consisted of using DPT techniques to collect groundwater samples from three Geoprobe borings for VOC analysis. The results from the Geoprobe screening locations, GP1, GP2, and GP3, were used to determine the characteristics of the leachate at this site. A vertical-profile boring, VP1, was installed in the center of the landfill to determine the vertical extent of groundwater contamination, and groundwater grab samples were collected at 10-foot intervals and analyzed for VOCs.

Originally, five soil borings and one new background monitoring well were installed at the site. Following redevelopment of the existing wells, it was determined that two of the existing wells (MW2 and MW3) were screened below the water table; therefore, two additional wells were installed near these existing wells. Boring logs and monitoring well diagrams are presented in Appendices A and B, respectively. A surface soil sample and a subsurface soil sample were collected from each boring/well. In addition, three surface soil samples were collected from within the boundary of the SWMU. All surface and subsurface soil samples were analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. Phase II RFI sampling locations are presented in Figure 10.1-1.

Monitoring well construction details for the Phase II wells are presented in Table 10.1-2. Geotechnical soil samples were collected from the three monitoring well boreholes, and the results are presented in Table 10.1-3. During site reconnaissance, a well was discovered north of the landfill and was labeled MW8. The origin of this well and the associated construction details are unknown; however, its purpose was for monitoring the LAS located adjacent to SWMU 2. All new and existing monitoring wells were

developed until the turbidity was in the range of less than 10 NTUs to 25 NTUs. Monitoring well development data are presented in Table 10.1-4. Existing well MW1 was dry and was abandoned in accordance with guidance from GEPD, DPW, and USACE. Groundwater samples were collected from seven monitoring wells and analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. Conductivity, temperature, pH, DO, Redox, and turbidity were measured in the field during sampling, and the results are presented in Table 10.1-5.

Two surface water samples and two sediment samples were collected from Canoochee Creek. The upstream location was northwest of the site, while the downstream location was north-northeast of the site.

10.1.3 Physical Characteristics of the Site

10.1.3.1 Topography

There ARE approximately 25 feet of relief across the site. The elevation of the site is approximately 150 feet amsl along the southern boundary and slopes gently to approximately 125 feet amsl along the northern boundary.

10.1.3.2 Surface drainage

Canoochee Creek is located approximately 450 feet north of the northern boundary of the landfill (i.e., MW3 and MW4). Based on the topography, the surface water flow direction is to the north toward Canoochee Creek.

10.1.3.3 Soils

The soils present across the site consist of alternating layers of sand and clayey sands, as indicated in cross sections A-A' and B-B' (Figures 10.1-2 and 10.1-3)

10.1.3.4 Hydrogeology

Groundwater was encountered at approximately 4 feet bgs at MW7 along the northern boundary of the site and at approximately 15 feet bgs along the southern boundary of the site. The shallow groundwater flow direction across the site is to the north toward Canoochee Creek, and the hydraulic gradient is 0.0148 foot/foot (Figure 10.1-4).

10.1.3.5 Ecology

As stated in Section 8.2, the habitats at SWMU 2 are classified as "unmanaged grasslands" and "aquatic habitats." The site includes successional fields of unmanaged grasses, with scattered mature hardwoods and immature pine. Unimproved roads run within 50 feet of the southern and eastern sides of the site's boundaries. A mature pine-oak forest borders the northern and northwestern sides. Surface water runoff flows into Canoochee Creek, which runs within 450 feet of the northern boundary of SWMU 2.

Groundhog holes were abundant throughout the grass-covered area of the site, and nine-banded armadillos were spotted on many occasions throughout the investigation. Evidence of white-tailed deer and coyote (*Canis latrans*) was also apparent.

10.1.4 Nature and Extent of Contamination

10.1.4.1 Surface soil

Eleven surface soil samples were collected from three surface soil locations, five soil boring locations, and three monitoring wells. The surface soil samples were analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. The results of the surface soil analyses are presented in Table 10.1-6 and Figure 10.1-5.

VOCs. 2-Butanone and acetone were detected in the surface soil sample from MW6 at concentrations of 0.0055 mg/kg and 0.174 mg/kg, respectively. No other VOCs were detected in the surface soil samples from the monitoring wells. Acetone was also detected at SB2 and SB5 at concentrations of 0.356 mg/kg and 0.511 mg/kg, respectively.

2-Butanone and acetone are considered to be SRCs in surface soil.

SVOCs. Bis(2-ethylhexyl)phthalate was detected in SB2 at a concentration of 1.1 mg/kg and is considered to be an SRC in surface soil.

Pesticides/PCBs. The site background sample (MW5) contained three pesticides: 4,4'-DDE (0.0011 mg/kg); 4,4'-DDT (0.0024 mg/kg); and methoxychlor (0.0029 mg/kg). Nine other pesticides were detected in only one surface soil sample. 4,4'-DDD and alpha-BHC were detected in sample SB1 at concentrations of 0.0032 mg/kg and 0.00024 mg/kg, respectively. The following pesticides were detected in SS2: aldrin (0.0011 mg/kg), endosulfan II (0.0018 mg/kg), and endrin ketone (0.0026 mg/kg). Pesticides detected in SB2 include: alpha-chlordane (0.00095 mg/kg), delta-BHC (0.0016 mg/kg), and heptachlor (0.001 mg/kg). Dieldrin was the only pesticide detected in sample SS3 at a concentration of 0.003 mg/kg. Endosulfan sulfate was detected in only SB4 at a concentration of 0.0032 mg/kg. Heptachlor epoxide was detected in three samples at concentrations ranging from 0.00018 mg/kg at SB5 to 0.00076 mg/kg at SS2. 4,4'-DDT was detected in four samples at concentrations ranging from 0.0018 mg/kg at SS2 to 0.0042 mg/kg at SB1. Methoxychlor was detected in five samples at concentrations ranging from 0.0029 mg/kg at SB5 to 0.012 mg/kg at SB2. 4,4'-DDE was detected in six samples at concentrations ranging from 0.0012 mg/kg at SB4 to 0.01 mg/kg at SS3.

4,4'-DDD; 4,4'-DDE; 4,4'-DDT; aldrin; alpha-chlordane; alpha-BHC; delta-BHC; dieldrin; endosulfan II; endosulfan sulfate; endrin ketone; heptachlor; heptachlor epoxide; and methoxychlor were detected and are considered to be SRCs in surface soil.

RCRA Metals. Arsenic, barium, chromium, and lead were detected in all 10 of the surface soil samples. Arsenic was detected at concentrations ranging from 0.71 mg/kg at MW7 to 3.4 mg/kg at SS3, barium at concentrations ranging from 13.6 mg/kg at SS2 to 29.5 mg/kg at SS3, chromium at concentrations ranging from 2.3 mg/kg at MW7 to 47.5 mg/kg at SB4, and lead at concentrations ranging from 3.1 mg/kg at MW7 to 19.7 mg/kg at SS3. Cadmium was detected at concentrations of 0.2 mg/kg at MW6 and 0.08 mg/kg at MW7. Mercury was detected at eight of 11 locations (including the site-specific background location, MW5) at concentrations ranging from 0.02 mg/kg at SB2, SB5, and SS1 to 0.04 mg/kg at MW5, MW7, and SB1; selenium was detected at MW6 at a concentration of 0.24 mg/kg.

Because arsenic, barium, cadmium, chromium, lead, and mercury were detected in surface soil samples from one or more of the monitoring wells, soil borings, and surface soil locations at concentrations above the reference background criteria, these metals are considered to be potential SRCs; however, arsenic,

barium, lead, and mercury might not be site-related. Arsenic was detected at less than two times the reference background criterion, including the site-specific background concentration. While barium was elevated above background at most locations, the maximum concentration was less than two times background, and the sampling locations at which the exceedances of reference background occurred were widely distributed, suggesting that barium occurs naturally in surface soils in this area. Cadmium was detected at only one location at a concentration only slightly above the reference background criterion. Lead was found in only one sample, SS3, at a concentration that was only slightly more than two times the reference background criterion. Mercury was detected at a maximum concentration of 0.04 mg/kg compared to the reference background criterion of 0.03 mg/kg. Chromium concentrations for much of the area within the SWMU (at SB4, SS2, SS3) were more than three times reference background.

10.1.4.2 Subsurface soil

Eight subsurface soil samples were collected from five soil borings and three monitoring wells. The subsurface soil samples were analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. The results of the subsurface soil analyses are presented in Table 10.1-7 and Figure 10.1-6.

VOCs. 2-Butanone was the only VOC detected in subsurface soil samples. 2-Butanone was detected in only one of the eight subsurface soil samples (SB3) at a concentration of 0.0076 mg/kg. 2-Butanone is considered to be an SRC in subsurface soil.

SVOCs. Bis(2-ethylhexyl)phthalate was detected in SB5 at a concentration of 0.229 mg/kg and is considered to be an SRC in subsurface soil.

Pesticides/PCBs. Alpha-BHC was detected in subsurface soil samples from MW5 and SB1 at concentrations of 0.00093 mg/kg and 0.00056 mg/kg, respectively. No other pesticides/PCBs were detected in the subsurface soil samples from the monitoring wells. 4,4'-DDE and 4,4'-DDT were also detected in soil boring SB1 at concentrations of 0.0088 mg/kg and 0.0089 mg/kg, respectively. No pesticides/PCBs were detected in the subsurface soil samples from SB2, SB3, SB4, and SB5.

4,4'-DDE; 4,4'-DDT; and alpha-BHC are considered to be SRCs in subsurface soil.

RCRA Metals. Analytical results from subsurface soil samples collected during the Phase I RFI did not indicate concentrations of RCRA metals that exceeded reference background concentrations. However, analytical results from subsurface soil samples collected from SB2, SB3, and SB5 during the subject Phase II RFI indicated concentrations of barium, chromium, and mercury that exceeded reference background concentrations. Barium and chromium were detected at all eight sampling locations. Barium concentrations ranged from 1.3 mg/kg at SB1 to 24.5 mg/kg at SB2; barium was detected in excess of the reference background criterion at only SB2. Chromium concentrations ranged from 3.4 mg/kg at MW7 to 22.5 mg/kg at SB2; chromium was detected in excess of the reference background criterion at SB2 and SB3. Mercury was detected in three of eight subsurface soil sampling locations at concentrations that ranged from 0.03 mg/kg at MW6 and SB2 to 0.23 mg/kg at SB3; mercury was detected in excess of the reference background criterion at only SB2. Arsenic, cadmium, lead, and selenium were also detected at two or more locations, each at concentrations below the reference background criteria. No metal concentrations exceeded the reference background criteria in MW5, MW6, MW7, SB1, and SB5. RCRA metals that exceeded the reference background criteria at this site were primarily detected at locations around its perimeter, with no metals detected at the most central sampling location (MW6).

Barium, chromium, and mercury are considered to be potential SRCs in subsurface soil.

10.1.4.3 Groundwater

Three groundwater screening wells and one vertical-profile boring (VP1) were installed within the boundary of the landfill using DPT techniques and were analyzed for VOCs. The analytical laboratory missed the holding times for VOCs for one of the intervals of the vertical-profile boring installed during the initial sampling endeavor (January 1998). Another vertical-profile boring (VP2) was installed next to the previous location, and groundwater was resampled in May 1998; however, the groundwater was inadvertently analyzed for only benzene, toluene, ethylbenzene, and total xylenes.

In addition, seven groundwater samples were collected from three newly installed monitoring wells and four existing monitoring wells. The groundwater samples from the monitoring wells were analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. The results of the groundwater analyses are presented in Table 10.1-8 and Figure 10.1-7.

VOCs. VOCs were detected in groundwater at relatively low concentrations. Toluene was detected at a concentration of 15.6 µg/L at MW6. Total xylenes were detected in groundwater samples collected in two sampling depths from VP1 at concentrations of 4.3 µg/L (28 feet to 32 feet) and 15.3 µg/L (41 feet to 45 feet). Total xylenes were also detected at a concentration of 4 µg/L from VP2 (40 feet to 45 feet). 4-Methyl-2-pentanone was detected in VP1 (41 feet to 45 feet) at a concentration of 9.9 µg/L. 4-Methyl-2-pentanone, toluene, and total xylenes are considered to be potential SRCs in groundwater.

SVOCs. Bis(2-ethylhexyl)phthalate was detected in MW8 at a concentration of 240 µg/L, which exceeds its MCL. Bis(2-ethylhexyl)phthalate was believed to be the result of field or laboratory contamination; therefore, with the concurrence of GEPD (SAIC 1999a), the groundwater at MW8 was resampled on July 10, 1999. Bis(2-ethylhexyl)phthalate was not detected in MW8 during the resampling. The elevated concentration of bis(2-ethylhexyl)phthalate initially detected was considered to be the result of field or laboratory contamination; therefore, bis(2-ethylhexyl)phthalate is not an SRC at SWMU 2.

Pesticides/PCBs. No pesticides/PCBs were detected in the groundwater samples.

RCRA Metals. Mercury was detected at four of the six groundwater sampling locations that were analyzed for RCRA metals at concentrations that slightly exceeded the reference background criterion and ranged from 0.18 µg/L at MW2 and MW8 to 0.21 µg/L at MW6. Lead was detected at two locations at concentrations that exceeded the reference background criterion: 25.5 µg/L at MW5 and 12.6 µg/L at MW8. Lead exceeded its MCL at MW5, which is a background sampling location. Selenium was detected at only one location, MW3, at a concentration of 2.5 µg/L, which slightly exceeded the reference background criterion. Barium was detected at all monitoring well sampling locations at concentrations that were below the reference background criterion.

Lead, mercury, and selenium are considered to be potential SRCs in groundwater. Lead is not considered to be site-related because lead was detected in an off-site location (MW8), it was not detected in any on-site wells above the reference background criterion, and it was detected at its highest concentration at the upgradient sampling location. Mercury was detected at levels near the detection limit and was detected above the reference background criterion at the upgradient sampling location; therefore, mercury is not considered to be site-related. Selenium was detected in only one well (MW3), which is downgradient, but at a concentration only slightly above the reference background criterion (i.e., 2.5 µg/L versus 1.90 µg/L); therefore, selenium is not considered to be site-related.

10.1.4.4 Surface water

Two surface water samples were collected from Canoochee Creek. The surface water samples were analyzed for VOCs, SVOCs, pesticides/PCBs, and metals. The results of the surface water analyses are presented in Table 10.1-9.

VOCs. No VOCs were detected in the surface water samples.

SVOCs. No SVOCs were detected in the surface water samples.

Pesticides/PCBs. No pesticides/PCBs were detected in the surface water samples.

RCRA Metals. No RCRA metals were detected above the respective reference background criteria in the surface water samples.

10.1.4.5 Sediment

Two sediment samples were collected from Canoochee Creek. The sediment samples were analyzed for VOCs, SVOCs, pesticides/PCBs, and RCRA metals. The results of the sediment analyses are presented in Table 10.1-9 and Figure 10.1-8.

VOCs. Acetone and 2-butanone were detected in the upstream sediment sample (SWS2); however, these compounds were not observed in the downstream sample (SWS1). Therefore, no VOCs are considered to be SRCs in sediment.

SVOCs. Bis(2-ethylhexyl)phthalate was detected in the upstream sediment sample (SWS2); however, this compound was not observed in the downstream sample (SWS1). Therefore, no SVOCs are considered to be SRCs in groundwater.

Pesticides/PCBs. 4,4'-DDE; alpha-chlordane; beta-BHC; delta-BHC; endosulfan sulfate; endrin ketone; and lindane were detected in the upstream sediment sample (SWS2). Only alpha-chlordane, at a concentration of 0.00071 mg/kg (SWS1), was observed in the downstream sample; however, the downstream concentration of alpha-chlordane was less than the upstream concentration. Alpha-chlordane is considered to be a potential SRC in sediment.

RCRA Metals. No RCRA metals were detected above the respective reference background criteria in the sediment samples.

10.1.4.6 Site-related contaminant summary

SRCs by medium and the corresponding maximum concentrations are presented in Table 10.1-10.

10.1.5 Fate and Transport Considerations

This section presents the site-specific components of the CSM developed for SWMU 2 and describes the contaminant release mechanisms through the primary transport medium (groundwater). This section also discusses the fate and transport of contaminants at the site with respect to their leachability and natural attenuation. This section is a site-specific extension for SWMU 2 of Chapter 6.0, which presents a general discussion on the contaminant fate and transport for the 16 SWMUs.

10.1.5.1 Conceptual Site Model

Water Balance Components

The annual average water balance estimates for SWMU 2 indicate an evapotranspiration rate of 65.1 percent (31.27 inches) of total precipitation (48 inches) as compared to 35 percent (16.79 inches) for rainwater available for flow. Of this 35 percent (16.79 inches), groundwater recharge (percolation) accounts for 33.81 percent (16.23 inches) and surface runoff accounts for the remaining 0.97 percent (0.47 inch).

The water balance estimations were based on the HELP model (EPA 1994b) calculations for an uncapped landfill cell using precipitation and temperature data for the years 1974 through 1978 at Savannah, Georgia.

Contaminant Release and Migration Pathways

Past and present contaminant migration pathways are listed below.

- Rainwater percolating through contaminated buried materials/debris and soil below the site leached contaminants and transported them to the water table.
- Buried material degraded and leached contaminants to adjacent soil and groundwater.
- Runoff from exposed material and surface contaminants migrated to surface water in Canoochee Creek.
- Fluctuating groundwater levels contacted contaminated buried material or soil and distributed contaminants in the soil at the water table interface.
- Groundwater flow transported contaminants within the water table aquifer to Canoochee Creek.

Additional current pathways might include the two described below.

- Organic compounds in groundwater and probably in soil are being biologically degraded.
- Organic compounds in soil and probably in groundwater are being volatilized.

The most likely pathways of contaminant migration are (1) groundwater flow to Canoochee Creek, located approximately 450 feet north of the site, and (2) overland flow to Canoochee Creek.

In the saturated zone, the contaminants are carried laterally either in solution or adsorbed to fine particulates (colloids) to the hypothetical receptor locations. Groundwater velocity is a function of hydraulic conductivity, hydraulic gradient, and effective porosity of soil. No slug test was performed at SWMU 2 to determine the horizontal hydraulic conductivity. To estimate horizontal hydraulic conductivity, conductivity measured in the laboratory for the site was used with a horizontal-to-vertical anisotropy ratio of 10. The horizontal hydraulic conductivity was estimated to be 1.40E-05 centimeters per second. The average hydraulic gradient was calculated to be 0.0148 foot/foot. Assuming an effective porosity of 0.16 [based on specific yield of clayey sands (Mills et al. 1985)], the groundwater velocity is calculated to be approximately 1.3 feet/year toward Canoochee Creek. At this velocity, it is expected that

site groundwater will take approximately 345 years to reach Canoochee Creek, which is located approximately 450 feet north of the site.

10.1.5.2 Fate and transport analysis

Results of Soil Leachability Analysis

The site characterization identified inorganic, organic, and pesticide SRCs in surface soil, subsurface soil, sediment, and groundwater. Six inorganics (metals) were identified as SRCs in soil. Based on the leachability analysis, arsenic, chromium, and mercury exceeded their respective GSSLs. Two VOCs and one SVOC were indicated as SRCs in surface and subsurface soils. None of the VOCs or SVOCs exceeded their respective GSSLs. Among the 14 pesticides detected in soil, only alpha-BHC and delta-BHC exceeded their respective GSSLs (Table 10.1-11). Chromium, arsenic, mercury, alpha-BHC, and delta-BHC were identified as CMCOPCs in soil.

Only alpha-chlordane was identified as an SRC in sediment. The maximum concentration did not exceed its GSSL (Table 10.1-11); therefore, there are no CMCOPCs in sediment.

Alpha-BHC, delta-BHC, chromium, and arsenic were not detected in groundwater. Mercury was detected, but did not exceed its MCL in groundwater.

Off-site migration of arsenic, chromium, and mercury is limited because of their high retardation factors. Off-site migration of alpha-BHC and delta-BHC is limited because of their relatively higher retardation factors. Alpha-BHC and delta-BHC can be degraded in the environment through processes including hydrolysis, Redox, photolysis, biodegradation, or volatilization.

Off-site migration of alpha-BHC and delta-BHC will be limited due to retardation and degradation through various processes, as well as slow groundwater movement (approximately 1.3 foot/year). At a flow rate of 1.3 foot/year, site groundwater will take more than 345 years to reach Canoochee Creek. In reality, contaminants will move more slowly than the groundwater due to retardation, and alpha-BHC and delta-BHC will gradually attenuate naturally.

Based on the leachability analysis, none of the constituents from SWMU 2 are expected to be of potential concern at the nearest receptor location (Canoochee Creek).

10.1.6 Human Health Preliminary Risk Evaluation of SWMU 2

SRCs were identified for the following media: surface soil, subsurface soil, sediment, and groundwater. Evaluation of the potential risks resulting from exposure to these constituents and the identification of HHCOPCs are addressed in this section.

10.1.6.1 Exposure evaluation

The exposure evaluation addresses which human receptor populations, both on-site and off-site, might be exposed to the contaminants present at the site. The exposure evaluation also addresses how contaminants might migrate and the potential exposure pathways for the various receptors.

Receptor Assessment

The landfill is currently inactive and is covered by vegetation (grasses and small trees). The landfill is located outside the garrison area near (less than 500 feet away from) an active training area. However, training activities do not take place on the landfill. Given its location, very few individuals are likely to enter the site; however, hunters and juvenile trespassers might cross the area. It is unlikely that any type of military activities would take place at this site; therefore, occupational receptors are not a viable exposure population.

The potential receptors include the following:

- hunters,
- recreational fishermen,
- juvenile trespassers, and
- off-site occupational receptors.

Migration and Exposure Pathway Analysis

Potential migration pathways for surface soils include leaching into groundwater, volatilization, and potential bioaccumulation of contaminants into game species. The site is currently vegetated; therefore, migration of contaminants via fugitive dust is not a viable migration pathway. Volatile organics, which might volatilize into the atmosphere, are present in surface soils. The presence of vegetation on the site would limit transport of contaminants as a result of surface runoff.

The vegetation might serve as a source of food for wildlife. Bioaccumulation of contaminants into plants that are then consumed by wildlife might result in bioaccumulation of contaminants into the tissue of game animals.

Analyses of contaminant migration (see Section 10.1.5.2) indicate that contaminants in soils might leach into groundwater. In addition, contaminants in groundwater might migrate to Canoochee Creek. However, based on the analysis given in Section 10.1.5, none of the constituents from SWMU 2 are expected to be of potential concern as a result of leaching into groundwater.

The migration and exposure pathways are shown in Figure 10.1-9. The on-site resident scenario is not considered to be a viable scenario for this site; however, in accordance with RBCA guidance, it is used to derive screening values. The exposure pathways associated with this scenario are presented to show what pathways would be associated with an on-site resident exposure scenario.

10.1.6.2 Risk evaluation

The human health results of the risk screening are given below.

SRCs for surface soils included two volatile organics, 14 pesticides, a phthalate [bis(2-ethylhexyl)phthalate], and six metals. The maximum concentrations of arsenic and chromium exceeded the screening values for soil ingestion (Table 10.1-12). The maximum concentrations for the remaining chemicals were below their respective screening levels.

The maximum concentration of arsenic was 3.4 mg/kg, which was higher than the screening value of 0.43 mg/kg. The maximum concentration of chromium (47.5 mg/kg) was higher than the screening value of 23 mg/kg. Arsenic and chromium are considered to be HHCOPCs in surface soil.

SRCs for subsurface soils included one volatile organic (2-butanone), three pesticides, a phthalate [bis(2-ethylhexyl)phthalate], and three metals. None of the SRCs exceeded their respective screening values for soil ingestion (Table 10.1-12); therefore, there are no HHCOPCs in subsurface soil.

SRCs for groundwater included three volatile organics and three metals. None of the SRCs in groundwater exceeded their respective screening values (Table 10.1-12); therefore, there are no HHCOPCs in groundwater.

Alpha-chlordane was the only chemical considered to be an SRC for sediment. The maximum concentration of this contaminant was more than two orders of magnitude below its screening value (Table 10.1-12); therefore, there are no HHCOPCs for sediment.

No SRCs were identified for surface water.

10.1.6.3 Uncertainties

Several chemicals were screened using screening values for other similar chemicals. These chemicals included endosulfan sulfate (screening values for endosulfan used), endrin ketone (screening values for endrin used), and alpha-chlordane (values for chlordane used). Small changes in the molecular structure of the molecule can affect the toxicity of the chemical; therefore, based on the screening values of similar chemicals, the chemicals that were eliminated might be HHCOPCs. The screening value for technical BHC was used to evaluate concentrations of delta-BHC. Technical-grade BHC is a mixture of various isomers; therefore, the toxicity of technical BHC might be greater or less than that of delta-BHC. Additional human health uncertainties have been addressed in Section 7.5 of the HHPRE (Chapter 7.0).

10.1.7 Ecological Preliminary Risk Evaluation of SWMU 2

The EPRE was conducted in accordance with GEPD (1996) guidance (see Chapter 8.0). At sites where surface water, sediment, or groundwater was collected, an ESV comparison was conducted. If ECOPCs for aquatic biota were identified in surface water, sediment, or groundwater based on the ESV comparison (Step i), then further evaluation was required for those media. If no ECOPCs were identified based on the Step i screening of those media, then those ECOPCs were not considered further. At sites where surface soil was collected, substances detected in surface soil were evaluated in EPRE Steps ii through v because there are no ESVs for surface soil. The results of the five steps of the EPRE are presented below.

10.1.7.1 Ecological screening value comparison (Step i)

No substances were detected in surface water at SWMU 2 at concentrations exceeding reference background criteria. Surface water was not evaluated further in the EPRE because there are no ECOPCs.

Alpha-chlordane, a pesticide, was the only organic substance detected in sediment at the site. None of the inorganic substances detected in sediment exceeded the background criteria for sediment collected upstream (sample SWS2). The results of the ESV comparison for sediment are presented in Table 10.1-13. Alpha-chlordane is not an ECOPC because its concentration did not exceed the ESV

value for chlordane. Sediment was not evaluated further in the EPRE because it has been eliminated as an ECOPC.

Three RCRA metals—lead, mercury, and selenium—were detected in groundwater at concentrations exceeding the reference background criteria. Three VOCs were also detected in groundwater. The results of the ESV comparison for groundwater are presented in Table 10.1-14. The ECOPCs identified by the ESV comparison for groundwater at the site were total xylenes, lead, and mercury, which were detected at concentrations exceeding the surface water ESV.

Because there are no ESVs for soil, all analytes detected in soil were evaluated further in EPRE Steps ii through v.

10.1.7.2 Preliminary problem formulation (Step ii)

The ecological habitat is described in Section 10.1.3.5. The preliminary assessment endpoints, ecological receptors, and surrogate species representative of those receptors selected for evaluation in the preliminary risk calculation are described in Section 8.2.

10.1.7.3 Preliminary effects (Step iii)

In the EPRE, TRVs were required for shrews and robins ingesting contaminated biota exposed to surface soil at the site, for raccoons ingesting water, and for mink and green herons ingesting aquatic biota. The derivation of TRVs is discussed in Section 8.3. The TRVs derived for shrews, raccoons, and mink are presented in Table 8-5, and TRVs for robins and green herons are presented in Table 8-6.

10.1.7.4 Preliminary exposure (Step iv)

Ecological receptors at the site are probably exposed by ingestion of contaminated soil or of biota exposed to contaminated soil, by ingestion of drinking water, or by ingestion of aquatic biota exposed to surface water. The exposure parameters for the surrogate species—shrews, raccoons, mink, green herons, and robins—are presented in Table 8-7.

10.1.7.5 Preliminary risk calculation (Step v)

The preliminary risk calculation (Step v) uses HQs, the ratios of the measured maximum concentrations and the TRVs, to evaluate the potential for risk. The HQs of ECOPCs with consistent modes of toxicity and effects endpoints are added to calculate an HI. Metals are assumed to have distinct modes of toxicity and effects endpoints; therefore, HIs are calculated for only VOCs and SVOCs when no individual ECOPC has an HQ greater than one and HQs are calculated for more than one chemical. ECOPCs with HQs and HIs less than one indicate little to no likelihood of risk to the ecological receptors. An ERA using site-specific data is indicated for those ECOPCs with calculated HQs or HIs exceeding one (GEPD 1996).

Surface Soil. The preliminary risk calculations for shrews and robins exposed to ECOPCs detected in surface soil are presented in Table 10.1-15. This table shows the maximum detected concentrations, ADDs, TRVs, and HQs for shrews and robins. HQs exceeding one are shown bordered by a double line.

The ECOPCs present in surface soil at concentrations resulting in ADDs exceeding the TRVs for the surrogate species are 4,4'-DDE; cadmium; chromium; and lead. The 4,4'-DDE HQ is 1.8 for robins, and the cadmium HQ is 2.3. The lead HQ for the robin is 10.6. The robin has an HQ of 11.6 for chromium.

Groundwater. The preliminary risk calculations for raccoons, mink, and green herons exposed to ECOPCs detected in groundwater are presented in Table 10.1-16. This table shows the maximum detected concentrations, ADDs, TRVs, and HQs for the receptors. HQs exceeding one are shown bordered by a double line.

The only ECOPC present in groundwater at a concentration resulting in an ADD exceeding the TRV for the surrogate species is mercury. The mink and green heron have mercury HQs of 1.8 and 5.6, respectively. The HI calculated for VOCs does not exceed one.

10.1.7.6 Uncertainties

The risks to ecological receptors from ECOPCs in surface soil and groundwater at SWMU 2 are overestimated by the preliminary risk calculations. Using the maximum detected concentrations in SWMU 2 groundwater, the modeled maximum concentrations of lead, mercury, and total xylenes predicted to occur at a distance of 450 feet from SWMU 2, the estimated distance to the nearest surface water body (Canoochee Creek), are 0.00 µg/L, 0.00396 µg/L, and 0.31 µg/L, respectively. These predicted maximum concentrations do not exceed the EPA Region IV surface water ESVs for lead (1.32 µg/L), mercury (0.012 µg/L), and total xylenes (1.8 µg/L); therefore, the ECOPCs in groundwater at SWMU 2 do not pose a current risk to aquatic biota.

Extrapolating from the leaching modeling results presented in Section 10.1.5, the mean mercury concentration in SWMU 2 subsurface soil (0.0967 mg/kg) would result in a predicted groundwater concentration at Canoochee Creek of approximately 0.168 µg/L, which exceeds the surface water ESV for mercury (0.012 µg/L) by a factor of 14. Thus, future risk to aquatic receptors from mercury leaching from subsurface soil to groundwater potentially discharging to Canoochee Creek is possible if dilution is less than a factor of 14. Groundwater is likely to discharge to Canoochee Creek only during periods of high rainfall, at which time dilution in the creek would probably exceed a factor of 14. Therefore, future risk to aquatic biota from groundwater at SWMU 2 potentially discharging to nearby surface waters is unlikely.

The supplemental risk calculations for robins exposed to 4,4'-DDE; cadmium; chromium; and lead in surface soil are presented in Table 10.1-17. The ADDs calculated using the realistic diet (EPA 1993), the site-specific AUF, and mean soil concentrations of ECOPCs do not exceed LOAEL-based TRVs (HQs less than 1). The supplemental risk evaluations for mink and green herons exposed to mercury in groundwater potentially discharging to surface water are presented in Tables 10.1-18 and 10.1-19, respectively. The mink ADD calculated using realistic diets (EPA 1993), the site-specific AUFs, and mean groundwater concentrations of ECOPCs does not exceed the LOAEL-based TRV (HQ less than 1). The HQ for the green heron is 2, assuming that the site-specific AUF is 1. Not only does an AUF of 1 overestimate exposure, but also the concentration of mercury predicted to occur at a distance of 450 feet would not result in HQs exceeding 1 for the green heron because the downgradient concentration (0.00396 µg/L) is less than the mean concentration (0.12 µg/L) by more than a factor of 3. Likewise, the predicted mean future mercury concentration in groundwater is unlikely to result in HQs exceeding 1 after dilution in Canoochee Creek. Therefore, ECOPCs in soil and groundwater at SWMU 2 do not currently pose a risk to wildlife receptors and will not in the future.

10.1.8 Conclusions and Risk Management and Site Recommendations for SWMU 2

10.1.8.1 Conclusions

Nature and Extent of Contamination

- Groundwater flows north toward Canoochee Creek and has an hydraulic gradient of 0.0148 foot/foot.
- SRCs for surface soils include two volatile organics (2-butanone and acetone), 14 pesticides, a phthalate [bis(2-ethylhexyl)phthalate], and six metals.
- SRCs for subsurface soils include one volatile organic (2-butanone), three pesticides, a phthalate [bis(2-ethylhexyl)phthalate], and three metals.
- SRCs for groundwater include three volatile organics and three metals (lead, mercury, and selenium).
- Bis(2-ethylhexyl)phthalate was also initially detected (at 240 µg/L) in groundwater at MW8, a previously unidentified well that was discovered northwest of and outside the boundaries of SWMU 2. The well is used for monitoring the adjacent LAS, and the construction characteristics of this well are unknown. Bis(2-ethylhexyl)phthalate was not detected at MW8 upon resampling conducted in July 1999, indicating that the initial detection was the result of field or laboratory contamination. Bis(2-ethylhexyl)phthalate is not considered to be an SRC in groundwater for SWMU 2.
- No contaminants were detected in surface water.
- One pesticide, alpha-chlordane, is considered to be an SRC in sediment.

Fate and Transport

- Based on the leachability analysis, arsenic, chromium, mercury, alpha-BHC, and delta-BHC were identified as possible CMCOPCs in soil.

Human Health Preliminary Risk Evaluation

- Arsenic and chromium were identified as possible HHCOPCs for surface soil.
- There are no HHCOPCs in other media at the site.

Ecological Preliminary Risk Evaluation

- 4,4'-DDE; cadmium; chromium; and lead in surface soil at SWMU 2 are ECOPCs because the preliminary HQs exceed 1. Supplemental risk calculations (Table 10.1-17) for 4,4'-DDE; cadmium; chromium; and lead result in HQs less than 1. Therefore, 4,4'-DDE; cadmium; chromium; and lead in surface soil are unlikely to pose a risk to robins, and further investigation and/or evaluation of these constituents in surface soil is not warranted.
- Total xylenes, lead, and mercury were identified as ECOPCs because they occur at levels that exceed the EPA Region IV surface water ESVs. These ECOPCs are unlikely to be potential hazards

to aquatic biota living in downgradient surface water bodies because the predicted maximum concentrations by modeling at the point of discharge to the nearest surface water body do not exceed the EPA Region IV surface water ESVs under current conditions and are unlikely to exceed ESVs under future conditions.

- Mercury in groundwater at SWMU 2 is an ECOPC for wildlife receptors because the preliminary HQs exceed 1. Mercury is unlikely to be a potential hazard to wildlife receptors ingesting aquatic biota living in downgradient surface water bodies now or in the future because the predicted maximum discharge concentration of mercury after dilution is unlikely to result in HQs exceeding 1.

10.1.8.2 Risk management and site recommendations

- Arsenic, chromium, mercury, alpha-BHC, and delta-BHC were identified as possible CMCOPCs based on their potential to leach to groundwater. The maximum concentration (0.00056 mg/kg) of alpha-BHC was collected below the water table and would not be a source of alpha-BHC leaching into the groundwater. The soil samples collected above the water table did not have concentrations of alpha-BHC that exceeded the GSSL. In addition, alpha-BHC was not detected in groundwater at this site; therefore, alpha-BHC is not considered to be a CMCOPC. The maximum concentration of arsenic in surface soil (3.4 mg/kg) does not exceed the reference subsurface background criterion (8.04 mg/kg). Given the relative thickness of the subsurface soil and the higher concentrations of arsenic present in this soil stratum, the contribution of arsenic from surface soil is not likely to be significant. In addition, arsenic was not identified as a COPC in groundwater, indicating that arsenic is not leaching to groundwater in significant concentrations; therefore, arsenic is not considered to be a CMCOPC in soil. Leachate modeling (see Attachment 10.1A) for delta-BHC indicated that delta-BHC naturally attenuates before reaching the water table. Leachate modeling (see Attachment 10.1A) for mercury and chromium estimated that the maximum groundwater concentrations of mercury and chromium (3.6 µg/L and 1,075 µg/L, respectively) will exceed their respective groundwater screening values (1.0 µg/L and 11.0 µg/L, respectively). Therefore, the CMCOPCs, mercury and chromium, were assessed in the human health risk characterization (see bullet 2), whereas arsenic, alpha-BHC, and delta-BHC were eliminated as CMCOPCs.
- As indicated in the previous bullet, chromium was determined to be a CMCOPC based on its potential to leach to groundwater. The potential risks associated with exposure to CMCOPCs in groundwater based on the estimated maximum groundwater concentrations derived from the leachate modeling were evaluated in a baseline risk assessment (see Attachment 10.1A). In addition, groundwater migration modeling was used to determine if chromium could migrate to Canoochee Creek. The results of the groundwater migration modeling indicated that chromium would not migrate to Canoochee Creek (see Attachment 10.1A); therefore, the baseline risk assessment addressed only the potential risk from exposure to groundwater. Chromium is considered to be an HHCOC based on the results of the risk characterization. The risk values for chromium in groundwater exceeded the target systemic target risk value of 0.1 for all of the potentially exposed receptors; therefore, risk-based remedial levels were developed. The recommended risk-based soil remedial level (4.6 mg/kg) for chromium based on leaching to groundwater was less than the reference subsurface soil background concentration (11.60 mg/kg). Given the comparative thickness of subsurface soil and its proximity to groundwater relative to surface soils, the amount of chromium (CMCOC) potentially leaching to groundwater from the subsurface soil is likely to be much greater than the contribution from the surface soil; therefore, the reference subsurface soil background criterion (11.60 mg/kg) is recommended as the remedial level (based on leaching) for chromium in soil.

- Mercury was determined to be a possible CMCOPC based on its potential to leach to groundwater. The potential risks associated with exposure to CMCOPCs in groundwater based on the estimated groundwater concentrations derived from the leachate modeling were evaluated in a baseline risk assessment. In addition, groundwater migration modeling was used to estimate potential surface water concentrations of mercury in Canoochee Creek (see Attachment 10.1A). The baseline risk assessment evaluated the potential risk to off-site receptors exposed to mercury in Canoochee Creek. Mercury is considered to be an HHCOC based on the results of the risk characterization for both on-site receptors (groundwater exposure) and off-site receptors (surface water exposure). The risk values for mercury in groundwater and surface water exceeded the systemic target risk value of 0.1 for all of the potentially exposed receptors; therefore, risk-based remedial levels were developed. Risk-based remedial levels were developed for mercury based on receptors exposed to groundwater and receptors exposed to surface water. In addition, a soil remedial level was developed based on the MCL for mercury. The risk-based remedial level for protection of surface water (0.003 mg/kg) was the lowest remedial level. However, this value is below the subsurface soil reference background concentration (0.05 mg/kg). Given the comparative thickness of subsurface soil and its proximity to groundwater relative to surface soils, the amount of mercury (CMCOC) potentially leaching to groundwater from the subsurface soil is likely to be much greater than the contribution from the surface soil; therefore, the reference subsurface soil background criterion (0.05 mg/kg) is recommended as the remedial level (based on leaching) for mercury in soil.
- Arsenic and chromium are potential HHCOPCs for surface soils at SWMU 2 based on direct exposure to soils. A baseline risk assessment addressing the risk associated with direct exposure to arsenic and chromium in surface soil was performed (see Attachment 10.1A). The risk values for arsenic in soil exceeded the target risk values for the current and future on-site Installation worker, the on-site resident child, and the on-site resident adult. The risk values for chromium in surface soil exceeded the target risk values for the on-site resident child. Remedial levels were derived for arsenic and mercury in surface soil based on direct exposure to soil. Risk-based remedial levels for arsenic were calculated for both carcinogenic and noncarcinogenic risk because it was the only COC that contributed significantly to the carcinogenic risks at this site. A risk-based remedial level for arsenic based on an HI of 0.5 (11.68 mg/kg) is recommended because it takes into account potential contributions of other COCs to noncarcinogenic risks and because it is well below the target ILCR of 1×10^{-4} . Because the recommended risk-based remedial level for arsenic (11.68 mg/kg) exceeds the maximum detected value of 3.4 mg/kg, no further investigation is recommended to address arsenic. All of the risk-based remedial levels for direct exposure to chromium in surface soils are less than the reference background concentration; therefore, the surface soil background criterion (6.21 mg/kg) was selected as the recommended remedial level for chromium in surface soil.
- The chromium concentrations detected above their respective remedial levels in surface and subsurface soil are presented in Figure 10.1-10. Chromium was also detected in surface soil at the background location (MW5) at a concentration above the remedial level (surface soil background reference criterion), indicating that chromium may be naturally elevated in this area. The observed chromium concentrations in surface and subsurface soil were on the low end of the concentration range (1 mg/kg to 1,000 mg/kg) established by the USGS for the eastern United States (USGS 1984) and may represent natural variability in the soil. Chromium was not detected in groundwater. Therefore, chromium in surface and subsurface soil does not require additional investigation and/or evaluation.
- Mercury was elevated in only one out of seven subsurface soil samples (Figure 10.1-10). The mercury concentrations in the remaining subsurface soil locations were either nondetect (four

locations) or below the subsurface reference background criterion (two locations). The observed mercury concentration in soil was on the low end of the concentration range (0.01 mg/kg to 3.4 mg/kg) established by the USGS for the eastern United States (USGS 1984). The only elevated mercury detection could be attributable to natural variability or an anomaly in the soil rather than representative of widespread contamination. Mercury was not detected above its MCL in groundwater, indicating that its movement/migration is highly retarded by its physiochemical properties and by site conditions. Therefore, additional investigation and/or evaluation of mercury in subsurface soil is not required.

- Total xylenes, lead, and mercury in surficial groundwater are potential ECOPCs for this site. These ECOPCs are a potential hazard to aquatic biota if groundwater discharges to nearby surface water bodies. However, as stated in Section 10.1.5.1, groundwater at this site will take approximately 345 years to reach the nearest downgradient surface water body, Canoochee Creek. ECOPCs detected in groundwater at the site are unlikely to be potential hazards to receptors because of the slow movement of groundwater relative to natural attenuation processes. Total xylenes (organic) are probably being reduced by the natural attenuation processes of hydrolysis, Redox, biodegradation, and volatilization. Movement/migration of inorganic constituents is highly retarded by their physiochemical properties. Therefore, total xylenes, lead, and mercury in groundwater are not considered to be ECOPCs for this site and do not require further investigation and/or evaluation.
- Mercury in surficial groundwater was identified as a potential ECOPC for terrestrial receptors at this site during the preliminary risk evaluation. Supplemental risk calculations resulted in an HQ of less than 1 (Table 10.1-18), and modeling (see Appendix K) indicated that maximum concentrations at the point of discharge to the nearest surface water body do not exceed surface water ESVs. Therefore, mercury is not considered to be an ECOPC in groundwater for terrestrial receptors, and further investigation and/or evaluation is not required.
- Based on the information presented in this section, an NFA status is recommended for SWMU 2 regarding further investigation of the site. Fort Stewart recommends that a CAP proposing and evaluating institutional controls (e.g., deed restrictions, land-use restrictions) be prepared for SWMU 2. Institutional controls would be protective of human health by the establishment of land-use controls for present and future land use. It is anticipated that the CAP will be submitted to GEPD in the first fiscal quarter (October through December) of 2001. The potential abandonment or use of the existing monitoring wells will be evaluated in the CAP.

Table 10.1-1. Summary of Phase I RFI Results, SWMU 2

| SUBSURFACE SOIL | | | |
|-----------------------|-------------------------------|---------------------|--------|
| Analyte | Reference Background Criteria | Sample Location | |
| | | SL1-12 ^a | SL4-8 |
| Depth (feet) | | 10 to 12 | 6 to 8 |
| <i>Metals (mg/kg)</i> | | | |
| Arsenic | 8.04 | 2 | 1.8 |
| Barium | 17.00 | 1.2 | 4.2 |
| Chromium | 11.60 | 9.5 | 7.9 |
| Lead | 11.10 | 1.4 | 3.1 |

| GROUNDWATER | | | | | | |
|---------------|-------------------------------|-------|--------------------|-------|-------|-------|
| Analyte | Reference Background Criteria | MCL | Sample Location | | | |
| | | | CO-M1 ^a | CO-M2 | CO-M3 | CO-M4 |
| Metals (µg/L) | | | | | | |
| Barium | 71.72 | 2,000 | <50 | <50 | <50 | 110 |
| Chromium | 3.56 | 100 | <50 | <50 | <50 | 110 |
| Lead | 4.69 | 15 | 8 | <5 | <5 | 136 |
| Mercury | 0.14 | 2 | <0.2 | <0.2 | <0.2 | 0.2 |

| SURFACE WATER | | | |
|--|-----------------|--------|----|
| Analyte | Sample Location | | |
| | S1 ^a | S1 Dup | S2 |
| <i>Volatile Organic Compounds (µg/L)</i> | | | |
| Toluene | <5 | 6 | <5 |
| <i>Metals (µg/L)</i> | | | |
| Barium | <50 | <50 | 60 |
| Lead | <5 | 6 | 13 |

^aSite-specific background location.

Bold indicates concentrations above reference background criteria.

Boxed *italic* indicates concentrations above MCLs.

Table 10.1-2. Monitoring Well Construction Summary, SWMU 2

| Well No. | Date Installed | Size/Type | Coordinates | Total Depth (feet) | Screen Interval (feet bgs) | Top of Filter Pack Elevation (feet bgs) | Top of Casing Elevation (feet) |
|----------|----------------|------------|----------------------------|--------------------|----------------------------|---|--------------------------------|
| 02-MW5 | 1/14/98 | 2-inch PVC | N 754656.90 E 765554.21 | 23.0 | 12.7 to 22.7 | 10.5 | 160.14 |
| 02-MW6 | 1/27/98 | 2-inch PVC | N 755946.61 E 765736.69 | 15.0 | 3.0 to 12.98 | 1.0 | 134.98 |
| 02-MW7 | 1/27/98 | 2-inch PVC | N 756365.34 E 765743.68 | 15.0 | 4.0 to 14.04 | 3.0 | 127.80 |

Note: All elevations are NGVD 1929.

Table 10.1-3. Summary of Geotechnical Analyses, SWMU 2

| Station | 02-MW5 | 02-MW6 | 02-MW7 |
|------------------------------|------------------|-----------|----------|
| Sample ID | 021513 | 021613 | 021713 |
| Depth (feet) | 10 to 12.3 | 7.5 to 10 | 11 to 12 |
| Moisture content (%) | 17 | 24.6 | 14.3 |
| Liquid limit | 69 | 58 | NP |
| Plastic limit | 21 | 20.8 | NP |
| Plasticity index | 48 | 37.2 | NP |
| Gravel (%) | 0 | 0 | 0 |
| Sand (%) | 96.3 | 88.2 | 94.7 |
| Fines (%) | 3.7 | 11.8 | 5.3 |
| Specific gravity | 2.61 | NA | NA |
| Porosity | 0.3 | NA | NA |
| Permeability (cm/sec) | 1.40E-06 | NA | NA |
| Total organic carbon (mg/kg) | 239 ^a | NA | NA |

^aSample ID is 021512, collected at a depth of 13 feet to 15 feet bgs.

NA = Not analyzed.

NP = Non-plastic.

Table 10.1-4. Well Development Summary, SWMU 2

| Well No. | Date | Total Development Time (hours) | Total Volume Removed (gallons) | Final Turbidity Reading (NTUs) | Total Well Depth (feet) |
|----------|---------|--------------------------------|--------------------------------|--------------------------------|-----------------------------|
| 02-MW1 | 1/15/98 | 1 hour, 46 minutes | 10 | 2.1 | Well abandoned ^a |
| 02-MW2 | 1/15/98 | 2 hours, 34 minutes | 260 | 3.3 | 47.70 |
| 02-MW3 | 2/3/98 | 1 hour, 2 minutes | 125 | 0.5 | 28.00 |
| 02-MW4 | 1/14/98 | 58 minutes | 67 | 5.0 | 17.26 |
| 02-MW5 | 1/19/98 | 3 hours, 20 minutes | 70 | 0.6 | 26.30 |
| 02-MW6 | 1/31/98 | 55 minutes | 80 | 4.8 | 14.00 |
| 02-MW7 | 1/31/98 | 55 minutes | 55 | 1.1 | 16.37 |
| 02-MW8 | 1/15/98 | 40 minutes | 70 | 5.5 | 16.42 |

^aWell was abandoned due to insufficient water in well.

**Table 10.1-5. Field Parameter Measurements during Groundwater and
Surface Water Sampling, SWMU 2**

| Parameter | Date | pH (su) | Conductivity (mS/cm) | Temperature (°C) | Turbidity (NTUs) | DO (mg/L) | Redox (mV) |
|----------------------|---------|------------|-------------------------|---------------------|---------------------|--------------|---------------|
| <i>Groundwater</i> | | | | | | | |
| 02-MW2 | 2/15/98 | 4.62 | 23 | 19.55 | 2.7 | 1.14 | 413 |
| 02-MW3 | 2/24/98 | 4.36 | 50 | 18.46 | 1.8 | 3.40 | -18.9 |
| 02-MW4 | 2/15/98 | 4.33 | 30.0 | 16.66 | 25.0 | N/A | 362 |
| 02-MW5 ^a | 2/15/98 | 4.67 | 51 | 18.21 | 2.4 | 1.27 | 404.1 |
| 02-MW6 | 2/15/98 | 6.15 | 434 | 16.87 | 8.9 | 4.74 | -26 |
| 02-MW7 | 2/15/98 | 4.72 | 19.00 | 13.45 | 4.2 | N/A | 306.0 |
| 02-MW8 | 2/15/98 | 4.69 | 28 | 14.84 | 9.8 | 4.15 | 400 |
| 02-MW8 | 7/10/99 | 4.70 | 23.0 | 23.74 | 9.8 | 6.4 | 307.7 |
| Average ^b | | 4.80 | | | | | |
| <i>Surface Water</i> | | | | | | | |
| 02-SWS1 | 1/14/98 | 6.84 | 60 | 13.27 | NA | 7.88 | 11.4 |
| 02-SWS2 ^a | 1/14/98 | 4.76 | 59 | 12.05 | NA | 10.01 | 200.4 |

^aSite-specific background location.

^bSite-specific background location not included in average.

NA = Not analyzed.

Table 10.1-6. Summary of Analytes Detected in Surface Soil, SWMU 2

| Station | Reference Background Criteria | 02-MW5 ^a | 02-MW6 | 02-MW7 | 02-SB1 | 02-SB2 | 02-SB3 |
|--|-------------------------------------|---------------------|----------|----------|----------|----------|----------|
| Sample ID | | 021511 | 021611 | 021711 | 021A11 | 021B11 | 021C11 |
| Date | | 01/14/98 | 01/27/98 | 01/29/98 | 01/14/98 | 01/14/98 | 01/15/98 |
| Depth (feet) | | 0 to 2 | 0 to 3 | 0 to 1 | 0 to 2 | 0 to 2 | 0 to 2 |
| Sample Type | | Grab | Grab | Grab | Grab | Grab | Grab |
| Volatile Organic Compounds (mg/kg) | | | | | | | |
| 2-Butanone | 0.00 | | 0.0055 | | | | |
| Acetone | 0.00 | | 0.174 | | | 0.356 | |
| Semivolatile Organic Compounds (mg/kg) | | | | | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | | | | | 1.1 | |
| Pesticides/PCBs (mg/kg) | | | | | | | |
| 4,4'-DDD | 0.00 | | | | 0.0032 | | |
| 4,4'-DDE | 0.00 | 0.0011 | | | 0.0096 | 0.0014 | |
| 4,4'-DDT | 0.00 | 0.0024 | | | 0.0042 | | |
| Aldrin | 0.00 | | | | | | |
| alpha-BHC | 0.00 | | | | 0.00024 | | |
| alpha-Chlordane | 0.00 | | | | | 0.00095 | |
| delta-BHC | 0.00 | | | | | 0.0016 | |
| Dieldrin | 0.00 | | | | | | |
| Endosulfan II | 0.00 | | | | | | |
| Endosulfan sulfate | 0.00 | | | | | | |
| Endrin ketone | 0.00 | | | | | | |
| Heptachlor | 0.00 | | | | | 0.001 | |
| Heptachlor epoxide | 0.00 | | | | | | |
| Methoxychlor | 0.00 | 0.0029 | | | | 0.012 | |
| Metals (mg/kg) | | | | | | | |
| Arsenic | 2.10 | 2.2 | 2.4 | 0.71 | 1.9 | 0.82 | 1.8 |
| Barium | 14.70 | 21.9 | 25.4 | 22.8 | 23.3 | 28.2 | 22 |
| Cadmium | 0.18 | | 0.2 | 0.08 | | | |
| Chromium | 6.21 | 12.1 | 13.9 | 2.3 | 10 | 4.4 | 10.8 |
| Lead | 8.81 | 6.8 | 5.9 | 3.1 | 7.6 | 4 | 5.1 |
| Mercury | 0.03 | 0.04 | 0.03 | 0.04 | 0.04 | 0.02 | |
| Selenium | 0.41 | | 0.24 | | | | |

Note: Footnotes appear on page 10.1-22.

Table 10.1-6. Summary of Analytes Detected in Surface Soil, SWMU 2 (continued)

| Station | Reference Background Criteria | 02-SB4 | 02-SB5 | 02-SS1 | 02-SS2 | 02-SS3 |
|--|-------------------------------------|----------|----------|----------|----------|----------|
| Sample ID | | 021D11 | 021E11 | 027111 | 027211 | 027311 |
| Date | | 01/15/98 | 01/15/98 | 01/16/98 | 01/16/98 | 01/16/98 |
| Depth (feet) | | 0 to 2 | 0 to 2 | 0 to 1 | 0 to 1 | 0 to 1 |
| Sample Type | | Grab | Grab | Grab | Grab | Grab |
| Volatile Organic Compounds (mg/kg) | | | | | | |
| 2-Butanone | 0.00 | | | | | |
| Acetone | 0.00 | | 0.511 | | | |
| Semivolatile Organic Compounds (mg/kg) | | | | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | | | | | |
| Pesticides/PCBs (mg/kg) | | | | | | |
| 4,4'-DDD | 0.00 | | | | | |
| 4,4'-DDE | 0.00 | 0.0012 | 0.0091 | 0.0029 | | 0.01 |
| 4,4'-DDT | 0.00 | | 0.0034 | | 0.0018 | 0.0023 |
| Aldrin | 0.00 | | | | 0.0011 | |
| alpha-BHC | 0.00 | | | | | |
| alpha-Chlordane | 0.00 | | | | | |
| delta-BHC | 0.00 | | | | | |
| Dieldrin | 0.00 | | | | | 0.003 |
| Endosulfan II | 0.00 | | | | 0.0018 | |
| Endosulfan sulfate | 0.00 | 0.0032 | | | | |
| Endrin ketone | 0.00 | | | | 0.0026 | |
| Heptachlor | 0.00 | | | | | |
| Heptachlor epoxide | 0.00 | 0.00032 | 0.00018 | | 0.00076 | |
| Methoxychlor | 0.00 | 0.0051 | 0.0029 | 0.0072 | | 0.0038 |
| Metals (mg/kg) | | | | | | |
| Arsenic | 2.10 | 1.3 | 1.6 | 1.4 | 2.2 | 3.4 |
| Barium | 14.70 | 24.5 | 28.8 | 21.7 | 13.6 | 29.5 |
| Cadmium | 0.18 | | | | | |
| Chromium | 6.21 | 47.5 | 9.8 | 5.8 | 19.6 | 21 |
| Lead | 8.81 | 5.4 | 5.4 | 4.8 | 5.1 | 19.7 |
| Mercury | 0.03 | 0.03 | 0.02 | 0.02 | | |
| Selenium | 0.41 | | | | | |

^aSite-specific background location.

Bold indicates concentrations above reference background criteria.

Table 10.1-7. Summary of Analytes Detected in Subsurface Soil, SWMU 2

| Station | Reference Background Criteria | 02-MW5 ^a | 02-MW6 | 02-MW7 | 02-SB1 | 02-SB2 | 02-SB3 | 02-SB4 | 02-SB5 |
|--|-------------------------------------|---------------------|----------|----------|----------|----------|----------|----------|----------|
| Sample ID | | 021512 | 021612 | 021712 | 021A12 | 021B12 | 021C12 | 021D12 | 021E12 |
| Date | | 01/14/98 | 01/27/98 | 01/29/98 | 01/14/98 | 01/14/98 | 01/15/98 | 01/15/98 | 01/15/98 |
| Depth (feet) | | 13 to 15 | 8 to 10 | 8 to 10 | 13 to 15 | 3 to 5 | 5 to 8 | 5 to 8 | 8 to 10 |
| Sample Type | | Grab | Grab | Grab | Grab | Grab | Grab | Grab | Grab |
| Volatile Organic Compounds (mg/kg) | | | | | | | | | |
| 2-Butanone | 0.00 | | | | | | 0.0076 | | |
| Semivolatile Organic Compounds (mg/kg) | | | | | | | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | | | | | | | | 0.229 |
| Pesticides/PCBs (mg/kg) | | | | | | | | | |
| 4,4'-DDE | 0.00 | | | | 0.0088 | | | | |
| 4,4'-DDT | 0.00 | | | | 0.0089 | | | | |
| alpha-BHC | 0.00 | 0.00093 | | | 0.00056 | | | | |
| Metals (mg/kg) | | | | | | | | | |
| Arsenic | 8.04 | 1.3 | 0.77 | | 1.1 | 4.3 | 1.5 | 0.85 | 1.5 |
| Barium | 17.00 | 1.8 | 3.8 | 2.7 | 1.3 | 24.5 | 8.5 | 13.7 | 8.7 |
| Cadmium | 0.24 | | 0.09 | 0.09 | | | | | |
| Chromium | 11.60 | 6.4 | 7.1 | 3.4 | 5.5 | 22.5 | 13.3 | 8.5 | 15 |
| Lead | 11.10 | 2.2 | 4 | 1.4 | 1.5 | 5.6 | 4.2 | 3.6 | 4 |
| Mercury | 0.05 | | 0.03 | | | 0.03 | 0.23 | | |
| Selenium | 1.12 | | 0.34 | | 0.22 | | 0.26 | | |

^aSite-specific background location.

Bold indicates concentrations above reference background criteria.

Table 10.1-8. Summary of Analytes Detected in Groundwater, SWMU 2

| Station | Reference Background Criteria | MCL | 02-GP1 | 02-GP2 | 02-GP3 | 02-MW2 | 02-MW3 | 02-MW4 | 02-MWS ^a |
|---------------------------------------|-------------------------------------|--------|----------|----------|----------|----------|----------|----------|---------------------|
| Sample ID | | | 024151 | 024251 | 024351 | 024211 | 024311 | 024411 | 024511 |
| Date | | | 01/16/98 | 01/16/98 | 01/16/98 | 02/15/98 | 02/24/98 | 02/15/98 | 02/15/98 |
| Depth (feet) | | | | | | | | | |
| Sample Type | | | Grab | Grab | Grab | Grab | Grab | Grab | Grab |
| Volatile Organic Compounds (µg/L) | | | | | | | | | |
| 4-Methyl-2-pentanone | 0.00 | | | | | | | | |
| Toluene | 0.00 | 1,000 | | | | | | | |
| Xylenes, total | 0.00 | 10,000 | | | | | | | |
| Semivolatile Organic Compounds (µg/L) | | | | | | | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | 6 | NA | NA | NA | | | | |
| Metals (µg/L) | | | | | | | | | |
| Barium | 71.72 | 2,000 | NA | NA | NA | 7.7 | 9.7 | 18.7 | 3.8 |
| Lead | 4.69 | 15 | NA | NA | NA | | | 2.1 | 25.5 |
| Mercury | 0.14 | 2 | NA | NA | NA | 0.18 | | | 0.2 |
| Selenium | 1.90 | 50 | NA | NA | NA | | 2.5 | | |
| Other Analytes (µg/L) | | | | | | | | | |
| Sulfate | 26,717.50 | | NA | NA | NA | NA | NA | 185 | NA |

Note: Footnotes appear on page 10.1-26.

Table 10.1-8. Summary of Analytes Detected in Groundwater, SWMU 2 (continued)

| Station | Reference Background Criteria | MCL | | | |
|--|-------------------------------------|----------|----------|----------|----------|
| Sample ID | | 02-VP2 | 02-VP2 | 02-VP2 | 02-VP2 |
| Date | | 024552 | 024553 | 024554 | 024555 |
| Depth (feet) | | 05/6/98 | 05/6/98 | 05/6/98 | 05/6/98 |
| Sample Type | | 10 to 15 | 20 to 25 | 30 to 35 | 40 to 45 |
| | | Grab | Grab | Grab | Grab |
| <i>Volatile Organic Compounds (µg/L)</i> | | | | | |
| 4-Methyl-2-pentanone | 0.00 | NA | NA | NA | NA |
| Toluene | 0.00 | | | | |
| Xylenes, total | 0.00 | 10,000 | | | |
| | | | | | 4 |
| <i>Semivolatile Organic Compounds (µg/L)</i> | | | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | 6 | NA | NA | NA |
| <i>Metals (µg/L)</i> | | | | | |
| Barium | 71.72 | 2,000 | NA | NA | NA |
| Lead | 4.69 | 15 | NA | NA | NA |
| Mercury | 0.14 | 2 | NA | NA | NA |
| Selenium | 1.90 | 50 | NA | NA | NA |
| <i>Other Analytes (µg/L)</i> | | | | | |
| Sulfate | 26,717.50 | NA | NA | NA | NA |

^aSite-specific background location.

^bBis(2-ethylhexyl)phthalate was believed to be the result of field or laboratory contamination. MW8 was resampled in July 1999. Bis(2-ethylhexyl)phthalate was not detected in the resampled groundwater; therefore, the initial bis(2-ethylhexyl)phthalate was considered to be due to field or laboratory contamination. Bis(2-ethylhexyl)phthalate is not considered to be an SRC in groundwater.

NA = Not analyzed.

Bold indicates concentrations above reference background criteria.
Boxed *italic* indicates concentrations above MCLs.

Table 10.1-9. Summary of Analytes Detected in
Surface Water and Sediment, SWMU 2

| SURFACE WATER | | | |
|----------------|-------------------------------------|----------|----------------------|
| Station | Reference Background Criteria | 02-SWS1 | 02-SWS2 ^a |
| Sample ID | | 023111 | 023211 |
| Date | | 01/14/98 | 01/14/98 |
| Sample Type | | Grab | Background |
| Metals (mg/kg) | | | |
| Barium | 46.8 | 21.4 | 23.4 |
| Chromium | 1.86 | 1.3 | 0.93 |

| SEDIMENT | | | |
|--|-------------------------------------|----------|----------------------|
| Station | Reference Background Criteria | 02-SWS1 | 02-SWS2 ^a |
| Sample ID | | 022111 | 022211 |
| Date | | 01/14/98 | 01/14/98 |
| Sample Type | | Grab | Grab |
| Volatile Organic Compounds (mg/kg) | | | |
| 2-Butanone | 0.00 | | 0.0272 |
| Acetone | 0.00 | | 0.149 |
| Semivolatile Organic Compounds (mg/kg) | | | |
| Bis(2-ethylhexyl)phthalate | 0.00 | | 0.388 |
| Pesticides/PCBs (mg/kg) | | | |
| 4,4'-DDE | 0.00 | | 0.0021 |
| alpha-Chlordane | 0.00 | 0.00071 | 0.0013 |
| beta-BHC | 0.00 | | 0.0022 |
| delta-BHC | 0.00 | | 0.0034 |
| Endosulfan sulfate | 0.00 | | 0.0115 |
| Endrin ketone | 0.00 | | 0.0027 |
| gamma-BHC (Lindane) | 0.00 | | 0.001 |
| Metals (mg/kg) | | | |
| Arsenic | 2.4 | 1.4 | 1.2 |
| Barium | 22.4 | 3 | 11.2 |
| Chromium | 16.2 | 7.2 | 8.1 |
| Lead | 8.6 | 1.2 | 4.3 |

^aSite-specific background sample location.

Bold indicates concentrations above reference background criteria.

Table 10.1-10. Summary of Site-related Contaminants, SWMU 2

| Analyte | Maximum Concentration (mg/kg) | | | Maximum Concentration (µg/L) | |
|---------------------------------------|-------------------------------|----------------------|----------|------------------------------|---------------|
| | Surface Soil | Subsurface Soil | Sediment | Groundwater | Surface Water |
| <i>Volatile Organic Compounds</i> | | | | | |
| 2-Butanone | 0.0055 | 0.0076 | ND | ND | ND |
| 4-Methyl-2-pentanone | ND | ND | ND | 9.9 | ND |
| Acetone | 0.511 | ND | ND | ND | ND |
| Toluene | ND | ND | ND | 15.6 | ND |
| Xylenes, total | ND | ND | ND | 15.3 | ND |
| <i>Semivolatile Organic Compounds</i> | | | | | |
| Bis(2-ethylhexyl)phthalate | 1.1 | 0.229 | ND | ND | ND |
| <i>Pesticides/PCBs</i> | | | | | |
| 4,4'-DDD | 0.0032 | ND | ND | ND | ND |
| 4,4'-DDE | 0.01 | 0.0088 | ND | ND | ND |
| 4,4'-DDT | 0.0042 | 0.0089 | ND | ND | ND |
| Aldrin | 0.0011 | ND | ND | ND | ND |
| alpha-BHC | 0.00024 | 0.00056 ^a | ND | ND | ND |
| alpha-Chlordane | 0.00095 | ND | 0.00071 | ND | ND |
| beta-BHC | ND | ND | ND | ND | ND |
| delta-BHC | 0.0016 | ND | ND | ND | ND |
| Dieldrin | 0.003 | ND | ND | ND | ND |
| Endosulfan II | 0.0018 | ND | ND | ND | ND |
| Endosulfan sulfate | 0.0032 | ND | ND | ND | ND |
| Endrin ketone | 0.0026 | ND | ND | ND | ND |
| gamma-BHC (Lindane) | ND | ND | ND | ND | ND |
| Heptachlor | 0.001 | ND | ND | ND | ND |
| Heptachlor epoxide | 0.00076 | ND | ND | ND | ND |
| Methoxychlor | 0.012 | ND | ND | ND | ND |
| <i>Metals</i> | | | | | |
| Arsenic | 3.4 | BRBC | ND | ND | ND |
| Barium | 29.5 | 24.5 | ND | BRBC | BRBC |
| Cadmium | 0.2 | BRBC | ND | ND | ND |
| Chromium | 47.5 | 22.5 | ND | ND | BRBC |
| Lead | 19.7 | BRBC | ND | 12.6 ^a | ND |
| Mercury | 0.04 | 0.23 | ND | 0.21 | ND |
| Selenium | BRBC | BRBC | ND | 2.5 | ND |

^aMaximum concentration detected excluding data from the site-specific background location (MW5).

BRBC = Below reference background criteria.

ND = Not detected.

NP = No pathway exists.

**Table 10.1-11. GSSL Screening of Site-related Contaminants
in Soil and Sediment, SWMU 2**

| Site-related Contaminant | Maximum Concentration | GSSL ^a | CMCOPC? |
|---|-----------------------|-------------------|---------|
| SOIL | | | |
| <i>Volatile Organic Compounds (mg/kg)</i> | | | |
| 2-Butanone ^b | 0.0076 | 7.685 | No |
| Acetone | 0.511 | 16 | No |
| <i>Semivolatile Organic Compounds (mg/kg)</i> | | | |
| Bis(2-ethylhexyl)phthalate | 1.1 | 3,600 | No |
| <i>Pesticides/PCBs (mg/kg)</i> | | | |
| 4,4'-DDD | 0.0032 | 16 | No |
| 4,4'-DDE | 0.01 | 54 | No |
| 4,4'-DDT | 0.0089 | 32 | No |
| Aldrin | 0.011 | 0.5 | No |
| alpha-BHC | 0.00056 | 0.0005 | Yes |
| alpha-Chlordane | 0.00095 | 10 | No |
| delta-BHC ^b | 0.0016 | 0.000638 | Yes |
| Dieldrin | 0.003 | 0.004 | No |
| Endosulfan II ^c | 0.0018 | 18 | No |
| Endosulfan sulfate ^c | 0.0032 | 18 | No |
| Endrin ketone ^d | 0.0026 | 1 | No |
| Heptachlor | 0.001 | 23 | No |
| Heptachlor epoxide | 0.00076 | 0.7 | No |
| Methoxychlor | 0.012 | 160 | No |
| <i>Metals (mg/kg)</i> | | | |
| Arsenic | 3.4 | 1 | Yes |
| Barium | 29.5 | 82 | No |
| Cadmium | 0.2 | 0.4 | No |
| Chromium | 47.5 | 2 | Yes |
| Lead ^e | 19.7 | 400 | No |
| Mercury | 0.23 | 0.1 | Yes |
| SEDIMENT | | | |
| <i>Pesticides/PCBs (mg/kg)</i> | | | |
| alpha-Chlordane | 0.00071 | 10 | No |

^aGSSL = EPA GSSL with a DAF of 20 for volatile and semivolatile organics, pesticides, and explosives and a DAF of 1 for inorganics (average pH of groundwater is less than 5; Table 10.1-5); unless otherwise indicated (SAIC 1999a), GSSL is taken from Soil Screening Guidance: Technical Background Document (EPA 1996a).

^bEPA-suggested GSSL is not available; GSSL is calculated following Soil Screening Guidance: Technical Background Document (EPA 1996a). GSSLs are back-calculated from MCL, if available; otherwise, GSSLs are back-calculated based on EPA Region III RBCs corresponding to 10⁻⁶ risk or HQ = 1 (SAIC 1999a).

^cBased on the GSSL of endosulfan.

^dBased on the GSSL of endrin.

^eA screening level of 400 mg/kg is used for lead based on Revised Interim Soil Lead Guidance for CERCLA Sites and RCRA Corrective Action Facilities (EPA 1994e).

Table 10.1-12. Human Health Risk Screening for Surface Soil, Subsurface Soil, Groundwater, and Sediment, SWMU 2

| SURFACE SOIL | | | | | | |
|---|---------------------------|----------------|----------------|----------------------------|---------|----------------------------|
| Analyte | Results > Detection Limit | Minimum Detect | Maximum Detect | EPA Region III Residential | HHCOPC? | Justification |
| <i>Volatile Organic Compounds (mg/kg)</i> | | | | | | |
| 2-Butanone | 1/10 | 0.0055 | 0.0055 | 4,700 | No | Max Detect < Risk Criteria |
| Acetone | 3/10 | 0.174 | 0.511 | 780 | No | Max Detect < Risk Criteria |
| <i>Semivolatile Organic Compounds (mg/kg)</i> | | | | | | |
| Bis(2-ethylhexyl)phthalate | 1/10 | 1.1 | 1.1 | 46 | No | Max Detect < Risk Criteria |
| <i>Pesticides/PCBs (mg/kg)</i> | | | | | | |
| 4,4'-DDD | 1/10 | 0.0032 | 0.0032 | 2.7 | No | Max Detect < Risk Criteria |
| 4,4'-DDE | 6/10 | 0.0011 | 0.01 | 1.9 | No | Max Detect < Risk Criteria |
| 4,4'-DDT | 4/10 | 0.0018 | 0.0042 | 1.9 | No | Max Detect < Risk Criteria |
| Aldrin | 1/10 | 0.0011 | 0.0011 | 0.04 | No | Max Detect < Risk Criteria |
| alpha-BHC | 1/10 | 0.00024 | 0.00024 | 0.10 | No | Max Detect < Risk Criteria |
| alpha-Chlordane | 1/10 | 0.00095 | 0.00095 | 1.8 ^a | No | Max Detect < Risk Criteria |
| delta-BHC | 1/10 | 0.0016 | 0.0016 | 0.35 ^b | No | Max Detect < Risk Criteria |
| Dieldrin | 1/10 | 0.003 | 0.003 | 0.040 | No | Max Detect < Risk Criteria |
| Endosulfan II | 1/10 | 0.0018 | 0.0018 | 47 ^c | No | Max Detect < Risk Criteria |
| Endosulfan sulfate | 1/10 | 0.0032 | 0.0032 | 47 | No | Max Detect < Risk Criteria |
| Endrin ketone | 1/10 | 0.0026 | 0.0026 | 2.3 ^d | No | Max Detect < Risk Criteria |
| Heptachlor | 1/10 | 0.001 | 0.001 | 0.14 | No | Max Detect < Risk Criteria |
| Heptachlor epoxide | 3/10 | 0.00018 | 0.00076 | 0.070 | No | Max Detect < Risk Criteria |
| Methoxychlor | 5/10 | 0.0029 | 0.012 | 39 | No | Max Detect < Risk Criteria |
| <i>Metals (mg/kg)</i> | | | | | | |
| Arsenic | 10/10 | 0.71 | 3.4 | 0.43 | Yes | Max Detect > Risk Criteria |
| Barium | 10/10 | 13.6 | 29.5 | 550 | No | Max Detect < Risk Criteria |
| Cadmium | 2/10 | 0.08 | 0.2 | 3.9 | No | Max Detect < Risk Criteria |
| Chromium | 10/10 | 2.3 | 47.5 | 23 | Yes | Max Detect > Risk Criteria |
| Lead | 10/10 | 3.1 | 19.7 | 400 | No | Max Detect < Risk Criteria |
| Mercury | 8/10 | 0.02 | 0.04 | 2.3 | No | Max Detect < Risk Criteria |

| SUBSURFACE SOIL | | | | | | |
|---|---------------------------|----------------|----------------|----------------------------|---------|----------------------------|
| Analyte | Results > Detection Limit | Minimum Detect | Maximum Detect | EPA Region III Residential | HHCOPC? | Justification |
| <i>Volatile Organic Compounds (mg/kg)</i> | | | | | | |
| 2-Butanone | 1/5 | 0.0076 | 0.0076 | 4,700 | No | Max Detect < Risk Criteria |

Note: Footnotes appear on page 10.1-31.

Table 10.1-12. Human Health Risk Screening for Surface Soil, Subsurface Soil, Groundwater, and Sediment, SWMU 2 (continued)

| SUBSURFACE SOIL (continued) | | | | | |
|---|---------------------------|----------------|----------------|----------------------------|-------------------------------|
| Analyte | Results > Detection Limit | Minimum Detect | Maximum Detect | EPA Region III Residential | HHCOPC? Justification |
| <i>Semivolatile Organic Compounds (mg/kg)</i> | | | | | |
| Bis(2-ethylhexyl)phthalate | 1/7 | 0.229 | 0.229 | 46 | No Max Detect < Risk Criteria |
| <i>Pesticides/PCBs (mg/kg)</i> | | | | | |
| 4,4'-DDE | 1/7 | 0.0088 | 0.0088 | 1.9 | No Max Detect < Risk Criteria |
| 4,4'-DDT | 1/7 | 0.0089 | 0.0089 | 1.9 | No Max Detect < Risk Criteria |
| alpha-BHC | 1/7 | 0.00056 | 0.00056 | 0.10 | No Max Detect < Risk Criteria |
| <i>Metals (mg/kg)</i> | | | | | |
| Barium | 7/7 | 1.3 | 24.5 | 550 | No Max Detect < Risk Criteria |
| Chromium | 7/7 | 3.4 | 22.5 | 23 | No Max Detect > Risk Criteria |
| Mercury | 3/7 | 0.03 | 0.23 | 2.3 | No Max Detect < Risk Criteria |

| GROUNDWATER | | | | | |
|--|---------------------------|----------------|----------------|-----------------------|-------------------------------|
| Analyte | Results > Detection Limit | Minimum Detect | Maximum Detect | Human Health Criteria | HHCOPC? Justification |
| <i>Volatile Organic Compounds (µg/L)</i> | | | | | |
| 4-Methyl-2-pentanone | 1/12 | 9.9 | 9.9 | 13.9 | No Max Detect < Risk Criteria |
| Toluene | 1/12 | 15.6 | 15.6 | 75 | No Max Detect < Risk Criteria |
| Xylenes, total | 3/12 | 4 | 15.3 | 1,200 | No Max Detect < Risk Criteria |
| <i>Metals (µg/L)</i> | | | | | |
| Lead | 4/6 | 2.1 | 12.6 | 15 | No Max Detect < Risk Criteria |
| Mercury | 3/6 | 0.18 | 0.21 | 0.37 | No Max Detect < Risk Criteria |
| Selenium | 1/6 | 2.5 | 2.5 | 18 | No Max Detect < Risk Criteria |

| SEDIMENT | | | | | |
|--------------------------------|---------------------------|----------------|----------------|----------------------------|-------------------------------|
| Analyte | Results > Detection Limit | Minimum Detect | Maximum Detect | EPA Region III Residential | HHCOPC? Justification |
| <i>Pesticides/PCBs (mg/kg)</i> | | | | | |
| alpha-Chlordane | 1/1 | 0.00071 | 0.00071 | 0.49 | No Max Detect < Risk Criteria |

^aThe screening value for chlordane was used for alpha-chlordane.^bThe screening value for technical-BHC was used for delta-BHC.^cThe screening value for endosulfan was used for endosulfan II.^dThe screening value for endrin was used for endrin ketone.

**Table 10.1-13. Ecological Screening Value Comparison for
Analytes Detected in Sediment, SWMU 2**

| Analyte | SWMU 2 Maximum | ESV | ECOPC Aquatic Biota? | Justification |
|--------------------------------|-------------------|---------------------|----------------------------|------------------|
| <i>Pesticides/PCBs (mg/kg)</i> | | | | |
| alpha-Chlordane ^a | 0.00071 | 0.0043 ^a | No | Max Detect < ESV |

^aEPA Region IV ESV value for chlordane.

ESV = EPA Region IV ESVs (EPA 1996d) and, where indicated, alternative values for analytes without ESVs.

**Table 10.1-14. Ecological Screening Value Comparison for
Analytes Detected in Groundwater, SWMU 2**

| Analyte | SWMU 2 Maximum | ESV | ECOPC Aquatic Biota? | Justification |
|--|-------------------|-------------------|----------------------------|------------------|
| <i>Volatile Organic Compounds (µg/L)</i> | | | | |
| 4-Methyl-2-pentanone | 9.9 | 170 ^a | No | Max Detect < ESV |
| Toluene | 15.6 | 175 | No | Max Detect < ESV |
| Xylenes, total | 15.3 | 1.8 ^a | Yes | Max Detect > ESV |
| <i>Metals (µg/L)</i> | | | | |
| Lead | 12.6 | 1.32 ^b | Yes | Max Detect > ESV |
| Mercury | 0.21 | 0.0123 | Yes | Max Detect > ESV |
| Selenium | 2.5 | 5 | No | Max Detect < ESV |

^aChronic National Ambient Water Quality Criteria or Tier II values as reported in Suter and Tsao (1996), Table 1 or Table 3.

^bHardness dependent; assumes 50 µg/L CaCO₃.

ESV = EPA Region IV ESVs (EPA 1996d) and, where indicated, alternative values for analytes without ESVs. Cells with double borders indicate concentrations exceeding ESV or, when there is no ESV, compounds that become ECOPCs by default.

Table 10.1-15. Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 2

| ECOPC | C _{Max} (mg/kg) | BAF _i | Short-tailed Shrew | | | American Robin | | |
|--------------------------------|-----------------------------|------------------|--|------------------|---------------------------|--|------------------|---------------------------|
| | | | ADD (mg/kg/d) = C _{Max} × BAF _i × IR _s | TRV (mg/kg/d) | HQ (mg/kg/d) = ADD/TRV | ADD (mg/kg/d) = C _{Max} × BAF _i × IR _R | TRV (mg/kg/d) | HQ (mg/kg/d) = ADD/TRV |
| | | | Volatile Organic Compounds | | | | | |
| 2-Butanone | 0.0055 | 5.00E-02 | 1.46E-04 | 3.89E+03 | 3.74E-08 | 4.18E-04 | No TRV | No HQ |
| Acetone | 0.511 | 5.00E-02 | 1.35E-02 | 2.20E+01 | 6.16E-04 | 3.88E-02 | No TRV | No HQ |
| | | | HI= 7.46E-04 | | | HI= 0.00E+00 | | |
| Semivolatile Organic Compounds | | | | | | | | |
| Bis(2-ethylhexyl)phthalate | 1.1 | 5.00E-02 | 2.92E-02 | 2.18E+01 | 1.34E-03 | 8.36E-02 | 1.10E+00 | 7.60E-02 |
| Pesticides/PCBs | | | | | | | | |
| 4,4'-DDD ^a | 0.0032 | 9.19E-02 | 1.56E-04 | 5.45E+00 | 2.86E-05 | 4.47E-04 | 2.80E-03 | 1.60E-01 |
| 4,4'-DDE ^a | 0.01 | 3.37E-01 | 1.79E-03 | 5.45E+00 | 3.28E-04 | 5.12E-03 | 2.80E-03 | 1.83E+00 |
| 4,4'-DDT ^a | 0.0042 | 4.71E-02 | 1.05E-04 | 1.24E-01 | 8.48E-04 | 3.01E-04 | 2.80E-03 | 1.07E-01 |
| Aldrin | 0.0011 | 5.60E-01 | 3.26E-04 | 4.40E-01 | 7.43E-04 | 9.36E-04 | 1.73E+00 | 5.41E-04 |
| alpha-BHC | 0.00024 | 2.60E+00 | 3.31E-04 | 3.52E+00 | 9.40E-05 | 9.48E-04 | 5.63E-01 | 1.68E-03 |
| alpha-Chlordane | 0.00095 | 1.60E+00 | 8.06E-04 | 5.45E+00 | 1.48E-04 | 2.31E-03 | 2.14E+00 | 1.08E-03 |
| delta-BHC | 0.0016 | 2.60E+00 | 2.20E-03 | 3.52E+00 | 6.27E-04 | 6.32E-03 | 5.63E-01 | 1.12E-02 |
| Dieldrin | 0.003 | 5.50E+00 | 8.75E-03 | 4.40E-02 | 1.99E-01 | 2.51E-02 | 7.70E-02 | 3.26E-01 |
| Endosulfan II | 0.0018 | 5.50E+00 | 5.25E-03 | 3.30E-01 | 1.59E-02 | 1.50E-02 | 1.00E+01 | 1.50E-03 |

Note: Footnotes appear on page 10.1-34.

Table 10.1-15. Preliminary Risk Calculations for ECOPCs in Surface Soil, SWMU 2 (continued)

| ECOPC | C_{Max} (mg/kg) | BAF_i | Short-tailed Shrew | | | American Robin | | |
|---------------------------------|----------------------|----------|---|------------------|-------------------|---|------------------|-------------------|
| | | | ADD (mg/kg/d) $= C_{Max} \times$ $BAF_i \times IR_S$ | TRV (mg/kg/d) | HQ $= ADD/TRV$ | ADD (mg/kg/d) $= C_{Max} \times$ $BAF_i \times IR_R$ | TRV (mg/kg/d) | HQ $= ADD/TRV$ |
| Endosulfan sulfate | 0.0032 | 5.50E+00 | 9.33E-03 | 3.30E-01 | 2.83E-02 | 2.68E-02 | 1.00E+01 | 2.68E-03 |
| Endrin ketone | 0.0026 | 1.90E+00 | 2.62E-03 | 1.09E-01 | 2.39E-02 | 7.51E-03 | 1.04E-02 | 7.25E-01 |
| Heptachlor ^a | 0.001 | 1.00E+00 | 5.30E-04 | 1.09E-01 | 4.84E-03 | 1.52E-03 | No TRV | No HQ |
| Heptachlor epoxide ^b | 0.00076 | 1.00E+00 | 4.03E-04 | 1.09E-01 | 3.68E-03 | 1.16E-03 | No TRV | No HQ |
| Methoxychlor | 0.012 | 5.70E-01 | 3.63E-03 | 8.79E+00 | 4.12E-04 | 1.04E-02 | 8.42E+00 | 1.23E-03 |
| | | | HI= 2.79E-01 | | | HI= 3.17E+00 | | |
| <i>Metals</i> | | | | | | | | |
| Arsenic | 3.4 | 6.60E-03 | 1.19E-02 | 1.50E-01 | 7.94E-02 | 3.41E-02 | 5.14E+00 | 6.64E-03 |
| Barium | 29.5 | 7.50E-03 | 1.17E-01 | 1.17E+01 | 9.99E-03 | 3.36E-01 | 2.08E+01 | 1.61E-02 |
| Cadmium | 0.2 | 1.10E+01 | 1.17E+00 | 2.12E+00 | 5.50E-01 | 3.34E+00 | 1.45E+00 | 2.31E+00 |
| Chromium | 47.5 | 1.60E-01 | 4.03E+00 | 6.02E+03 | 6.70E-04 | 1.16E+01 | 1.00E+00 | 1.16E+01 |
| Lead ^c | 19.7 | 4.0E-01 | 4.18E+00 | 1.76E+01 | 2.38E-01 | 1.20E+01 | 1.13E+00 | 1.06E+01 |
| Mercury ^d | 0.04 | 3.40E-01 | 7.21E-03 | 2.89E+00 | 2.50E-03 | 2.07E-02 | 4.50E-01 | 4.59E-02 |

^aTRVs for 4,4'-DDD; 4,4'-DDE; and 4,4'-DDT are based on NOAEL for DDT and its metabolites (Sample, Opresko, and Suter 1996); see Tables 8-5 and 8-6.

^bDefault BAF; assumed to be 1.

^cCalcium-dependent BAF for lead (Corp and Morgan 1991, as cited in HAZWRAP 1994); default value = 0.4, assumes calcium concentration in soil > 500 mg/kg and lead concentration > 1 mg/kg.

^dAssumes mercuric sulfide for short-tailed shrew, mercuric chloride for American robin.

ADD = Average daily dose (mg/kg/d).

BAF_i = Soil-to-invertebrate bioaccumulation factor (HAZWRAP 1994).

C_{Max} = Maximum detected surface soil concentration (mg/kg).

HQ = Hazard quotient; HI = hazard index = sum of HQs.

IR_R = Robin food ingestion rate (kg/kgBW/d) = 1.52.

IR_S = Shrew food ingestion rate (kg/kgBW/d) = 0.53.

TRV = NOAEL (mg/kg/d); see Tables 8-5 and 8-6.

Cells with double borders indicate HQ > 1.

^aTRVs for 4,4'-DDD; 4,4'-DDE; and 4,4'-DDT are based on NOAEL for DDT and its metabolites (Sample, Opresko, and Suter 1996); see Tables 8-5 and 8-6.

^bDefault BAF; assumed to be 1.

^cCalcium-dependent BAF for lead (Corp and Morgan 1991, as cited in HAZWRAP 1994); default value = 0.4, assumes calcium concentration in soil > 500 mg/kg and lead concentration > 1 mg/kg.

^dAssumes mercuric sulfide for short-tailed shrew, mercuric chloride for American robin.

ADD = Average daily dose (mg/kg/d).

BAF_i = Soil-to-invertebrate bioaccumulation factor (HAZWRAP 1994).

C_{Max} = Maximum detected surface soil concentration (mg/kg).

HQ = Hazard quotient; HI = hazard index = sum of HQs.

IR_R = Robin food ingestion rate (kg/kgBW/d) = 1.52.

IR_S = Shrew food ingestion rate (kg/kgBW/d) = 0.53.

TRV = NOAEL (mg/kg/d); see Tables 8-5 and 8-6.

Cells with double borders indicate HQ > 1.

Table 10.1-16. Preliminary Risk Calculations for ECOPCs in Groundwater, SWMU 2

| ECOPC | C_{Max} ($\mu\text{g/L}$) | BCF | Raccoon | | | Mink | | | Green Heron | | |
|----------------------|----------------------------------|-------|--|------------------|-----------------|--|------------------|-----------------|--|------------------|-----------------|
| | | | ADD (mg/kg/d) $= C_{Max} \times$ $0.001 \times$ IR_W | TRV (mg/kg/d) | HQ = ADD/TRV | ADD (mg/kg/d) $= C_{Max} \times$ $BCF \times$ $0.001 \times$ IR_M | TRV (mg/kg/d) | HQ = ADD/TRV | ADD (mg/kg/d) $= C_{Max} \times$ $0.001 \times$ $BCF \times$ IR_H | TRV (mg/kg/d) | HQ = ADD/TRV |
| | | | Volatile Organic Compounds | | | | | | | | |
| Xylenes, total | 15.3 | 17 | 1.22E-03 | 1.08E-01 | 1.14E-02 | 3.56E-02 | 1.69E-01 | 2.11E-01 | 4.99E-02 | No TRV | -- |
| Metals | | | | | | | | | | | |
| Lead | 12.6 | 300 | 1.01E-03 | 3.93E+00 | 2.56E-04 | 5.18E-01 | 6.15E+00 | 8.42E-02 | 7.26E-01 | 1.13E+00 | 6.42E-01 |
| Mercury ^a | 0.21 | 63000 | 1.68E-05 | 6.46E-01 | 2.60E-05 | 1.81E+00 | 1.01E+00 | 1.79E+00 | 2.54E+00 | 4.50E-01 | 5.64E+00 |

^aAssumes mercuric sulfide for raccoon, methyl mercury for mink and heron.

0.001 (mg/ μg) = Conversion from μg to mg.

ADD = Average daily dose (mg/kg/d).

BCF = Water-to-fish bioconcentration factor (HAZWWRAP 1994).

C_{Max} = Maximum detected concentration ($\mu\text{g/L}$).

HQ = Hazard quotient; HI = hazard index = sum of HQs.

IR_H = Heron food ingestion rate (kg/kg/d) = 0.192.

IR_M = Mink food ingestion rate (kg/kg/d) = 0.137.

IR_W = Raccoon water ingestion rate (L/kg/d) = 0.080.

TRV = NOAEL (mg/kg/d).

-- = Cannot be calculated due to the lack of data.

Cells with double borders indicate HQ > 1.

Table 10.1-17. Supplemental Risk Calculations for ECOPCs in Surface Soil for American Robin, SWMU 2

| American Robin | | | | | | | | | |
|-----------------|---------------------------------|-----------------|--|------------------|---|--|---|--|--------------------------------|
| ECOPC | Site Concentration Mean (mg/kg) | SP _r | ADD _p (mg/kgBW/d) Mean × SP _r × I _p × AUF | BAF _i | ADD _A (mg/kgBW/d) Mean × BAF _i × I _A × AUF | ADD _S (mg/kgBW/d) Mean × I _S × AUF | ADD _{total} (mg/kgBW/d) ADD _p + ADD _A + ADD _S | Body-weight-adjusted LOAEL TRV (mg/kgBW/d) | HQ = ADD _{total} /TRV |
| Inorganics | | | | | | | | | |
| Cadmium | 4.45E-02 | 3.00E-02 | 7.93E-04 | 1.10E+01 | 2.91E-01 | 5.50E-03 | 2.97E-01 | 2.00E+01 | 1.49E-02 |
| Chromium | 1.45E+01 | 9.00E-04 | 7.75E-03 | 1.60E-01 | 1.38E+00 | 1.79E+00 | 3.18E+00 | 5.00E+00 | 6.35E-01 |
| Lead | 6.61E+00 | 1.80E-03 | 7.07E-03 | 4.00E-01 | 1.57E+00 | 8.17E-01 | 2.39E+00 | 1.13E+01 | 2.12E-01 |
| Organics | | | | | | | | | |
| Pesticides/PCBs | | | | | | | | | |
| 4,4'-DDE | 3.73E-03 | 2.00E-03 | 4.43E-06 | 1.70E+00 | 3.77E-03 | 4.61E-04 | 4.23E-03 | 2.80E-02 | 1.51E-01 |

ADD_A = Average daily dose; animal.

ADD_p = Average daily dose; plant.

ADD_S = Average daily dose; soil.

ADD_{total} = Average daily dose; total.

AF = Animal fraction.

AUF = 9.90E-01.

BAF_i = Soil-to-animal bioaccumulation factor; invertebrates.

HQ = Hazard quotient.

I_A = TUF × IR_f × AF.

I_A (kg/kgBW/d) = 6.00E-01.

I_p = TUF × IR_f × PF.

I_p (kg/kgBW/d) = 6.00E-01.

I_S = TUF × IR_f × SF.

I_S (kg/kgBW/d) = 1.25E-01.

IR_f = Food ingestion rate.

PF = Plant fraction.

SF = Soil fraction.

SP_r = Soil-to-plant bioaccumulation factor; reproductive parts.

TRV = Toxicity reference value.

TUF = Temporal use factor.

Table 10.1-18. Supplemental Risk Calculations for ECOPCs in Groundwater for Mink, SWMU 2

| Mink | | | | | | | |
|------------|---|----------|--|--|--|--|-----------------------------------|
| ECOPC | Site Concentration Mean (µg/L) | BCF | ADD _A | ADD _w | ADD _{total} | Body-weight- adjusted LOAEL TRV (mg/kgBW/d) | HQ = ADD _{total} /TRV |
| | | | (mg/kgBW/d) Mean × 0.001 × BCF × I _A × AUF | (mg/kgBW/d) Mean × 0.001 × IR _w × AUF | (mg/kgBW/d) ADD _A + ADD _w | | |
| Inorganics | | | | | | | |
| Mercury | 1.20E-01 | 6.30E+04 | 8.29E-02 | 9.50E-07 | 8.29E-02 | 1.01E+01 | 8.20E-03 |

0.001 = Conversion from µg/L to mg/L.

ADD_A = Average daily dose; animal.

ADD_{total} = Average daily dose; total.

ADD_w = Average daily dose; drinking water.

AF = Animal fraction.

AUF = 8.00E-02. AUF assumes 328 feet (100 meters) of stream are contaminated at mean concentration.

BCF = Water-to-animal bioconcentration factor.

HQ = Hazard quotient.

I_A = TUF × IR_r × AF.

I_A(kg/kgBW/d) = 1.37E-01.

IR_r = Food ingestion rate.

IR_w(L/kgBW/d) = 9.90E-02.

TRV = Toxicity reference value.

TUF = Temporal use factor.

Table 10.1-19. Supplemental Risk Calculations for ECOPCs in Groundwater for Green Heron, SWMU 2

| Green Heron | | | | | | |
|-------------|--------------------------------|----------|--|---|--|---|
| ECOPC | Site Concentration Mean (µg/L) | BCF | ADD _A (mg/kgBW/d) Mean × 0.001 × BCF × I _A × AUF | ADD _W (mg/kgBW/d) Mean × 0.001 × IR _W × AUF | ADD _{total} (mg/kgBW/d) ADD _A + ADD _W | Body-weight-adjusted LOAEL TRV (mg/kgBW/d) HQ = ADD _{total} /TRV |
| Inorganics | | | | | | |
| Mercury | 1.20E-01 | 6.30E+04 | 1.97E+00 | 1.13E-05 | 1.97E+00 | 9.00E-01 |

0.001 = Conversion from µg/L to mg/L.

ADD_A = Average daily dose; animal.

ADD_{total} = Average daily dose; total.

ADD_W = Average daily dose; drinking water.

AF = Animal fraction.

AUF = 1.00E+00. AUF assumes 328 feet (100 meters) of stream are contaminated at mean concentration.

BCF = Water-to-animal bioconcentration factor.

I_A = TUF × IR_r × AF.

I_A(kg/kgBW/d) = 2.60E-01.

IR_r = Food ingestion rate.

IR_W(L/kgBW/d) = 9.40E-02.

TRV = Toxicity reference value.

TUF = Temporal use factor.

Cells with double border indicate HQ > 1.

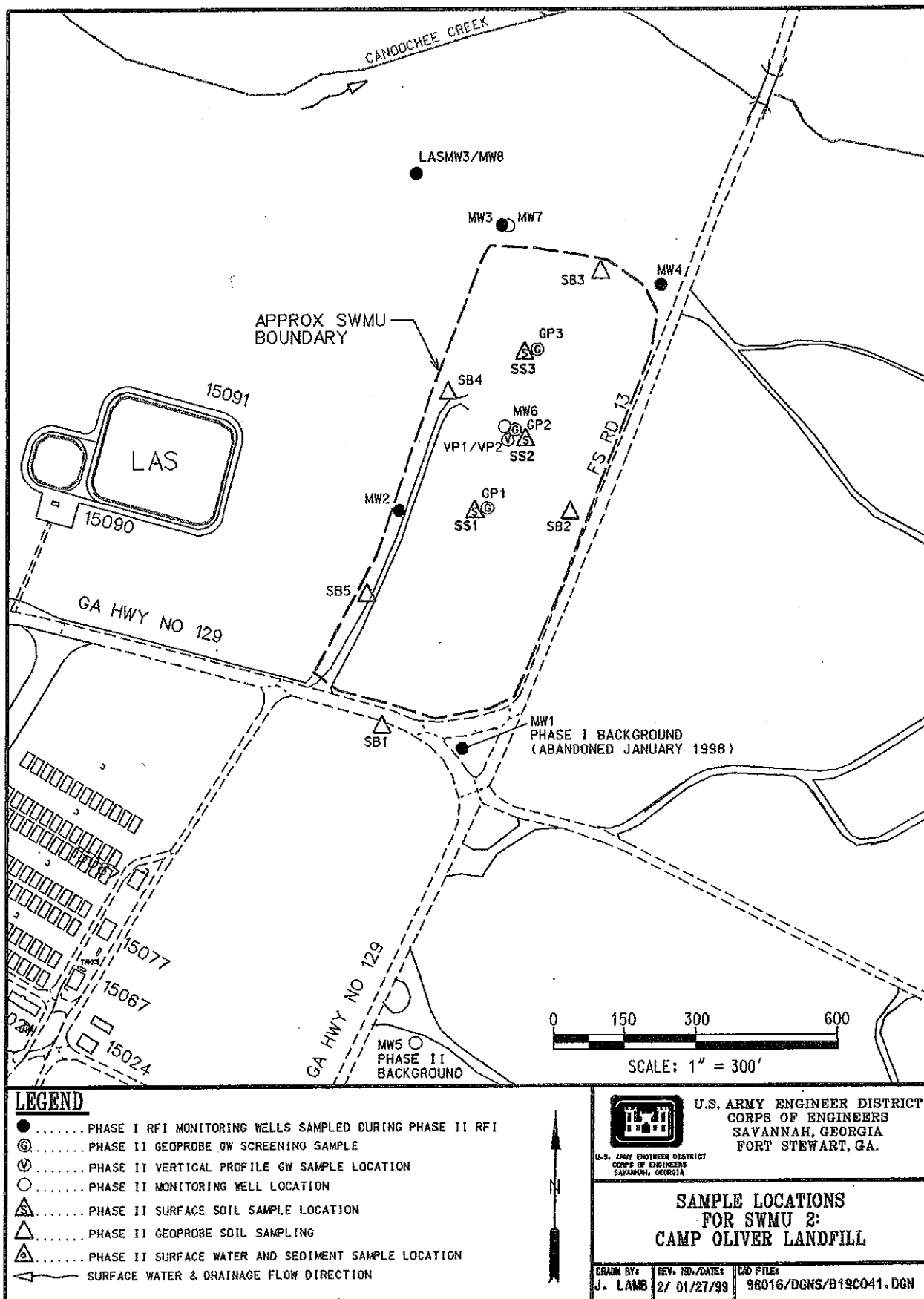


Figure 10.1-1. Sampling Locations and Phase I RFI Analytical Results in Surface Soil, SWMU 2

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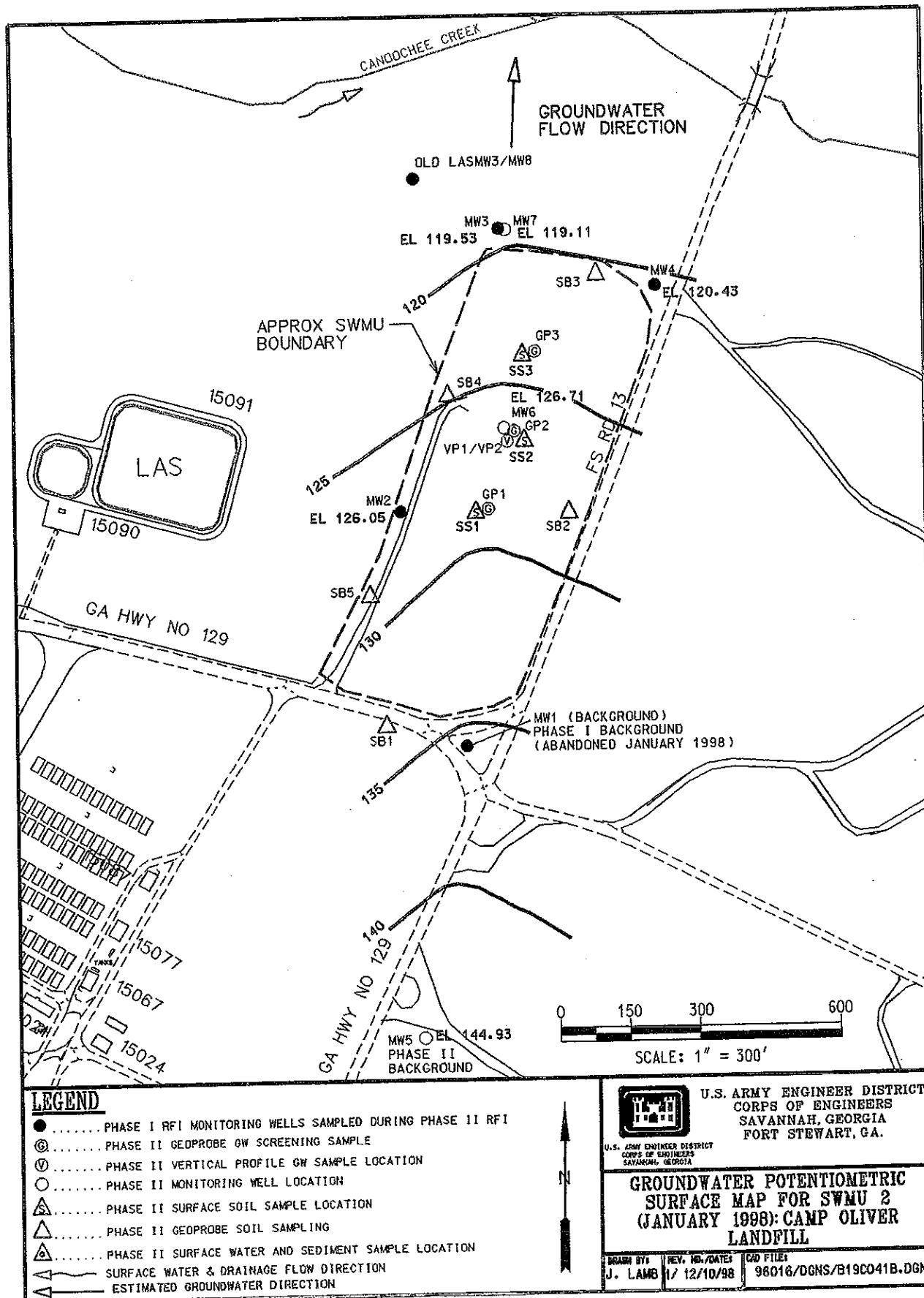


Figure 10.1-4. Groundwater Potentiometric Surface Map, SWMU 2

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- = Exposure pathway not complete.
- * = Exposure pathway complete.
- 1 = Exposure pathway evaluated for future screening only.
- 2 = This scenario is used to derive screening values. It is not a viable scenario for this site.

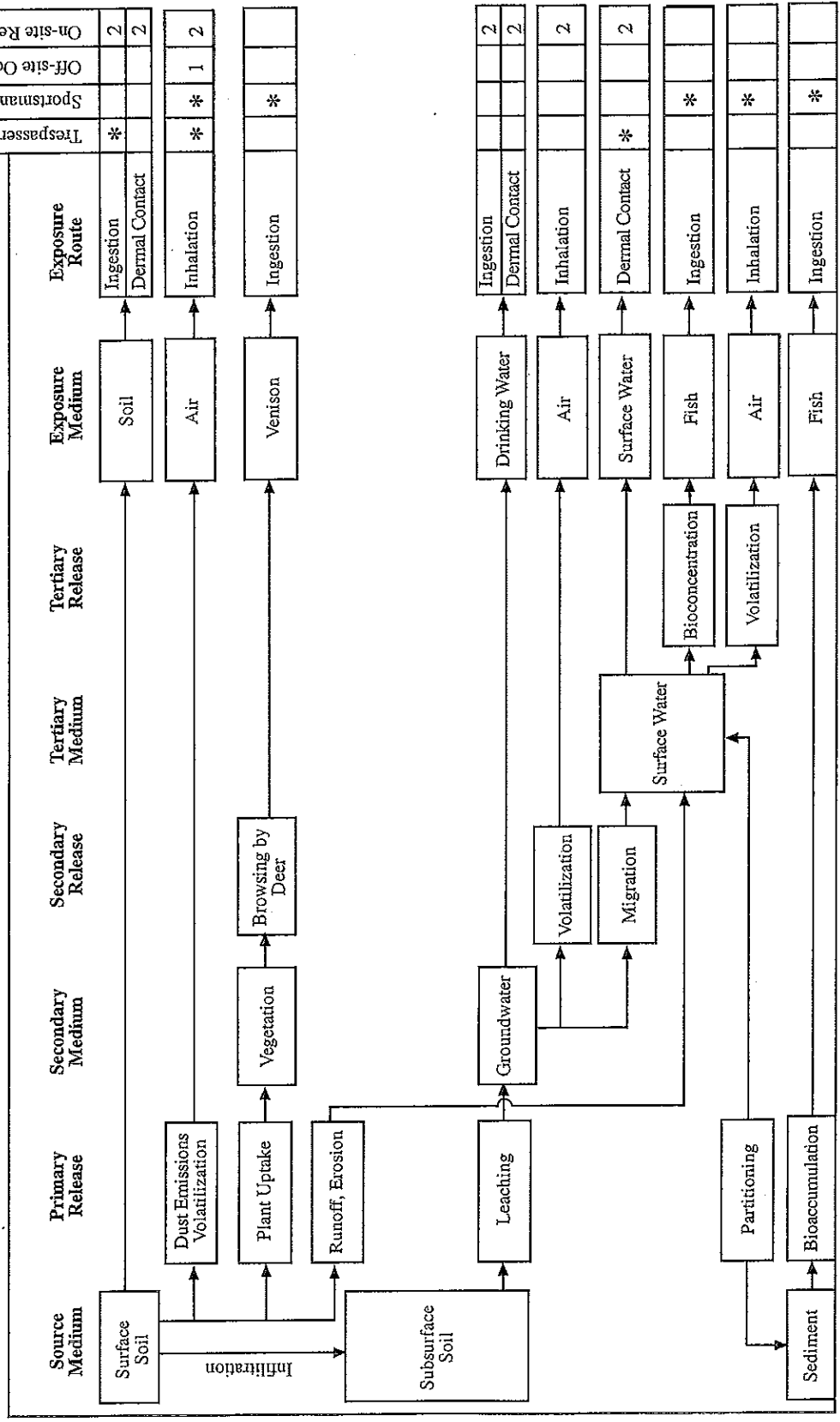
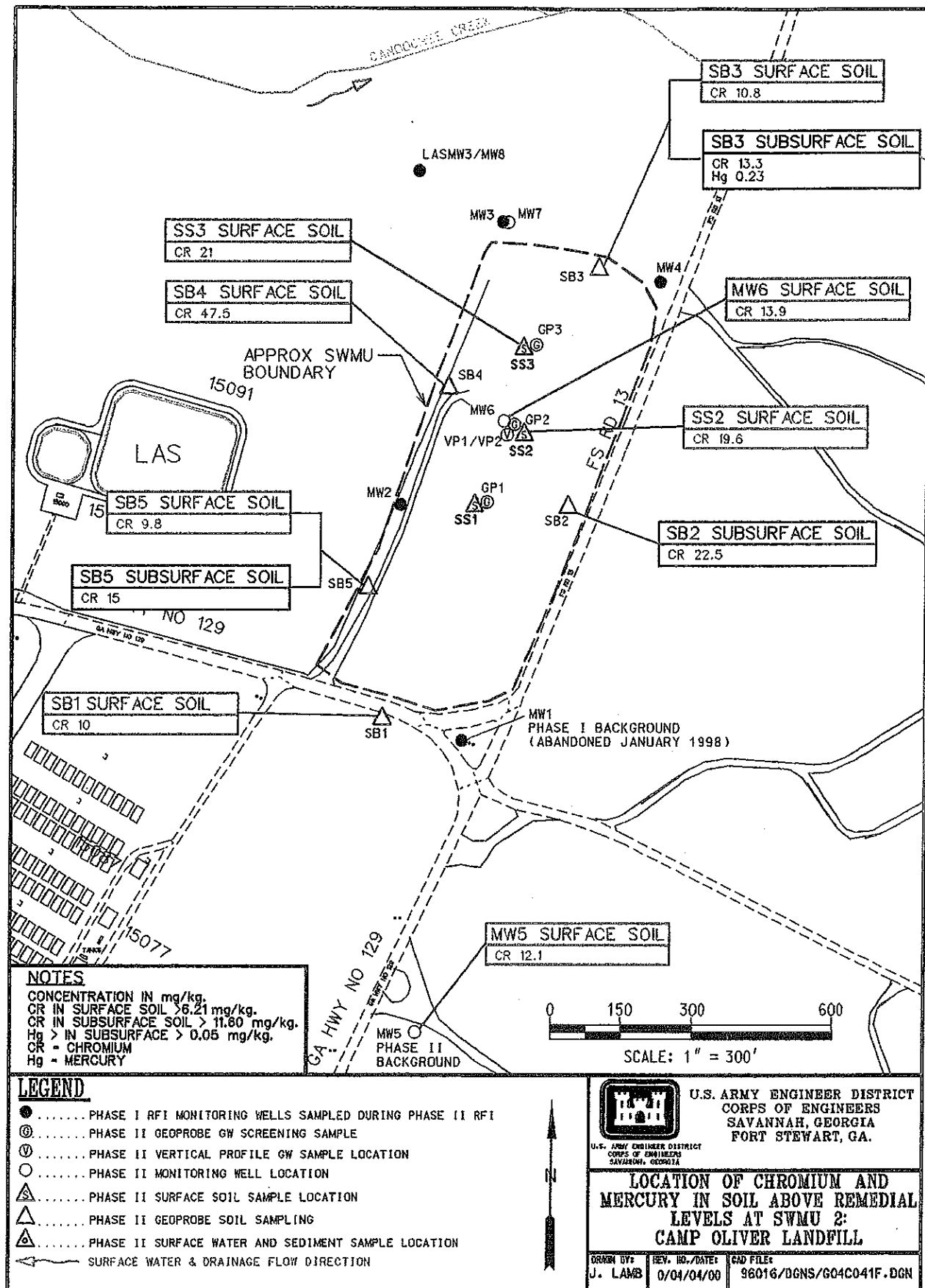


Figure 10.1-9. Potential Migration and Exposure Pathways, SWMU 2



ATTACHMENT 10.1A
SWMU 2: CAMP OLIVER LANDFILL
HUMAN HEALTH BASELINE RISK ASSESSMENT

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ATTACHMENT 10.1A

SWMU 2: CAMP OLIVER LANDFILL

HUMAN HEALTH BASELINE RISK ASSESSMENT

The human health baseline risk assessment consists of five elements: (1) identification of COPCs, (2) exposure assessment, (3) toxicity assessment, (4) risk characterization, and (5) assessment of uncertainty.

10.1A.1 IDENTIFICATION OF COPCS

The CMCOPCs and HHCOPCs have been discussed in the sections on contaminant fate and transport (Section 10.1.5) and the HHPRE (Section 10.1.6), respectively. The CMCOPCs include arsenic, chromium, mercury, and two pesticides (alpha-BHC and delta-BHC). Arsenic and chromium were identified as HHCOPCs in surface soils (Section 10.1.8.1). No COPCs were identified in the other media (see Section 10.1.8.1).

10.1A.2 EXPOSURE ASSESSMENT

The exposure assessment quantifies the amount of a COPC an individual may come in contact with at each site. The exposure assessment considers all pathways of potential human exposure, the magnitude of exposure, and the frequency and duration of exposure. The process for estimating exposure consists of the following elements: (1) characterization of the exposure setting in terms of the physical and demographic characteristics of the site, (2) identification of receptor populations, (3) identification of the exposure pathways by which an individual may come in contact with a COPC, (4) estimation of the exposure point concentration, and (5) quantification of the intake or dose to which an individual may be exposed.

10.1A.2.1 Exposure Setting

The exposure setting describes the physical features at the site that are important when identifying the human populations that may be exposed to COPCs, either currently or in the future.

The landfill is located approximately 17 miles northwest of the Fort Stewart garrison area. It is just north of the bivouac area, in an area used for training. The site is approximately 2 acres in size. The area is covered with grass, small trees, and bushes.

There are no surface water bodies present within the site boundary; however, Canoochee Creek runs within 450 feet of the northern boundary of SWMU 2.

10.1A.2.2 Identification of Potential Receptor Populations and Exposure Pathways

A complete exposure pathway consists of four elements: (1) a source of contamination, (2) a transport or retention medium, (3) a point of contact with the chemical, and (4) a route of exposure (ingestion, dermal absorption, or inhalation) at the point of contact through which the chemical may be taken into the body. When all of these elements are present, the pathway is considered to be complete.

The CSM for risk assessment is shown in Figure 10.1A-1. This model illustrates the potential receptor populations and their routes of exposure for COPCs.

The following receptor populations were used to evaluate potential risks associated with current land use: an on-site Installation worker and a juvenile trespasser. At present, there is no work being done by FSMR at this site. However, a worker may be present on the site in the near future, and as a conservative measure, this receptor population will be addressed. The on-site Installation worker and the trespasser may be exposed to COPCs in surface soil via incidental ingestion and dermal contact. The site is currently vegetated; therefore, exposure via inhalation of fugitive dust is not considered to be a viable pathway.

The sportsman may cross the area while hunting. However, the COPCs in surface soils are not likely to bioaccumulate in game animals. The BAFs for the uptake of arsenic from soil into plants and from forage into venison are 0.04 and 0.002, respectively. Similarly the BAFs for chromium are 0.0075 for plants and 0.0055 for beef. Given that bioaccumulation is not likely to be a significant exposure pathway, the potential exposure pathways for the sportsman would be limited to direct contact with surface soils. Other receptor populations, such as the on-site Installation worker and the juvenile trespasser, are likely to come in greater contact with soils and are at greater potential risks; therefore, the sportsman will not be addressed at this site.

It is unlikely that COPCs are currently migrating off-site in significant concentrations. COPCs in surface soils may migrate via wind erosion or runoff into Canoochee Creek. However, the presence of vegetation is likely to limit migration via these pathways. In addition, sampling of Canoochee Creek failed to show significant concentrations of constituents (i.e., concentrations greater than their respective screening values). Therefore, there are no current exposure pathways for off-site receptors.

The potential on-site receptor populations for the future land-use scenario receptor populations include an Installation worker, a trespasser, and a resident. Although no changes in land use are expected at this site, for the purposes of this risk assessment it was assumed that the vegetative cover had been removed. The exposure pathways for the on-site Installation worker and the juvenile trespasser include all of the potential pathways addressed under the current scenarios, with the addition of exposure via inhalation of fugitive dust.

The on-site resident is presented for comparative purposes and is not considered to be a viable receptor population. Potential exposure pathways for the on-site resident include incidental ingestion, dermal contact, and inhalation of fugitive dust. If the site were developed for residential purposes, it would be landscaped and vegetated; therefore, exposure via inhalation of fugitive dust would be unlikely. However, as a conservative assumption, exposure via inhalation of fugitive dust will be evaluated.

COPCs in surface soil may migrate off-site via wind erosion. Based on this potential migration pathway, future off-site receptor populations include an Installation worker, a resident, a trespasser, a juvenile wading in surface water, and a sportsman. All of these receptors may be exposed via inhalation of fugitive dust. The Installation worker is at the greatest potential risk because of the amount of time this receptor spends in the area and the potential duration of the exposure; therefore, this receptor will be evaluated. An off-site resident will also be evaluated. The residential exposure scenario is used for comparative purposes only and is not considered to be a viable exposure scenario.

COPCs in groundwater may migrate to Canoochee Creek in the future. Potential off-site receptors include a child wading in the creek and a sportsman fishing. The exposure pathways for the child would be limited to dermal exposure. The sportsman might also be exposed via dermal contact and ingestion of fish that have bioaccumulated COPCs in their tissue.

10.1A.2.3 Estimation of Exposure Concentrations

The estimation of exposure concentrations for on-site receptors to COPCs in soil is discussed in Appendix I, Section I.2.3. Exposure concentrations were calculated using either analytical results or environmental fate and transport models. The analytical results from the surface soil samples were used to calculate the exposure concentrations for HHCOPCs in soil (arsenic and chromium). The exposure point concentrations are equal to the 95 percent upper confidence limit of the mean, unless this value was greater than the maximum detected concentration. If that was the case, then the exposure concentration defaulted to the maximum concentration. The selected exposure concentrations for the HHCOPCs are presented in Table 10.1A-1.

Exposure concentrations of fugitive dust in air were calculated using the formulas described in Appendix I, Section I.2.3. These values were based on the exposure concentrations for surface soil. For the purposes of estimating exposure of an off-site receptor to fugitive dust, it was assumed that no dilution of the air concentrations occurred and that the exposure concentrations for both on-site and off-site receptors were the same.

Potential groundwater concentrations resulting from leaching of CMCOPCs (arsenic, chromium, mercury, alpha-BHC, and delta-BHC) were estimated using the SESOIL leachate model (Appendix K). For the purposes of this risk assessment, the maximum concentrations of constituents detected above the water table were used as the source concentrations for the SESOIL model. The results of the SESOIL modeling are summarized in Table 10.1A-1.

Alpha-BHC was identified as a CMCOPC based on a maximum concentration of 0.00056 mg/kg. However, this sample was collected below the water table and would not be a source of alpha-BHC leaching into the groundwater from the unsaturated zone. The other soil samples collected above the water table did not have concentrations of alpha-BHC that exceeded its respective GSSL. In addition, alpha-BHC was not detected in groundwater at this site; therefore, alpha-BHC is not considered to be a CMCOPC and was not evaluated further.

Arsenic was identified as a CMCOPC based on the maximum concentration of arsenic in surface soils. However, the maximum concentration in surface soil (3.4 mg/kg) does not exceed the subsurface soil reference background criterion for arsenic, 8.04 mg/kg. Given the relative thickness of subsurface soil and the higher concentrations of arsenic present in this soil stratum, the contribution of arsenic to groundwater from surface soil is not likely to be significant. In addition, arsenic was not identified as a COPC in groundwater, indicating that arsenic is not leaching to groundwater in significant concentrations. Therefore, the potential for arsenic to leach into groundwater was not addressed further in the risk assessment.

Leachate modeling input parameters and results for delta-BHC, mercury, and chromium are presented in Appendix K, Tables K-2.1, K-3.1, and K-4.1. Modeling results indicated that delta-BHC naturally attenuates before reaching the water table; therefore, the potential risks associated with the leaching of delta-BHC into groundwater were not assessed.

The estimated (SESOIL model) maximum groundwater concentrations of mercury and chromium are 0.0025 mg/L and 1.73 mg/L, respectively. Both of these concentrations exceeded the respective groundwater screening values of 0.001 mg/L for mercury and 0.011 mg/L for chromium; therefore, these estimated groundwater concentrations from the SESOIL model (Appendix K, Table K-4.1 and Figures K-1.1 and K-1.2), were assessed in the risk characterization.

COPCs in groundwater may migrate to Canoochee Creek. The ODAST model was used to estimate groundwater concentrations of mercury and chromium at Canoochee Creek. For the purposes of this risk assessment, it was assumed that the concentrations in Canoochee Creek are equal to the concentrations in the adjacent groundwater. This assumption is conservative, given that it assumes that there is no dilution of the constituents upon discharge of groundwater into the surface water body.

In the ODAST modeling, it was assumed that the concentration of chromium at the source location remains constant at the SESOIL-predicted concentration for a period of 70 years. The ODAST model was simulated for a period of 1,000 years. The ODAST input parameters are presented in Appendix K, Table K-5.1. The ODAST modeling results (Appendix K, Table K-6.1) indicated that chromium does not migrate to Canoochee Creek in the 1,000-year simulation.

The maximum concentration of mercury in groundwater adjacent to Canoochee Creek was 0.00045 mg/L. Groundwater discharging into Canoochee Creek will be diluted by the surface water upstream at the point of discharge. For the purposes of this risk assessment, it was assumed that the mercury migrating from the groundwater to Canoochee Creek will not be diluted and that the mercury concentration in surface water is equal to the concentration in groundwater adjacent to the creek.

The potential receptor populations may potentially be exposed to COPCs in soils as a result of direct exposure (HHCOPCs) and leaching to groundwater (CMCOPC). The COPCs for each environmental medium are:

- HHCOPCs in surface soils: arsenic and chromium,
- CMCOPCs in soils leaching to groundwater: chromium and mercury, and
- CMCOPC in soils leaching to groundwater and migrating to surface water: mercury.

10.1A.2.4 Quantification of Exposure

The equations used to estimate exposures to receptor populations are discussed in Appendix I, Section I.2.4. The default exposure factors for the current on-site trespasser assume that the juvenile spends all of his time on the site. The exposure parameter values used to estimate potential exposure are given in Table 10.1A-2.

A potential intake was estimated for each receptor population for all applicable pathways. The estimated intakes for the current on-site receptors, the Installation worker and the juvenile trespasser, are given in Tables 10.1A-3 and 10.1A-4, respectively. There are no current off-site receptor populations.

Future on-site receptor populations include an Installation worker, a juvenile trespasser, and a resident. The estimated intakes for the Installation worker and the juvenile trespasser are given in Tables 10.1A-5 and 10.1A-6, respectively. The resident population is divided into a child and an adult resident because the differences in behavior, exposure duration, and physiology between an adult and a child result in different intakes. Because of the increased amount of hand-to-mouth behavior in children, the incidental soil ingestion for a child is twice that of an adult. Given the higher ingestion rate and the lower body weight of a child, the resident child is at greater risk from incidental ingestion of surface soil than the resident adult. However, the increased exposure duration for the adult makes this receptor more sensitive to carcinogenic effects for certain exposure pathways. For the purposes of this risk assessment, only carcinogenic risks will be calculated for the resident adult, given that the juvenile receptor is more sensitive to noncarcinogenic risks. However, given the child's increased exposure rates for soils, the adult is not always the most sensitive receptor for carcinogens; therefore, the potential carcinogenic and

noncarcinogenic risks will be calculated for the resident child. The estimated intakes for the child resident and the adult resident are given in Tables 10.1A-7 and 10.1A-8, respectively.

Future off-site receptors include an Installation worker, a resident child, a resident adult, a juvenile wader, and a sportsman. The estimated intakes for the Installation worker, resident child, resident adult, juvenile wader, and sportsman are given in Tables 10.1A-9, 10.1A-10, 10.1A-11, 10.1A-12, and 10.1A-13, respectively.

10.1A.3 TOXICITY ASSESSMENT

The purpose of the toxicity assessment is to determine the increased likelihood and magnitude of adverse human health effects based on the extent of exposure to contamination. The toxicity assessment for SWMU 2 was carried out as described in Appendix I, Section I.3. The toxicity values for arsenic, chromium, and mercury are given in Table 10.1A-14.

Elemental mercury does not have an oral reference dose listed in either IRIS or HEAST (EPA 1999a; EPA 1997c). An oral reference dose has been developed for mercuric chloride; therefore, this value was used to calculate oral HQs and the dermal reference dose for mercury.

10.1A.4 RISK CHARACTERIZATION RESULTS

The risk characterization followed the procedure outlined in Appendix I, Section I.4. Quantitative estimates of noncarcinogenic risks were calculated for acetone for each potentially complete exposure pathway.

10.1A.4.1 Current Land-use Scenarios

Current potential receptors are the Installation worker and the juvenile trespasser. There is no current use of groundwater at this site; therefore, current land use receptors are not at risk from exposure to chromium in groundwater. The potential risks to each of these receptor populations are discussed below.

On-site Installation Worker. The calculated risk values for the on-site Installation worker exposed to surface soil are given in Table 10.1A-15. The total HI for this receptor is 0.0256, which is more than an order of magnitude less than the target risk value; therefore, adverse systemic health risks are not expected.

The total ILCR for this receptor is 1.36×10^{-6} . This ILCR value is above the target risk value of 1×10^{-6} . Arsenic is the only carcinogenic risk driver in surface soil.

Juvenile Trespasser. The calculated risk values for the juvenile trespasser exposed to surface soil are given in Table 10.1A-16. The HI for this receptor is 3.84×10^{-3} , which is more than two orders of magnitude below the target value of 1.0; therefore, adverse systemic health risks are not expected for this receptor population.

The total ILCR is 6.86×10^{-8} , which is more than an order of magnitude below the target value of 1×10^{-6} ; therefore, carcinogenic risks are within an acceptable range for this receptor.

10.1A.4.2 Future Land-use Scenarios

Future potential on-site receptors are an Installation worker, a juvenile trespasser, and a resident. Future off-site receptors include an Installation worker, a resident, a juvenile wader, and a sportsman. The potential risks to each of these receptor populations are discussed below.

On-site Installation Worker. The calculated risk values for the on-site Installation worker are given in Table 10.1A-17. The total HI for this receptor is 5.74, which is greater than the target value of 1.0. The major risk drivers are chromium (HI = 5.64) and mercury (HI = 0.0815) in groundwater.

The total ILCR for this receptor is 1.38×10^{-6} . This ILCR value is above the target risk value of 1×10^{-6} . Arsenic in surface soil is the primary carcinogenic risk driver (ILCR = 1.36×10^{-6}).

Juvenile Trespasser. The calculated risk values for the juvenile trespasser are given in Table 10.1A-18. The HI for this receptor is 3.85×10^{-3} , which is more than two orders of magnitude below the target value of 1.0; therefore, adverse systemic health risks are not expected for this receptor population.

The total ILCR is 7.00×10^{-8} , which is more than an order of magnitude below the target value of 1×10^{-6} ; therefore, carcinogenic risks are within an acceptable range for this receptor.

On-site Resident Child. The calculated risk values for the on-site resident child are given in Table 10.1A-19.

The total HI for this receptor is 38.7, which is more than an order of magnitude greater than the target value of 1.0. Chromium in groundwater is the primary risk driver (HI = 37.9), followed by mercury in groundwater (HI = 0.537). Arsenic and chromium in surface soils also contribute significantly to the total risk, with HIs of 0.109 and 0.162, respectively.

The total ILCR for this receptor is 1.17×10^{-5} . This ILCR value is more than an order of magnitude above the target risk value of 1×10^{-6} . Arsenic is the primary carcinogenic risk driver (ILCR = 1.17×10^{-5}).

On-site Resident Adult. The calculated risk values for the on-site resident adult are given in Table 10.1A-20. Only ILCRs were calculated for this receptor population. The noncarcinogenic risks were calculated for the resident child.

The total ILCR for this receptor is 2.31×10^{-6} . This ILCR value is above the target risk value of 1×10^{-6} . Arsenic in surface soil is the primary carcinogenic risk driver (ILCR = 2.29×10^{-6}).

Off-site Installation Worker. The calculated risk values for the off-site Installation worker are given in Table 10.1A-21. The total HI for this receptor is 5.72, which is greater than the target value of 1.0. This risk value is the result of exposure to chromium (HI = 5.64) in groundwater.

The total ILCR for this receptor is 1.91×10^{-8} , which is below the target risk level of 1.0×10^{-6} ; therefore, the ILCR for this receptor is within an acceptable range.

Off-site Resident Child. The calculated risk values for the on-site resident child are given in Table 10.1A-22.

The total HI for this receptor is 38.4, which is more than an order of magnitude greater than the target value of 1.0. Chromium in groundwater is the primary risk driver (HI = 37.9), followed by mercury in groundwater (HI = 0.537).

The total ILCR for this receptor is 1.85×10^{-9} , which is below the target risk level of 1.0×10^{-6} ; therefore, the ILCR for this receptor is within the acceptable range.

Off-site Resident Adult. The calculated risk values for the off-site resident adult are given in Table 10.1A-23. Only ILCRs were calculated for this receptor population. The noncarcinogenic risks were calculated for the resident child.

The total ILCR for this receptor is 2.36×10^{-8} , which is more than an order of magnitude below the target value of 1×10^{-6} ; therefore, the cancer risk for this receptor is within an acceptable range.

Off-site Juvenile Wader. The calculated risk values for the off-site juvenile wader are given in Table 10.1A-24. Mercury was the only COPC identified for this receptor. Mercury does not have a cancer slope factor; therefore, only the noncarcinogenic risks could be calculated for this receptor.

The HI for this receptor is 6.38×10^{-4} , which is more than three orders of magnitude less than the target value of 1.0; therefore, adverse systemic health risks are not expected for this receptor population.

Off-site Sportsman. The calculated risk values for the off-site sportsman are given in Table 10.1A-25. Mercury was the only COPC identified for this receptor. Mercury does not have a cancer slope factor; therefore, only the noncarcinogenic risks could be calculated for this receptor.

The HI for this receptor is 36.5, which is above the target risk value of 1.0. This exceedance is the result of exposure to mercury bioaccumulated in fish tissue (HI = 36.5).

10.1A.5 UNCERTAINTY ASSESSMENT

A discussion of the general uncertainties associated with the analysis of risks at sites within the 16 SWMUs is provided in Appendix I, Section I.5.

To be conservative, it was assumed that the surficial aquifer that was sampled would be used as a source of drinking water. However, the surficial aquifer is not considered to be a source of potable water, and drinking water wells at the FSMR are likely to use the deeper Floridan aquifer.

For the purposes of estimating the surface water concentrations, it was assumed that groundwater discharging into surface water was not diluted. This is a very conservative assumption, given that the groundwater discharging into Canoochee Creek is likely to be diluted by more than an order of magnitude.

The exact chemical forms of chromium and mercury were not known. As a conservative measure, it was assumed that chromium existed in the more toxic hexavalent state, although this form of chromium is very unstable and readily oxidizes to the less toxic trivalent state. None of the mercuric compounds had reference doses calculated for both oral and inhalation pathways; therefore, the oral reference dose for mercuric chloride and the inhalation reference dose for inorganic mercury were used in the risk assessment.

10.1A.6 RISK SUMMARY

The purpose of the risk summary is to provide an overview of the risk assessment results, including identification of the COPCs assessed, the receptor populations, and the risk characterization results.

The COPCs addressed in the baseline risk assessment included HHCOPCs (arsenic and chromium) and CMCOPCs (alpha-BHC, delta-BHC, arsenic, chromium, and mercury). After further analysis of the potential for the CMCOPCs to migrate to groundwater (see Section 10.1A.2.3, Estimation of Exposure Concentrations), it was concluded that alpha-BHC, delta-BHC, and arsenic are not likely to leach into groundwater at concentrations that present an unacceptable risk to human health. The potential risks associated with these constituents as CMCOPCs were not quantified.

The baseline risk assessment addressed the risks associated with exposure to the following COPCs: arsenic (surface soil), chromium (surface soil and groundwater), and mercury (groundwater and surface water). The potential risks associated with exposure to COPCs in groundwater were based on the estimated groundwater concentrations derived from the leachate modeling. The surface water concentration for mercury was estimated based on the groundwater migration model. It was assumed that the concentration of mercury in surface water is equal to the modeled concentration in groundwater (0.00045 mg/L) adjacent to the surface water body; therefore, it was assumed that COPCs in groundwater do not get diluted in Canoochee Creek.

Given the absence of potential migration pathways resulting in exposure of a receptor population, current land-use receptor populations are limited to on-site receptors. The receptor populations assessed for current land use include an Installation worker and a juvenile trespasser. The hunter is also a potential current land-use receptor, but it was determined that exposure via bioaccumulation into venison is not likely to be a significant pathway, so this receptor was not assessed. Future land-use populations include an Installation worker, an on-site trespasser, an off-site juvenile wader, and off-site sportsman fishing in Canoochee Creek. The Installation worker and resident adult represent both on-site and off-site receptors. The resident population was divided into a child and an adult resident because the differences in behavior, exposure duration, and physiology between an adult and a child result in different intakes. The child is more sensitive to noncarcinogenic toxicity because this receptor has a higher intake relative to body weight. Although the resident adult is often more sensitive to carcinogenic effects, given the receptor's longer exposure duration, the resident child has significantly higher intake rates for certain exposure pathways that offset the adult's longer exposure duration, resulting in higher carcinogenic risks for the resident child. Therefore, the resident adult was evaluated for only carcinogenic risks, but the resident child was evaluated for both noncarcinogenic and carcinogenic risks. The reader is referred to Appendix I, Section I.2.2 (Identification of Potential Receptor Populations and Associated Exposure Pathways) for a more detailed discussion on the potential exposure pathways and the differences between the exposures of adult and child resident receptors.

All of the identified COPCs are considered to be COCs based on the results of the risk characterization. The risk values for chromium and mercury in groundwater exceeded the systemic target risk value for all of the potentially exposed receptors. The risk values for arsenic in soils exceeded target risk values for the current and future on-site Installation worker, the on-site resident child, and the on-site resident adult. The risk values for chromium in surface soils exceeded the target risk values for the on-site resident child. Mercury migrating to surface water exceeded the systemic target risk value for the sportsman.

Remedial levels in surface soils will be derived for arsenic and chromium based on direct exposure of a receptor population. Remedial levels in soils will be derived for chromium and mercury based on the potential of these constituents to leach into groundwater and for mercury to subsequently migrate to surface water.

10.1A.7 REMEDIAL LEVELS

Remedial levels are derived for each HHCOC and CMCOC for all applicable environmental media. The first step in determining the remedial levels for a site is to derive remedial levels for each HHCOC and CMCOC based on regulatory and risk-based criteria. These remedial levels are reviewed and a final remedial level for each COC is recommended.

10.1A.7.1 Derivation of Remedial Levels

Remedial levels were derived for the HHCOCs in surface soil (arsenic and chromium). Remedial levels for CMCOCs were derived based on the protection of groundwater (chromium and mercury) and migration to surface water (mercury). The development of remedial levels followed the protocols given in Appendix I, Section I.6.

Risk-based remedial values were derived for the most sensitive receptor population. By protecting the most sensitive receptors, other less sensitive receptor populations will also be protected. If the most sensitive receptor population was not well defined, then remedial levels were derived for those populations that were considered to be representative of the sensitive receptors.

The on-site resident was the most sensitive receptor population for surface soil. The resident child was used to estimate the potential systemic risk associated with exposure to COPCs in surface soils. The resident adult was used to estimate the potential ILCR for an on-site resident population. Therefore, the resident child and the resident adult were used to estimate the risk-based remedial levels based on the systemic risk and the carcinogenic risk, respectively. The remedial levels are given in Table 10.1A-26.

Remedial levels for arsenic were calculated for the resident child. The remedial levels for direct exposure to arsenic in surface soils were calculated based on both the HI and the ILCR, given that the risk values for both noncarcinogenic and carcinogenic risks exceeded the target risk value. The remedial levels for direct exposure to chromium in surface soils were calculated based on exposure of a resident child. The ILCR associated with exposure to chromium in surface soil was below the target risk value for all of the identified receptor populations; therefore, the child resident represents the most sensitive receptor population, and the risk-based remedial levels were derived based on systemic risk only.

The remedial level for a CMCOC represents the soil concentration that is not likely to leach into groundwater and/or subsequently migrate to surface waters in concentrations that present a significant threat to human health. The potential risk associated with CMCOCs is not direct exposure to soils, but exposure to these constituents in either groundwater or surface water; therefore, the soil remedial levels are based upon target groundwater concentrations. These values are the concentrations of a CMCOC in either groundwater or surface water that present a defined risk to a receptor. For example, if the target groundwater concentration is based on an HI of 1.0, the risk value of 1.0 represents the potential risk to a receptor population exposed to the risk-based target concentration of the CMCOC in groundwater. The corresponding risk-based soil remedial value would represent the concentration of the CMCOC in soil that is likely to leach into groundwater, resulting in a CMCOC groundwater concentration equal to the target groundwater concentration.

The most sensitive receptor population for CMCOCs in groundwater is the on-site resident. The on-site resident adult scenario was used to calculate risk-based remedial levels based on carcinogenic risks. The on-site resident child was used to calculate the risk-based remedial levels based on noncarcinogenic risk. Some CMCOCs have both a cancer slope factor and a reference dose; therefore, the target groundwater and surface water concentrations were calculated for both carcinogenic and noncarcinogenic risks.

The CMCOC mercury is a risk driver for the off-site sportsman as a result of its migration to surface water (Canoochee Creek). The target groundwater concentration for this receptor represents the concentration of the CMCOC in groundwater underlying the site that, upon migration and discharge into Canoochee Creek, presents a defined level of risk to the sportsman.

The target groundwater concentrations are given in Table 10.1A-27.

10.1A.7.2 Remedial Level Recommendations

The selection of a remedial level must take into consideration the following factors:

- regulatory standards,
- target risk values for risk-based remedial levels, and
- background concentrations of inorganic COCs.

Regulatory standards that are considered for remedial levels must be derived based on the potential risk to receptors. If regulatory standards are not used for the recommended remedial level, then risk-based remedial values will be recommended based on a target risk value for the receptor population. Finally, the background concentrations of inorganic COCs must be taken into consideration because the remedial actions cannot reduce the concentration of a constituent to levels below the background concentrations.

10.1A.7.2.1 Regulatory Standards

There are no regulatory standards that are directly applicable for the COCs addressed at this site. However, the MCLs should be taken into consideration when selecting remedial levels for CMCOCs based on the protection of groundwater.

The MCLs are appropriate for use as the remedial levels for groundwater HHCOCs. The MCL takes into consideration both the potential human health risks associated with exposure to the contaminant in drinking water and the technological limitations in the removal of that contaminant from water. Therefore, these values are appropriate for use as remedial levels, given that if a risk-based concentration cannot be obtained as a result of technological limitations, the lowest attainable concentration is used as the target concentration.

The selection of a target groundwater concentration for a CMCOC based on an on-site resident must take into consideration the MCL, if available, and the potential risks associated with the presence of all CMCOCs and groundwater HHCOCs. The risk-based target groundwater concentration will be estimated for the appropriate target risk values. However, if the recommended risk-based target groundwater concentration exceeds the MCL, the final recommended value will default to the MCL. An MCL that is based on the acceptable human health risks as defined in the SDWA may be more stringent than the possible target risk values allowed under the current GEPD RCRA guidance (GEPD 1996); therefore, the recommended target groundwater concentration will not exceed the MCL, if available.

10.1A.7.2.2 Target Risk Values for Risk-based Remedial Levels

The selection of a target risk value for remedial levels must take into account the total risk for that receptor population from all of the potential COCs present at the site. The total potential risks associated with the COCs should not result in a cumulative HI that exceeds 3.0 or an ILCR of greater than 1×10^{-4} .

(GEPD 1996). The recommended target risk values for the derivation of the risk-based remedial levels for the on-site resident and the off-site sportsman are discussed below.

On-site Resident. The on-site resident may be exposed to arsenic and chromium in surface soil as well as to the CMCOs chromium and mercury. Arsenic is the only carcinogenic risk driver identified for this site. It is recommended that the risk-based remedial values for carcinogens be based upon an ILCR of 1×10^{-5} , which is below the maximum total acceptable ILCR of 1×10^{-4} .

There are two surface soil constituents and two groundwater constituents that contribute to the total HI for the on-site resident; therefore, the risk-based remedial values should be based on an HI of 0.5, resulting in a total HI of 2.0, which is below the maximum acceptable total HI of 3.0.

Off-site Sportsman. The off-site sportsman may be exposed to mercury migrating into Canoochee Creek. Given that mercury is the only COC identified for this receptor, an HI of 3.0 is recommended as the target risk value for this receptor. This risk value is equal to the maximum target value and takes into consideration that dilution of mercury discharging into the creek is not used in deriving the remedial level. Therefore, the actual potential risk to this receptor would be less than 3.0.

10.1A.7.2.3 Recommended Remedial Levels for the Constituents of Concern

The remedial levels for the HHCOs and CMCOs are given in Tables 10.1A-26 and 10.1A-28, respectively. The selection of the recommended remedial level takes into consideration the MCLs, risk-based remedial levels, and reference background concentrations of inorganics. The recommended remedial level for each COC is discussed in the following paragraphs.

Arsenic. The risk-based remedial levels for this HHCO were calculated for both carcinogenic and noncarcinogenic risks. Arsenic is the only HHCO that contributed significantly to the carcinogenic risks at this site. A risk-based remedial level based on a HI of 0.5 (11.68 mg/kg) is recommended. This value takes into account the potential contribution of other HHCOs to the noncarcinogenic risks associated with the site, and the ILCR associated with this remedial level would be below the target ILCR of 1×10^{-4} (Table 10.1A-26).

The recommended risk-based remedial level for arsenic (11.68 mg/kg) is more than the maximum detected value of 3.4 mg/kg; therefore, no further investigation is recommended to address this HHCO.

Chromium. Chromium was identified as both an HHCO and a CMCO. All of the risk-based remedial levels for direct exposure to chromium in surface soils are less than the reference background concentration. Similarly, all of the risk-based remedial levels based on leaching to groundwater are less than the reference background concentration. Therefore, the reference surface soil background criterion (6.21 mg/kg) is recommended as the remedial level for chromium in soil.

Mercury. Mercury was identified as a CMCO for both groundwater and surface water. The remedial levels for protection of groundwater would be based upon the MCL, given that this value is less than the remedial level based on the recommended HI of 0.5. However, the risk-based remedial level for the protection of surface water (0.003 mg/kg) is less than the remedial level based on the protection of groundwater, 0.13 mg/kg based on the MCL for mercury (Table 10.1A-28).

The risk-based remedial level of 0.003 mg/kg is less than the subsurface soil background concentration (0.05 mg/kg); therefore, the recommended remedial value for mercury is 0.05 mg/kg.

10.1A.7.2.4 Summary of Recommended Remedial Levels

The recommended remedial levels for HHCOCs and CMCOCs are given in Tables 10.1A-26 and 10.12A-28, respectively. The following remedial levels are recommended:

- chromium in surface soil: 6.21 mg/kg (surface soil background),
- chromium in subsurface soil: 11.60 mg/kg (subsurface soil background), and
- mercury in soil: 0.05 mg/kg (subsurface soil background).

The maximum concentration of arsenic in surface soil was less than the recommended remedial level; therefore, no further investigations are required for this constituent.

Table 10.1A-1. Selected Exposure and Modeled Concentrations, SWMU 2

| HHCOPCs | | | |
|---------|----------|--|-----------------------------------|
| Medium | Chemical | Maximum Detected Concentration (mg/kg) | 95 Percent Upper Confidence Limit |
| Soil | Arsenic | 3.4 | 2.54 |
| | Chromium | 47.5 | 35 |

| CMCOPCs | | | |
|---------|-----------|--|---|
| Medium | Chemical | Maximum Detected Concentration (mg/kg) | Predicted SESOIL C(gw,max) at the Source (mg/L) |
| Soil | alpha-BHC | 0.00056 | E ^a |
| | Arsenic | 3.4 | E ^b |
| | Chromium | 11.8 | 1.73 |
| | delta-BHC | 0.0016 | E ^c |
| | Mercury | 0.23 | 0.0025 |

E^a = Alpha-BHC was eliminated as a CMCOPC because the maximum concentration was detected below the water table and would not be a source of alpha-BHC through leaching from soil above the water table. Detections above the water table did not exceed the GSSL.

E^b = Arsenic was eliminated as a CMCOPC because the maximum concentration of arsenic in surface soil (3.4 mg/kg) was below the subsurface soil reference background criterion (8.04 mg/kg). The potential contribution of arsenic from surface soil is not likely to be significant.

E^c = Delta-BHC was eliminated as a CMCOPC because modeling results indicated that delta-BHC naturally attenuates before reaching the water table.

Bold indicates exposure concentration selected.

Table 10.1A-2. Exposure Parameters for Potential Receptor Populations, SWMU 2

| Parameter | Units | On-site Installation Worker | Trespasser | On-site Resident Adult | On-site Resident Child | Off-site Installation Worker | Off-site Resident Adult | Off-site Resident Child | Sportsman | Off-site Juvenile Wader |
|---------------------------------|------------------------|-----------------------------------|------------|------------------------------|------------------------------|------------------------------------|-------------------------------|-------------------------------|-----------|-------------------------------|
| SURFACE SOIL | | | | | | | | | | |
| Incidental Ingestion | | | | | | | | | | |
| Soil ingestion rate | g/day | 0.1 | 0.1 | 0.1 | 0.2 | NA | NA | NA | NA | NA |
| Fraction ingested from area | unitless | 1 | 0.38 | 1 | 1 | NA | NA | NA | NA | NA |
| Exposure frequency | days/year | 250 | 52 | 350 | 350 | NA | NA | NA | NA | NA |
| Exposure duration | years | 25 | 10 | 30 | 6 | NA | NA | NA | NA | NA |
| Body weight | kg | 70 | 45 | 70 | 15 | NA | NA | NA | NA | NA |
| Carcinogen averaging time | days | 25,550 | 25,550 | 25,550 | NA | NA | NA | NA | NA | NA |
| Noncarcinogen averaging time | days | 9,125 | 3,650 | NA | 2,190 | NA | NA | NA | NA | NA |
| Dermal Contact | | | | | | | | | | |
| Skin area | cm ² /event | 5,000 | 3,700 | 5,000 | 1,700 | NA | NA | NA | NA | NA |
| Adherence factor | mg/cm ² | 0.2 | 0.2 | 0.2 | 0.2 | NA | NA | NA | NA | NA |
| Exposure frequency | events/year | 250 | 52 | 350 | 350 | NA | NA | NA | NA | NA |
| Exposure duration | years | 25 | 10 | 30 | 6 | NA | NA | NA | NA | NA |
| Body weight | kg | 70 | 45 | 70 | 15 | NA | NA | NA | NA | NA |
| Carcinogen averaging time | days | 25,550 | 25,550 | 25,550 | NA | NA | NA | NA | NA | NA |
| Noncarcinogen averaging time | days | 9,125 | 3,650 | NA | 2,190 | NA | NA | NA | NA | NA |
| Inhalation of Dust | | | | | | | | | | |
| Inhalation rate | m ³ /hr | 2.5 | 1.9 | 0.80 | 0.68 | 2.5 | 0.80 | 0.68 | NA | NA |
| Exposure time | hr/day | 8 | 6 | 18.4 | 18.4 | 8 | 18.4 | 18.4 | NA | NA |
| Exposure frequency | days/year | 250 | 52 | 350 | 350 | 250 | 350 | 350 | NA | NA |
| Exposure duration | years | 25 | 10 | 30 | 6 | 25 | 30 | 6 | NA | NA |
| Body weight | kg | 70 | 45 | 70 | 15 | 70 | 70 | 15 | NA | NA |
| Carcinogen averaging time | days | 25,550 | 25,550 | 25,550 | NA | 25,550 | 25,550 | NA | NA | NA |
| Noncarcinogen averaging time | days | 9,125 | 3,650 | NA | 2,190 | 9,125 | NA | 2,190 | NA | NA |
| GROUNDWATER | | | | | | | | | | |
| Drinking Water Ingestion | | | | | | | | | | |
| Drinking water ingestion | L/day | 1 | NA | 2 | 1 | 1 | 2 | 1 | NA | NA |
| Fraction ingested from area | unitless | 1 | NA | 1 | 1 | 1 | 1 | 1 | NA | NA |
| Exposure frequency | days/year | 250 | NA | 350 | 350 | 250 | 350 | 350 | NA | NA |
| Exposure duration | years | 25 | NA | 30 | 6 | 25 | 30 | 6 | NA | NA |
| Body weight | kg | 70 | NA | 70 | 15 | 70 | 70 | 15 | NA | NA |
| Carcinogen averaging time | days | 25,550 | NA | 25,550 | NA | 25,550 | 25,550 | NA | NA | NA |
| Noncarcinogen averaging time | days | 9,125 | NA | NA | 2,190 | 9,125 | NA | 2,190 | NA | NA |

Note: Footnotes appear on page 10.1A-17.

Table 10.1A-2. Exposure Parameters for Potential Receptor Populations, SWMU 2 (continued)

| Parameter | Units | On-site Installation Worker | Trespasser | On-site Resident Adult | On-site Resident Child | Off-site Installation Worker | Off-site Resident Adult | Off-site Resident Child | Sportsman | Off-site Juvenile Wader |
|-------------------------------------|----------------|-----------------------------------|------------|------------------------------|------------------------------|------------------------------------|-------------------------------|-------------------------------|-----------|-------------------------------|
| Dermal Contact while Bathing | | | | | | | | | | |
| Skin area | m ² | NA | NA | NA | 0.170 | NA | NA | 0.170 | NA | NA |
| Exposure time | hours/day | NA | NA | NA | 0.33 | NA | NA | 0.33 | NA | NA |
| Exposure frequency | days/year | NA | NA | NA | 350 | NA | NA | 350 | NA | NA |
| Exposure duration | years | NA | NA | NA | 6 | NA | NA | 6 | NA | NA |
| Body weight | kg | NA | NA | NA | 15 | NA | NA | 15 | NA | NA |
| Carcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Noncarcinogen averaging time | days | NA | NA | NA | 2,190 | NA | NA | 2,190 | NA | NA |
| SURFACE WATER | | | | | | | | | | |
| Incidental Ingestion | | | | | | | | | | |
| Water ingestion rate | L/hour | NA | NA | NA | NA | NA | NA | NA | 0.01 | 0.05 |
| Exposure time | hours/day | NA | NA | NA | NA | NA | NA | NA | 4 | 2 |
| Exposure frequency | days/year | NA | NA | NA | NA | NA | NA | NA | 52 | 52 |
| Exposure duration | years | NA | NA | NA | NA | NA | NA | NA | 30 | 10 |
| Body weight | kg | NA | NA | NA | NA | NA | NA | NA | 70 | 45 |
| Carcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 25,550 | 25,550 |
| Noncarcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 10,950 | 3,650 |
| Dermal Contact while Wading | | | | | | | | | | |
| Skin area | m ² | NA | NA | NA | NA | NA | NA | NA | 0.41 | 0.4 |
| Exposure time | hours/day | NA | NA | NA | NA | NA | NA | NA | 4 | 2 |
| Exposure frequency | days/year | NA | NA | NA | NA | NA | NA | NA | 52 | 52 |
| Exposure duration | years | NA | NA | NA | NA | NA | NA | NA | 30 | 10 |
| Body weight | kg | NA | NA | NA | NA | NA | NA | NA | 70 | 45 |
| Carcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 25,550 | 25,550 |
| Noncarcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 10,950 | 3,650 |
| BIOTA | | | | | | | | | | |
| Ingestion of Fish | | | | | | | | | | |
| Ingestion Rate | kg/day | NA | NA | NA | NA | NA | NA | NA | 0.027 | NA |
| Fraction ingested from area | unitless | NA | NA | NA | NA | NA | NA | NA | 1 | NA |
| Exposure frequency | days/year | NA | NA | NA | NA | NA | NA | NA | 365 | NA |
| Exposure duration | years | NA | NA | NA | NA | NA | NA | NA | 30 | NA |
| Body weight | kg | NA | NA | NA | NA | NA | NA | NA | 70 | NA |
| Carcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 25,550 | NA |
| Noncarcinogen averaging time | days | NA | NA | NA | NA | NA | NA | NA | 10,950 | NA |

NA = Not applicable; this receptor is not exposed via this pathway.

Table 10.1A-3. Estimated Intakes for Current Installation Worker, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Dermal Exposure ^a | |
|----------------------|----------|------------------------|-------|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | 2.49E-06 | 8.88E-07 | 2.49E-08 | 8.88E-09 |
| | Chromium | 35 | mg/kg | 3.42E-05 | 1.22E-05 | 3.42E-07 | 1.22E-07 |

^aThe equations used to calculate oral and dermal exposures for surface soil are presented in Appendix I, Section 1.2.4.2.

Table 10.1A-4. Estimated Intakes for Current Juvenile Trespasser, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Dermal Exposure ^a | |
|----------------------|----------|------------------------|-------|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | 3.06E-07 | 4.37E-08 | 5.95E-09 | 8.50E-10 |
| | Chromium | 35 | mg/kg | 4.21E-06 | 6.02E-07 | 8.20E-08 | 1.17E-08 |

^aThe equations used to calculate oral and dermal exposures for surface soil are presented in Appendix I, Section 1.2.4.2.

Table 10.1A-5. Estimated Intakes for Future On-site Installation Worker, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^{a,b} | | Dermal Exposure ^a | | Inhalation Exposure ^a | |
|----------------------|----------|------------------------|-------|---|--|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | 2.49E-06 | 8.88E-07 | 2.49E-08 | 8.88E-09 | 9.23E-11 | 3.30E-11 |
| | Chromium | 35 | mg/kg | 3.42E-05 | 1.22E-05 | 3.42E-07 | 1.22E-07 | 1.27E-09 | 4.54E-10 |
| Groundwater | Chromium | 1.73 | mg/L | 1.69E-02 | 6.05E-03 | NA ^c | NA ^c | NA ^d | NA ^d |
| | Mercury | 0.0025 | mg/L | 2.45E-05 | 8.74E-06 | NA ^c | NA ^c | NA ^d | NA ^d |

^aThe equations used to calculate oral, dermal, and inhalation exposures for soil are presented in Appendix I, Section I.2.4.2.^bThe equation used to calculate oral exposure for groundwater is presented in Appendix I, Section I.2.4.3.^cNA = Not applicable; this pathway was not assessed for this receptor.^dNA = Not applicable; this pathway was not assessed for this chemical.

Table 10.1A-6. Estimated Intakes for Future On-site Juvenile Trespasser, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Dermal Exposure ^a | | Inhalation Exposure ^a | |
|----------------------|----------|------------------------|-------|---|--|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | 3.06E-07 | 4.37E-08 | 5.95E-09 | 8.50E-10 | 1.70E-11 | 2.43E-12 |
| | Chromium | 35 | mg/kg | 4.21E-06 | 6.02E-07 | 8.20E-08 | 1.17E-08 | 2.35E-10 | 3.35E-11 |

^aThe equations used to calculate oral, dermal, and inhalation exposures for soil are presented in Appendix I, Section I.2.4.2.

Table 10.1A-7. Estimated Intakes for Future On-site Resident Child, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^{a,b} | | Dermal Exposure ^{a,b} | | Inhalation Exposure ^a | |
|----------------------|----------|------------------------|-------|---|--|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | 3.25E-05 | 7.77E-06 | 5.52E-08 | 1.32E-08 | 3.77E-10 | 9.03E-11 |
| Groundwater | Chromium | 35 | mg/kg | 4.47E-04 | NA ^c | 7.61E-07 | NA ^c | 5.20E-09 | 4.46E-10 |
| | Chromium | 1.73 | mg/L | 1.1E-01 | NA ^c | 6.2E-05 | NA ^c | NA ^d | NA |
| | Mercury | 0.0025 | mg/L | 1.6E-04 | NA ^c | 8.97E-08 | NA ^c | NA ^d | NA |

^aThe equations used to calculate oral, dermal, and inhalation exposures for soil are presented in Appendix I, Section 1.2.4.2.

^bThe equations used to calculate oral and dermal exposures for groundwater are presented in Appendix I, Section 1.2.4.3.

^cNA = Not applicable; applicable toxicity data were not available.

^dNA = Not applicable; this pathway was not assessed for this chemical.

Table 10.1A-8. Estimated Intakes for Future On-site Resident Adult, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Dermal Exposure ^a | Inhalation Exposure ^a |
|----------------------|----------|------------------------|-------|--|--|------------------------------|----------------------------------|
| | | | | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | | |
| Surface soil | Arsenic | 2.54 | mg/kg | 1.49E-06 | 1.49E-08 | 4.08E-11 | 5.62E-10 |
| Groundwater | Chromium | 35 | mg/kg | NA | NA | NA | NA |
| | Mercury | 0.00356 | mg/L | NA | NA | NA | NA |

^aThe equations used to calculate oral, dermal, and inhalation exposures for soil are presented in Appendix I, Section 1.2.4.2.

NA = Not applicable; applicable toxicity data were not available.

Table 10.1A-9. Estimated Intakes for Future Off-site Installation Worker, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Inhalation Exposure ^b | |
|----------------------|----------|------------------------|-------|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | NA ^c | NA ^c | 9.23E-11 | 3.30E-11 |
| | Chromium | 35 | mg/kg | NA ^c | NA ^c | 1.27E-09 | 4.54E-10 |
| Groundwater | Chromium | 1.73 | mg/L | 1.69E-02 | 6.05E-03 | NA ^d | NA ^d |
| | Mercury | 0.0025 | mg/L | 2.45E-05 | 8.74E-06 | NA ^d | NA ^d |

^aThe equation used to calculate oral exposure for groundwater is presented in Appendix I, Section 1.2.4.3.

^bThe equation used to calculate inhalation exposure for soil is presented in Appendix I, Section 1.2.4.2.

^cNA = Not applicable; this pathway was not assessed for this receptor.

^dNA = Not applicable; this pathway was not assessed for this chemical.

Table 10.1A-10. Estimated Intakes for Future Off-site Resident Child, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | | Dermal Exposure ^a | | Inhalation Exposure ^b | |
|----------------------|----------|------------------------|-------|---|--|---|--|---|--|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | NA ^c | NA ^c | NA ^c | NA ^c | 3.77E-10 | 9.03E-11 |
| | Chromium | 35 | mg/kg | NA ^c | NA ^c | NA ^c | NA ^c | 5.20E-09 | 4.46E-10 |
| Groundwater | Chromium | 1.73 | mg/L | 1.11E-01 | NA ^d | 6.2E-05 | NA ^d | NA ^e | NA ^e |
| | Mercury | 0.0025 | mg/L | 1.60E-04 | NA ^d | 8.97E-08 | NA ^d | NA ^e | NA ^e |

^aThe equations used to calculate oral and dermal exposures for groundwater are presented in Appendix I, Section 1.2.4.3.

^bThe equation used to calculate inhalation exposure for soil is presented in Appendix I, Section 1.2.4.2.

^cNA = Not applicable; this pathway was not assessed for this receptor.

^dNA = Not applicable; applicable toxicity data were not available.

^eNA = Not applicable; this pathway was not assessed for this chemical.

Table 10.1A-11. Estimated Intakes for Future Off-site Resident Adult, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | Dermal Exposure ^b | Inhalation Exposure ^c |
|----------------------|----------|------------------------|-------|--|--|--|
| | | | | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) | Average Daily Dose for Carcinogens (mg/kg/d) |
| Surface soil | Arsenic | 2.54 | mg/kg | NA ^c | NA ^c | 4.08E-11 |
| | Chromium | 35 | mg/kg | NA ^c | NA ^c | 5.62E-10 |
| Groundwater | Chromium | 1.73 | mg/L | NA ^d | NA ^d | NA ^e |
| | Mercury | 0.0025 | mg/L | NA ^d | NA ^d | NA ^e |

^aThe equations used to calculate oral, dermal, and inhalation exposures for soil are presented in Appendix I, Section I.2.4.2.

^bThe equations used to calculate oral, dermal, and inhalation exposures for groundwater are presented in Appendix I, Section I.2.4.3.

^cNA = Not applicable; this pathway was not assessed for this receptor.

^dNA = Not applicable; this pathway was not assessed for this chemical.

^eNA = Not applicable; applicable toxicity data were not available.

Table 10.1A-12. Estimated Intakes for the Future Off-site Juvenile Wader, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^a | Dermal Exposure ^a |
|----------------------|----------|------------------------|-------|---|---|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) |
| Surface water | Mercury | 4.50E-04 | mg/L | 2.85E-08 | 1.14E-08 |

^aThe equations used to calculate oral and dermal exposures for surface water are presented in Appendix I, Section I.2.4.4.

Table 10.1A-13. Estimated Intakes for the Future Off-site Sportsman, SWMU 2

| Environmental Medium | Chemical | Exposure Concentration | Units | Oral Exposure ^{a,b} | Dermal Exposure ^a |
|----------------------|----------|------------------------|-------|---|---|
| | | | | Average Daily Dose for Noncarcinogens (mg/kg/d) | Average Daily Dose for Noncarcinogens (mg/kg/d) |
| Surface water | Mercury | 4.50E-04 | mg/L | 3.66E-08 | 1.50E-08 |
| Fish tissue | Mercury | 2.84E+01 | mg/kg | 1.09E-02 | NA |

^aThe equations used to calculate oral and dermal exposures for surface water are presented in Appendix I, Section I.2.4.4.

^bThe equation used to calculate exposure for fish uptake is presented in Appendix I, Section I.2.4.6.

NA = Not applicable; this is not a viable exposure pathway for this environmental medium.

Table 10.1A-14. Toxicity Values for Constituents of Potential Concern, SWMU 2

| Chemical | Oral Reference Dose (mg/kg/d) | Ref. ^a | Oral Cancer Slope Factor 1/(mg/kg/d) | Ref. ^a | Gastrointestinal Absorption Factor | Dermal Reference Dose ^b (mg/kg/d) | Dermal Cancer Slope Factor ^c 1/(mg/kg/d) | Inhalation Reference Dose (mg/kg/d) | Ref. ^a | Inhalation Cancer Slope Factor 1/(mg/kg/d) | Ref. ^a |
|-----------------------|--|-------------------|---|-------------------|--|---|--|--|-------------------|---|-------------------|
| Arsenic | 3.00E-04 | I | 1.50E+00 | I | 4.10E-01 | 1.23E-04 | 3.66E+00 | ND | | 1.51E+01 | I |
| Chromium ^d | 3.00E-03 | I | ND | | 2.00E-02 | 6.00E-05 | ND | 2.86E-05 | I | 4.10E+01 | H |
| Mercury ^e | 3.00E-04 | I | ND | | 7.00E-02 | 2.10E-05 | ND | 8.60E-05 | I | ND | |

^aReferences: I = IRIS (EPA 1999a); H = HEAST (EPA 1997c).

^bDermal reference dose calculated using the following formula: oral reference dose × gastrointestinal absorption factor.

^cDermal cancer slope factor calculated using the following formula: oral reference dose ÷ gastrointestinal absorption factor.

^dToxicity values given are for hexavalent chromium.

^eOral and dermal reference dose value are for mercuric chloride.

ND = Toxicological values were not available to assess this route of exposure.

**Table 10.1A-15. Hazard Indices and Carcinogenic Risks for
Current On-site Installation Worker, SWMU 2**

| Chemical | Surface Soil ^a | | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Total | |
| Arsenic | 8.28E-03 | 2.02E-04 | 8.49E-03 | 8.49E-03 |
| Chromium | 1.14E-02 | 5.71E-03 | 1.71E-02 | 1.71E-02 |
| Pathway Total | 1.97E-02 | 5.91E-03 | 2.56E-02 | 2.56E-02 |
| Chemical | Surface Soil ^b | | | Total Cancer Risk ^b |
| | Oral ILCR | Dermal ILCR | Total | |
| Arsenic | 1.33E-06 | 3.25E-08 | 1.36E-06 | 1.36E-06 |
| Chromium | NA | NA | NA | — |
| Pathway Total | 1.33E-06 | 3.25E-08 | 1.36E-06 | 1.36E-06 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; applicable toxicity data were not available.

— = No data.

**Table 10.1A-16. Hazard Indices and Carcinogenic Risks for
Current Juvenile Trespasser, SWMU 2**

| Chemical | Surface Soil ^a | | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Total | |
| Arsenic | 1.02E-03 | 4.84E-05 | 1.07E-03 | 1.07E-03 |
| Chromium | 1.40E-03 | 1.37E-03 | 2.77E-03 | 2.77E-03 |
| Pathway Total | 2.42E-03 | 1.41E-03 | 3.84E-03 | 3.84E-03 |
| Chemical | Surface Soil ^b | | | Total Cancer Risk ^b |
| | Oral ILCR | Dermal ILCR | Total | |
| Arsenic | 6.55E-08 | 3.11E-09 | 6.86E-08 | 6.86E-08 |
| Chromium | NA | NA | NA | — |
| Pathway Total | 6.55E-08 | 3.11E-09 | 6.86E-08 | 6.86E-08 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; applicable toxicity data were not available.

— = No data.

Table 10.1A-17. Hazard Indices and Carcinogenic Risks for Future On-site Installation Worker, SWMU 2

| Chemical | Surface Soil ^a | | | | Groundwater ^a | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|-----------------|-----------------|--------------------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Inhalation HQ | Total | Oral HQ | Total | |
| Arsenic | 8.28E-03 | 2.02E-04 | ND | 8.49E-03 | NA | NA | 8.49E-03 |
| Chromium | 1.14E-02 | 5.71E-03 | 4.45E-05 | 1.72E-02 | 5.64E+00 | 5.64E+00 | 5.66E+00 |
| Mercury | NA | NA | NA | NA | 8.15E-02 | 8.15E-02 | 8.15E-02 |
| Pathway Total | 1.97E-02 | 5.91E-03 | 4.45E-05 | 2.57E-02 | 5.72E+00 | 5.72E+00 | 5.74E+00 |
| Chemical | Surface Soil ^b | | | | Groundwater ^b | | Total Cancer Risk ^b |
| | Oral ILCR | Dermal ILCR | Inhalation ILCR | Total | Oral ILCR | Total | |
| Arsenic | 1.33E-06 | 3.25E-08 | 4.96E-10 | 1.36E-06 | NA | NA | 1.36E-06 |
| Chromium | ND | ND | 1.86E-08 | 1.86E-08 | ND | NA | 1.86E-08 |
| Mercury | NA | NA | NA | NA | ND | NA | NA |
| Pathway Total | 1.33E-06 | 3.25E-08 | 1.91E-08 | 1.38E-06 | — | — | 1.38E-06 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

— = No data.

Table 10.1A-18. Hazard Indices and Carcinogenic Risks for Future On-site Juvenile Trespasser, SWMU 2

| Chemical | Surface Soil ^a | | | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|-----------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Inhalation HQ | Total | |
| Arsenic | 1.02E-03 | 4.84E-05 | ND | 1.07E-03 | 1.07E-03 |
| Chromium | 1.40E-03 | 1.37E-03 | 8.21E-06 | 2.78E-03 | 2.78E-03 |
| Pathway Total | 2.42E-03 | 1.41E-03 | 8.21E-06 | 3.85E-03 | 3.85E-03 |
| Chemical | Surface Soil ^b | | | | Total Cancer Risk ^b |
| | Oral ILCR | Dermal ILCR | Inhalation ILCR | Total | |
| Arsenic | 6.55E-08 | 3.11E-09 | 3.66E-11 | 6.86E-08 | 6.86E-08 |
| Chromium | ND | ND | 1.37E-09 | 1.37E-09 | 1.37E-09 |
| Pathway Total | 6.55E-08 | 3.11E-09 | 1.41E-09 | 7.00E-08 | 7.00E-08 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable.

ND = Toxicological values were not available to assess this route of exposure.

Table 10.1A-19. Hazard Indices and Carcinogenic Risks for Future On-site Resident Child, SWMU 2

| Chemical | Surface Soil ^a | | | | Groundwater ^a | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|-----------------|-----------------|--------------------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Inhalation HQ | Total | Oral HQ | Dermal HQ | |
| Arsenic | 1.08E-01 | 4.49E-04 | ND | 1.09E-01 | NA | NA | 1.09E-01 |
| Chromium | 1.49E-01 | 1.27E-02 | 1.82E-04 | 1.62E-01 | 3.69E+01 | 1.04E+00 | 3.81E+01 |
| Mercury | NA | NA | NA | NA | 5.33E-01 | 4.27E-03 | 5.37E-01 |
| Pathway Total | 2.57E-01 | 1.31E-02 | 1.82E-04 | 2.71E-01 | 3.74E+01 | 1.04E+00 | 3.87E+01 |

| Chemical | Surface Soil ^b | | | | Groundwater ^b | | Total Cancer Risk ^b |
|----------------------|---------------------------|-----------------|-----------------|-----------------|--------------------------|-------------|--------------------------------|
| | Oral ILCR | Dermal ILCR | Inhalation ILCR | Total | Oral ILCR | Dermal ILCR | |
| Arsenic | 1.17E-05 | 4.83E-08 | 1.36E-09 | 1.17E-05 | NA | NA | 1.17E-05 |
| Chromium | ND | ND | 1.83E-08 | 1.83E-08 | ND | ND | 1.83E-08 |
| Mercury | NA | NA | NA | NA | ND | ND | — |
| Pathway Total | 1.17E-05 | 4.83E-08 | 1.96E-08 | 1.17E-05 | — | — | 1.17E-05 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

Table 10.1A-20. Carcinogenic Risks for Future On-site Resident Adult, SWMU 2

| Chemical | Surface Soil ^a | | | | Groundwater ^a | | Total Cancer Index ^a |
|----------------------|---------------------------|-----------------|-----------------|-----------------|--------------------------|-------------|---------------------------------|
| | Oral ILCR | Dermal ILCR | Inhalation ILCR | Total | Oral ILCR | Dermal ILCR | |
| Arsenic | 2.24E-06 | 5.46E-08 | 6.14E-10 | 2.29E-06 | NA | NA | 2.29E-06 |
| Chromium | ND | ND | 2.30E-08 | 2.30E-08 | ND | ND | 2.30E-08 |
| Mercury | NA | NA | NA | NA | ND | ND | — |
| Pathway Total | 2.24E-06 | 5.46E-08 | 2.36E-08 | 2.31E-06 | — | — | 2.31E-06 |

^aThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

— = No data.

**Table 10.1A-21. Hazard Indices and Carcinogenic Risks for
Future Off-site Installation Worker, SWMU 2**

| Chemical | Surface Soil ^a | | Groundwater ^a | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|--------------------------|-----------------|---------------------------------|
| | Inhalation HQ | Total | Oral HQ | Total | |
| Arsenic | ND | ND | NA | NA | NA |
| Chromium | 4.45E-05 | 4.45E-05 | 5.64E+00 | 5.64E+00 | 5.64E+00 |
| Mercury | NA | NA | 8.15E-02 | 8.15E-02 | 8.15E-02 |
| Pathway Total | 4.45E-05 | 4.45E-05 | 5.72E+00 | 5.72E+00 | 5.72E+00 |
| Chemical | Surface Soil ^b | | Groundwater ^b | | Total Cancer Risk ^b |
| | Inhalation ILCR | Total | Oral ILCR | Total | |
| Arsenic | 4.96E-10 | 4.96E-10 | NA | NA | 4.96E-10 |
| Chromium | 1.86E-08 | 1.86E-08 | ND | ND | 1.86E-08 |
| Mercury | NA | NA | ND | ND | NA |
| Pathway Total | 1.91E-08 | 1.91E-08 | — | — | 1.91E-08 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

— = No data.

**Table 10.1A-22. Hazard Indices and Carcinogenic Risks for
Future Off-site Resident Child, SWMU 2**

| Chemical | Surface Soil ^a | | Groundwater ^a | | | Total Hazard Index ^a |
|----------------------|---------------------------|-----------------|--------------------------|-----------------|-----------------|---------------------------------|
| | Inhalation HQ | Total | Oral HQ | Dermal HQ | Total | |
| Arsenic | ND | ND | NA | NA | NA | — |
| Chromium | 1.82E-04 | 1.82E-04 | 3.69E+01 | 1.04E+00 | 3.79E+01 | 3.79E+01 |
| Mercury | NA | NA | 5.33E-01 | 4.27E-03 | 5.37E-01 | 5.37E-01 |
| Pathway Total | 1.82E-04 | 1.82E-04 | 3.74E+01 | 1.04E+00 | 3.84E+01 | 3.84E+01 |
| Chemical | Surface Soil ^b | | Groundwater ^b | | | Total Cancer Risk ^b |
| | Inhalation ILCR | Total | Oral ILCR | Dermal ILCR | Total | |
| Arsenic | 1.36E-09 | 1.36E-09 | NA | NA | NA | 1.36E-09 |
| Chromium | 4.88E-10 | 4.88E-10 | ND | ND | ND | 4.88E-10 |
| Mercury | ND | ND | ND | ND | ND | — |
| Pathway Total | 1.85E-09 | 1.85E-09 | — | — | — | 1.85E-09 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

— = No data.

Table 10.1A-23. Carcinogenic Risks for Future Off-site Resident Adult, SWMU 2

| Chemical | Surface Soil ^a | | Groundwater ^a | | | Total Cancer Risk ^a |
|----------------------|---------------------------|-----------------|--------------------------|-------------|----------|--------------------------------|
| | Inhalation ILCR | Total | Oral ILCR | Dermal ILCR | Total | |
| Arsenic | 6.14E-10 | 6.14E-10 | NA | NA | NA | 6.14E-10 |
| Chromium | 2.30E-08 | 2.30E-08 | ND | ND | ND | 2.30E-08 |
| Mercury | NA | NA | ND | ND | ND | — |
| Pathway Total | 2.36E-08 | 2.36E-08 | — | — | — | 2.36E-08 |

^aThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = Toxicological values were not available to assess this route of exposure.

— = No data.

Table 10.1A-24. Hazard Indices for Future Off-site Juvenile Wader, SWMU 2

| Chemical | Surface Water ^a | | | Total Hazard Index ^a |
|----------------------|----------------------------|-----------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Total | |
| Mercury ^b | 9.50E-05 | 5.43E-04 | 6.38E-04 | 6.38E-04 |
| Pathway Total | 9.50E-05 | 5.43E-04 | 6.38E-04 | 6.38E-04 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThere are no cancer slope factors for mercury; therefore, only noncarcinogenic risk values are presented.

Table 10.1A-25. Hazard Indices for Future Off-site Sportsman, SWMU 2

| Chemical | Surface Water ^a | | | Fish Tissue ^a | | Total Hazard Index ^a |
|----------------------|----------------------------|-----------------|-----------------|--------------------------|-----------------|---------------------------------|
| | Oral HQ | Dermal HQ | Total | Oral HQ | Total | |
| Mercury ^b | 1.22E-04 | 7.15E-04 | 8.37E-04 | 3.65E+01 | 3.65E+01 | 3.65E+01 |
| Pathway Total | 1.22E-04 | 7.15E-04 | 8.37E-04 | 3.65E+01 | 3.65E+01 | 3.65E+01 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThere are no cancer slope factors for mercury; therefore, only noncarcinogenic risk values are presented.

Table 10.1A-26. Remedial Levels for Soil Based on Direct Exposure, SWMU 2

| Constituent of Concern | Maximum Detected Concentration (mg/kg) | Reference Background Concentration Surface Soil (mg/kg) | Risk-based Remedial Levels (mg/kg) | | | |
|------------------------|--|---|------------------------------------|-------|------|----------------------|
| | | | HI | | ILCR | |
| | | | 1 | 0.5 | 0.1 | 1 × 10 ⁻⁶ |
| Arsenic | 3.4 | 2.10 | 23.37 | 11.68 | 2.34 | 0.6 |
| Chromium | 47.5 | 6.21 | 1.53 | 0.77 | 0.15 | NA |
| | | | | | | 1 × 10 ⁻⁵ |
| | | | | | | 60.6 |
| | | | | | | NA |

NA = Not applicable; toxicity data required for calculation of remedial level were not available.
Bold indicates values that are the recommended remedial values.

Table 10.1A-27. Target Groundwater Concentrations for Contaminant Migration Constituents of Concern, SWMU 2

| Constituent of Concern | Point of Exposure ^a | Maximum Modeled Groundwater Concentration (µg/L) | Maximum Contaminant Level (µg/L) | Risk-based Target Groundwater Concentrations (µg/kg) | | | |
|------------------------|--------------------------------|--|----------------------------------|--|-------|-------|------|
| | | | | HI | | | |
| | | | | 3 | 1 | 0.5 | 0.1 |
| Chromium | Groundwater | 1041 | 100.0 | 136.2 | 45.41 | 22.70 | 4.54 |
| Mercury | Groundwater | 3.56 | 2,000 | 14.0 | 4.66 | 2.33 | 0.47 |
| Mercury | Surface water | 3.56 | NA | 0.3 | 0.10 | 0.05 | 0.01 |

^aGroundwater represents groundwater underlying the site, and surface water represents surface water in Canoochee Creek.

NA = Not applicable; MCLs are not applicable to surface water.

Bold indicates values that are the recommended remedial values.

**Table 10.2A-24. Hazard Indices and Carcinogenic Risks for
Future Off-site Resident Child, SWMU 3**

| Chemical | Groundwater ^a | | | Surface Soil ^a | | Total Hazard Index ^a |
|----------------------|--------------------------|-----------------|-----------------|---------------------------|-----------------|---------------------------------------|
| | Oral HQ | Dermal HQ | Total | Inhalation HQ | Total | |
| Arsenic | NA | NA | NA | ND | ND | — |
| delta-BHC | ND | ND | — | NA | NA | — |
| Mercury | 4.16E-02 | 3.33E-04 | 4.19E-02 | NA | NA | 4.19E-02 |
| Pathway Total | 4.16E-02 | 3.33E-04 | 4.19E-02 | — | — | 4.19E-02 |
| Chemical | Groundwater ^b | | | Surface Soil ^b | | Total Cancer Risk ^b |
| | Oral ILCR | Dermal ILCR | Total | Inhalation ILCR | Total | |
| Arsenic | NA | NA | NA | 1.36E-09 | 1.36E-09 | 1.36E-09 |
| delta-BHC | 3.99E-07 | 7.85E-09 | 4.07E-07 | NA | NA | 4.07E-07 |
| Mercury | ND | ND | — | NA | NA | — |
| Pathway Total | 3.99E-07 | 7.85E-09 | 4.07E-07 | 1.36E-09 | 1.36E-09 | 4.08E-07 |

^aThe equations used to calculate noncarcinogenic risk are presented in Appendix I, Section I.4.2.

^bThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = The toxicity data required to quantify the risk are not available.

— = No data.

Table 10.2A-25. Carcinogenic Risks for Future Off-site Resident Adult, SWMU 3

| Chemical | Groundwater ^a | | | Surface Soil ^a | | Total Cancer Risk ^a |
|----------------------|--------------------------|-----------------|-----------------|---------------------------|-----------------|--------------------------------------|
| | Oral ILCR | Dermal ILCR | Total | Inhalation ILCR | Total | |
| Arsenic | NA | NA | NA | 1.71E-09 | 1.71E-09 | 1.71E-09 |
| delta-BHC | 8.56E-07 | 5.00E-08 | 9.06E-07 | NA | NA | 9.06E-07 |
| Mercury | ND | ND | — | NA | NA | — |
| Pathway Total | 8.56E-07 | 5.00E-08 | 9.06E-07 | 1.71E-09 | 1.71E-09 | 9.08E-07 |

^aThe equations used to calculate carcinogenic risk are presented in Appendix I, Section I.4.1.

NA = Not applicable; this chemical is not present in this environmental medium.

ND = The toxicity data required to quantify the risk are not available.

— = No data.

11.0 CONCLUSIONS AND RECOMMENDATIONS

11.1 CONCLUSIONS

Nature and Extent of Contamination

The results of the chemical analyses on surface soil, subsurface soil, and groundwater were screened against the reference background criteria for the FSMR. Surface water and sediment were screened against the site-specific background criteria.

Inorganics were considered to be SRCs if their concentrations were above the reference background concentrations, and organics were considered to be SRCs if they were simply detected because organic constituents are considered to potentially be man-made. The maximum concentration of SRCs was carried through to fate and transport, HHPRE, and EPRE analysis. A summary of SRCs by medium for each SWMU is presented in Table 11-1.

Fate and Transport Analysis

Fate and transport analysis was performed on each SWMU. This analysis included developing a site-specific CSM and determining the potential for SRCs in surface soil, subsurface soil, and/or sediment to migrate to groundwater.

The maximum concentrations of the identified SRCs determined from nature and extent analysis were compared to EPA GSSLs. Generally, if contaminant concentrations in soil and/or sediment fall below their respective GSSLs and there are no significant ecological receptors of concern, then no further study or action is warranted. SRCs were identified as CMCOPCs if they were detected at concentrations that exceeded their respective GSSLs. A summary of the results of the fate and transport analysis (CMCOPCs) is presented in Table 11-2.

A weight-of-evidence approach was used to evaluate each CMCOPC identified based on leaching to groundwater. In some instances, the potential impact of CMCOPCs to groundwater, and possibly to surface water, was evaluated (modeled concentrations were compared to risk-based criteria) in a human health baseline risk assessment (see following section). CMCOPCs that indicated a potential risk to human health (i.e., that exceeded risk-based screening criteria) were identified as CMCOs, and remedial levels were developed based on protection of groundwater. SWMUs for which a human health baseline risk assessment was performed are identified in Table 11-2.

Human Health Preliminary Risk Evaluation

An HHPRE using a Step i risk evaluation approach based on guidance from GEPD was performed on each SWMU to determine the potential human health risks associated with the maximum concentrations of the identified SRCs. A summary of the HHPRE results (HHCOPCs) is presented in Table 11-2.

A weight-of-evidence approach was used to evaluate each HHCOPC identified in the preliminary risk assessment. In some instances, HHCOPCs were evaluated further in a human health baseline risk assessment. HHCOPCs and/or CMCOPCs (see previous section) that either had HIs of 0.1 or ILCRs of 1×10^{-6} were identified as HHCOs. Remedial levels were developed that were protective of the most sensitive receptor population, based on a minimum risk level of 3.0 for the total HI and 1×10^{-4} for the total ILCR. SWMUs for which a human health baseline risk assessment was performed are identified in Table 11-2.

Ecological Preliminary and Supplemental Risk Evaluation

An EPRE based on guidance from GEPD was performed on each SWMU to determine the potential risk to ecological receptors associated with the maximum concentrations of the identified SRCs. The EPRE compared measured concentrations of detected constituents to conservative ESVs to identify constituents detected at the facility that pose a potential hazard to ecological receptors and that are identified as ECOPCs to ecological receptors. A summary of the results of the EPRE (ECOPCs) is presented in Table 11-2.

A weight-of-evidence approach was used to evaluate each ECOPC identified in the preliminary risk evaluation. In some instances, ECOPCs were evaluated further in a supplemental preliminary risk evaluation (SPRE). The SPRE presented a comparison of more realistic exposure estimates to TRVs based on LOAELs. The exposure estimates were calculated using measured concentrations and more realistic exposure assumptions such as diets, absorption efficiencies, and AUFs. SWMUs for which a SPRE was performed are identified in Table 11-2.

11.2 RECOMMENDATIONS

A weight-of-evidence approach was used with the results from the fate and transport evaluation, HHPRE, and EPRE to determine the recommendation for each SWMU. The recommendations fell into following three categories:

- **No Further Action:** NFA was recommended for a SWMU if: (1) the contaminant levels in soil, groundwater, surface water, and sediment were below the reference background criteria, fate and transport values (GSSLs), or human health or ecological screening criteria or (2) significant uncertainty was evident, indicating minimal potential risk of migration to groundwater and/or surface water bodies and/or to human health and ecological receptors.
- **Additional Investigation (Phase II RFI or additional monitoring):** A Phase II RFI or additional monitoring was recommended if the nature and extent of potential contaminants had not been determined, and further investigation or additional monitoring was required to evaluate the extent or potential migration of contaminants in the future.
- **Corrective Action Plan:** A CAP was recommended if the nature and extent of contamination at a SWMU was determined by the Phase II RFI, there was a potential risk of migration of contaminants to groundwater and/or surface water bodies or a potential risk to human health and ecological receptors, or institutional controls need to be applied to protect the health and safety of humans coming in contact with the site (i.e., inactive EOD areas). Such a site requires a CAP to evaluate appropriate remedial actions to eliminate or minimize these potential risks.

The recommendations for each SWMU are presented in Table 11-3.

Table 11-1. Summary of Site-related Contaminants

| SWMU | Type of Investigation | Site-related Contaminants | | | | |
|-------------------|-----------------------|--|--------------------------------------|---|----------------------------|--|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment |
| 2 | Phase II | 2 VOCs, 14 pest., 1 SVOC, and 6 metals | 1 VOC, 3 pest., 1 SVOC, and 3 metals | 3 VOCs and 3 metals | None | alpha-Chlordane |
| 3 | Phase II | 4 pest., BEHP, As, Cr, and Pb | 2 VOCs, BEHP, 3 pest., Cr, and Cd | 3 VOCs, 3 pest., Ba, Cd, Cr, Pb, and Hg | 1 SVOC, As, Ba, Cr, and Pb | 6 VOCs, As, Ba, Cr, Pb, Hg, and Se |
| 9 | Phase I ^a | As, Cr, and Ag | NC | NC | NP | NP |
| 10 | Phase II | As, Ba, Cr, and Pb | NC ^b | None | Cd, Cr, and Hg | As, Ba, and Pb |
| 11 | Phase II | As, Ba, Cr, Pb, and Ag | NC ^b | None | NP | NP |
| 12A | Phase II | 3 SVOCs, 4 exp., and 16 metals | Al, As, Ba, Cr, Fe, Pb, and V | BEHP, 1 exp., and 8 metals | RDX, Pb, Mn, and Hg | 1 SVOC, 1 exp., and 9 metals |
| 14 | Phase I | 2 VOCs, BEHP, and Hg | 5 VOCs, Cr, and Hg | 1 VOC, Pb, and Hg | NP | NP |
| 17 | Phase II | 1 VOC | 3 VOCs | 3 VOCs and Pb | None | None |
| 18 | Phase II | 1 VOC, Pb, and Hg | 5 VOCs, 2 SVOCs, Ba, Cr, Pb, and Hg | 9 VOCs, Ba, Cd, and Pb | 1 SVOC and Ba | (6 VOCs, 4 SVOCs, As, Ba, Cd, Cr, Pb, Hg, Se, and Ag) ^c |
| 19 | Phase I | 4 VOCs, 7 pest., and 5 metals | 6 VOCs, 9 pest., and 5 metals | BEHP, 7 pest., and 3 metals | NP | NP |
| 24B | Phase I | 1 VOC, 10 SVOCs, and 6 metals | 2 VOCs | 1 VOC, 11 SVOCs, and Hg | NP | NP |
| 27A (Bldg. 1339A) | Phase I | None | 2 VOCs and 3 SVOCs | 2 VOCs and BEHP | NP | NP |
| 27A (Bldg. 1339B) | Phase I | BEHP and Pb | 2 VOCs | 1 VOC | NP | NP |
| 27A (Bldg. 1322) | Phase I | 3 VOCs and Pb | 3 VOCs | Acetone | NP | NP |
| 27B | Phase I | None | 1 VOC | ND | NP | NP |
| 27C | Phase I | 1 VOC | 2 VOCs and 1 SVOC | 4 VOCs | NP | NP |
| 27D | Phase I | 3 VOCs | 1 VOC | None | NP | NP |
| 27E (Bldg. 1628) | Phase I | None | 1 VOC | None | NP | NP |

Note: Footnotes appear on page 11-5.

Table 11-1. Summary of Site-related Contaminants (continued)

| SWMU | Type of Investigation | Site-related Contaminants | | | | |
|------------------------|-----------------------|---------------------------|------------------------------|---------------------|-----------------|-----------------|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment |
| 27E (Bldg. 1720) | Phase I | NC | 2 VOCs and BEHP | 1 SVOC | NP | NP |
| 27F (NW Bldg. 1340) | Phase I | NC | 3 VOCs and Pb | 10 VOCs and 4 SVOCs | NP | NP |
| 27F (NE Bldg. 1340) | Phase I | 3 VOCs | 8 VOCs and 4 SVOCs | None | NP | NP |
| 27G | Phase I | NC | 3 VOCs | 1 SVOC | NP | NP |
| 27H (Bldg. 1071) | Phase I | NC | 2 VOCs, 11 SVOCs, Pb, and Hg | 1 VOC and 9 SVOCs | NP | NP |
| 27H (Bldg. 1056) | Phase I | NC | 1 VOC, 1 SVOC, Cd, and Pb | 2 VOCs and 4 SVOCs | NP ^d | NP ^d |
| 27I (Block 9900) | Phase I | NC | 1 VOC and Pb | None | NC | 1 VOC and Pb |
| 27I (Block 10300) | Phase I | NC | None | None | Pb | None |
| 27J (Bldg. 10535) | Phase I | None | None | 1 VOC and 1 SVOC | NP | NP |
| 27J (Bldg. 10531) | Phase I | 1 VOC and 1 SVOC | NC | 2 SVOCs | NP | NP |
| 27K | Phase I | NC | 4 VOCs | 1 VOC | NP | NP |
| 27L (Block 10200) | Phase I | None | 1 VOC and 1 SVOC | 8 VOCs and 2 SVOCs | Acetone | None |
| 27M (Block 10100) | Phase I | 1 VOC and Pb | 2 SVOCs and Pb | 1 VOC | NC | Pb |
| 27N (Block 9800) | Phase I | NC | 2 SVOCs and Pb | None | NC | 5 SVOCs |
| 27O (Block 9700) | Phase I | Pb | None | 1 SVOC | 1 VOC | Pb |
| 27P (Block 9500) | Phase I | 1 VOC and 1 SVOC | 1 VOC, 6 SVOCs, and Pb | None | NC | 1 VOC and Pb |

Note: Footnotes appear on page 11-5.

Table 11-1. Summary of Site-related Contaminants (continued)

| SWMU | Type of Investigation | Site-related Contaminants | | | |
|---------------------|-----------------------|-------------------------------------|---------------------------|---------------------------------|---|
| | | Surface Soil | Subsurface Soil | Groundwater | Surface Water |
| 27Q (Block 9400) | Phase I | Pb | None | None | NC |
| 27R | Phase I | None | 1 VOC and 2 SVOCs | None | NP |
| 27S | Phase I | NC | 6 VOCs | None | NP |
| 27T | Phase I | 4 SVOCs | None | 1 VOC and 1 SVOC | NC |
| 27U | Phase I | 1 VOC and Pb | 2 VOCs and Pb | 4 VOCs | NP |
| 27V | Phase I | 1 VOC and Pb | 1 VOC and Pb | None | NP |
| 29 | Phase II | 8 VOCs and Ag | 16 VOCs and 14 SVOCs | 3 VOCs, 3 SVOCs, As, Ba, and Cr | NP |
| 31 | Phase II and IRA | None | 6 VOCs and 17 SVOCs | 4 VOCs | NP |
| 32 | Phase II | 2 VOCs, Ba, Cd, Cr, Pb, and Hg | 2 VOCs, Pb, and Hg | 4 VOCs and 2 SVOCs | NP |
| 34 | Phase II | 4 VOCs, 2 SVOCs, Ba, Cd, Pb, and Hg | 1 VOC, Ba, Cd, Cr, and Pb | 3 VOCs | NP |
| 37 | Phase I | 1 VOC and Hg | 2 VOCs and Hg | 4 VOCs | (4 VOCs, Ba, Cd, Cr, Pb, Hg, and Se) ^f |

^aPhase II RFI was not required at this time. The Phase II RFI will be conducted upon closure of the Red Cloud Range, Hotel Area.

^bPer the GEPD-approved SAP, subsurface soil was not collected because subsurface soil sampling in an EOD area requires approval by the Secretary of the Army.

^cResults from sediment within the NGTC Equalization Basin.

^dSediment was collected; however, the OWS does not discharge to the drainage ditch.

BEHP = Bis(2-ethylhexyl)phthalate.

NA = Not applicable.

NC = Not collected based on field screening results or because no medium (i.e., surface water) was available during the RFI.

ND = Not detected.

NP = No pathway exists.

Table 11-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs

| SWMU | CMCOPCs | | | HHCOPCs | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|-------------------|--|--|---------------------|-----------------|------------------------|------------------------|-----------------|--------------------------|-----------------------------|--------------------|------------------------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 2 | 2 pest., Ar, Ca, and Hg | None | As and Cr | None | None | None | None | 4,4'-DDE, Cd, Cr, and Pb | 1 VOC, Pb, and Hg | None | None | HHBRA and SPRE |
| 3 | None | As | As | None | 1 pest. and Hg | 1 SVOC, As, Cr, and Pb | As | Pb and Cr | 2 pest., Ba, Cd, Pb, and Hg | 1 SVOC, Ba, and Pb | 2 VOCs, As, Ba, and Se | HHBRA and SPRE |
| 9 ^a | NA ^a | NA ^a | NA ^a | NC ^a | NC ^a | NP | NP | NA ^a | NC ^a | NP | NP | |
| 10 | NA ^b | NA ^b | NA ^b | NC | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | |
| 11 | NA ^b | NA ^b | NA ^b | NC | NA ^b | NA ^b | NA ^b | NA ^b | NA ^b | NP | NP | |
| 12A | Ar, Cd, Cr, Pb, Ag, 1 SVOC, and 2 exp. | None | As and Pb | As | BEHP | Hg | None | 1 SVOC, Cd, Cr, and Pb | BEHP | Pb and Hg | Ba | HHBRA and SPRE |
| 14 | 1 VOC | NA | None | None | None | NP | NP | None | Pb, Hg, and 1 VOC | NP | NP | |
| 17 | None | None | None | None | 1 VOC | None | None | None | 1 VOC and Pb | None | None | HHBRA |
| 18 | Cr and Hg | (1 VOC, 1 SVOC, Ar, Ba, Cd, Cr, Hg, and Se) ^f | None | None | 3 VOCs and Pb | 1 SVOC | As | Pb | 4 VOCs, Ba, and Pb | Ba and BEHP | None | HHBRA and SPRE |
| 19 | 2 pest. | NA | None | None | BEHP, 2 pest., and As | NP | NP | Cd, Pb, and 1 pest. | BEHP, 5 pest., Ba, and Hg | NP | NP | HHBRA and SPRE |
| 24B | 1 VOC, 3 SVOCs, and Pb | NA | 4 SVOCs, As, and Pb | None | 1 VOC, 9 SVOCs, and Hg | NP | NP | NP | Hg and 9 SVOCs | NP | NP | |
| 27A (Bldg. 1339A) | None | NA | None | None | BEHP | NP | NP | None | 1 VOC and BEHP | NP | NP | |
| 27A (Bldg. 1339B) | None | NA | None | None | Benzene | NP | NP | Pb | Xylenes | NP | NP | |
| 27A (Bldg. 1322) | None | NA | None | None | Acetone | NA | NA | Pb | None | NP | NP | HHBRA |

Note: Footnotes appear on page 11-8.

Table 11-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs (continued)

| SWMU | CMCOPCs | | | HHCOPCs | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|------------------------|---------|----------|--------------|-----------------|-----------------------|---------------|----------|--------------|-----------------------|---------------|----------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 27B | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27C | None | NP | None | None | None | NP | NP | None | 2 VOCs | NP | NP | |
| 27D | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27E (Bldg. 1628) | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27E (Bldg. 1720) | None | NP | NA | None | None | NP | NP | NA | 1 SVOC | NP | NP | |
| 27F (NW Bldg. 1340) | None | NP | NA | None | 4 VOCs and 4 SVOCs | NP | NP | NA | 2 VOCs and 4 SVOCs | NP | NP | |
| 27F (NE Bldg. 1340) | 1 VOC | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27G | None | NP | NA | None | 1 SVOC | NP | NP | NA | None | NP | NP | HHBRA |
| 27H (Bldg. 1071) | 2 SVOCs | NP | NC | 1 SVOC | 1 VOC and 7 SVOCs | NP | NP | NC | 8 SVOCs | NP | NP | |
| 27H (Bldg. 1056) | None | NP | NC | None | 3 SVOCs | NP | NP | NC | 2 SVOCs | NP | NP | |
| 27I (Block 9900) | None | None | NC | None | None | NC | None | NC | None | NC | Pb | |
| 27I (Block 10300) | None | NA | NC | None | None | Pb | None | NC | None | Pb | None | |
| 27J (Bldg. 10535) | None | NP | None | None | None | NP | NP | None | 1 VOC and 1 SVOC | NP | NP | |
| 27J (Bldg. 10531) | None | NP | None | NC | 1 SVOC | NP | NP | None | 2 SVOCs | NP | NP | |
| 27K | None | NP | NA | None | None | NP | NP | NA | None | NP | NP | |
| 27L (Block 10200) | None | None | None | None | 4 VOCs and 2 SVOCs | Acetone | None | None | 2 VOCs and 1 SVOC | None | None | |
| 27M (Block 10100) | 1 VOC | None | None | None | 1 VOC | NC | None | Pb | None | NC | Pb | |
| 27N (Block 9800) | None | None | NA | None | None | NC | 1 SVOC | NA | None | NC | None | HHBRA |

Note: Footnotes appear on page 11-8.

Table 11-2. Summary of CMCOPCs, HHCOPCs, and ECOPCs (continued)

| SWMU | CMCOPCs | | HHCOPCs | | | | | ECOPCs | | | | HHBRA or SPRE Performed? |
|---------------------|------------------|---------------------------|--------------|-----------------|------------------------|---------------|----------|----------------|------------------------|---------------|----------|--------------------------|
| | Soil | Sediment | Surface Soil | Subsurface Soil | Groundwater | Surface Water | Sediment | Surface Soil | Groundwater | Surface Water | Sediment | |
| 27O (Block 9700) | None | None | None | NA | None | None | None | Pb | 1 SVOC | None | Pb | |
| 27P (Block 9500) | None | None | None | None | None | NC | None | None | None | NC | Pb | |
| 27Q (Block 9400) | None | None | None | NA | None | NC | None | Pb | None | NC | Pb | |
| 27R | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 27S | None | NP | NA | None | None | NP | NP | NA | None | NP | NP | |
| 27T | None | Cd | 1 SVOC | None | None | NA | 4 SVOCs | None | 1 SVOC | NA | Cd | HHBRA |
| 27U | None | NP | None | None | Benzene | NP | NP | Pb | None | NP | NP | |
| 27V | None | NP | None | None | None | NP | NP | None | None | NP | NP | |
| 29 | 7 VOCs | NP | None | None | 1 VOC, 2 SVOCs, and As | NP | NP | None | 1 VOC, 2 SVOCs, and Ba | NP | NP | HHBRA |
| 31 | 1 VOC and 1 SVOC | NP | None | None | Acetone | NP | NP | None | Xylenes | NP | NP | HHBRA |
| 32 | 1 VOC | NP | None | None | Acetone | NP | NP | Cd, Pb, and Cr | 1 VOC and 1 SVOC | NP | NP | HHBRA |
| 34 | 2 VOCs | NP | None | None | Acetone | NP | NP | Cd and Pb | 1 VOC | NP | NP | HHBRA |
| 37 | 1 VOC | 1 VOC ^b and Cd | None | None | Benzene | NP | NP | None | Xylenes | NP | NA | |

^aPhase II RFI was not required at this time. The Phase II RFI will be conducted upon closure of the Red Cloud Range, Hotel Area.

^bWith the concurrence of GEPA, fate and transport analysis and human health and ecological preliminary risk assessments were deemed unnecessary. SRCs were determined solely on comparison to background criteria (see Table 11-1).

^cResults from sediment within the NGTC Equalization Basin.

BEHP = Bis(2-ethylhexyl)phthalate.

HHBRA = Human health baseline risk assessment.

NA = Not applicable.

NC = Sample not collected based on field screening results or because no medium (i.e., surface water) was available during the RFI.

NP = No pathway exists.

SPRE = Supplemental Preliminary Risk Evaluation.

Table 11-3. SWMU-specific Recommendations

| SWMU | Recommendation |
|---------------------------|---|
| 2 | CAP |
| 3 | CAP |
| 9 | CAP |
| 10 | CAP |
| 11 | CAP |
| 12A | Long-term compliance monitoring and CAP |
| 14 | NFA |
| 17 | NFA |
| 18 | Long-term monitoring and CAP |
| 19 | NFA |
| 24B | Phase II RFI |
| 27A (Building 1339A) | NFA |
| 27A (Building 1339B) | NFA |
| 27A (Building 1322) | NFA |
| 27B | NFA |
| 27C | NFA |
| 27D | NFA |
| 27E (Building 1628) | NFA |
| 27E (Building 1720) | NFA |
| 27F (NW Building 1340) | Phase II RFI |
| 27F (NE Building 1340) | NFA |
| 27G | NFA |
| 27H (Building 1071) | Phase II RFI |

| SWMU | Recommendation |
|-------------------------|----------------|
| 27H (Building 1056) | Phase II RFI |
| 27I (Block 9900) | NFA |
| 27I (Block 10300) | NFA |
| 27J (Building 10535) | NFA |
| 27J (Building 10531) | Phase II RFI |
| 27K | NFA |
| 27L (Block 10200) | Phase II RFI |
| 27M (Block 10100) | NFA |
| 27N (Block 9800) | NFA |
| 27O (Block 9700) | NFA |
| 27P (Block 9500) | NFA |
| 27Q (Block 9400) | NFA |
| 27R | NFA |
| 27S | NFA |
| 27T | Phase II RFI |
| 27U | NFA |
| 27V | NFA |
| 29 | CAP |
| 31 | NFA |
| 34 | NFA |
| 32 | NFA |
| 37 | NFA |

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